Corrections

BIOCHEMISTRY. For the article “Genetic characterization of glucose transporter function in Leishmania mexicana,” by Richard J. S. Burchmore, Dayana Rodriguez-Contreras, Kathleen McBride, Michael P. Barrett, Govind Modi, David Sacks, and Scott M. Landfear, which appeared in issue 7, April 1, 2003, of Proc. Natl. Acad. Sci. USA (100, 3901–3906; First Published March 21, 2003; 10.1073/pnas.0630165100), the authors request that Patrick Merkel, Physiologisch-chemisches Institut, University of Tübingen, Hoppe-Seyler Strasse 4, 72076 Tübingen, Germany, be added to the list of authors between Kathleen McBride and Michael P. Barrett. The revised author line appears below.

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BIOPHYSICS. For the article “Single pilus motor forces exceed 100 pN,” by Berenike Maier, Laura Potter, Magdalene So, Hank S. Seifert, and Michael P. Sheetz, which appeared in issue 25, December 10, 2002, of Proc. Natl. Acad. Sci. USA (99, 16012–16017; First Published November 22, 2002; 10.1073/pnas.242523299), the authors request that Cynthia D. Long, Department of Microbiology-Immunology, Northwestern Medical School, Chicago, IL 60611, be added to the list of authors between Magdalene So and Hank S. Seifert. Dr. Long’s present address is Molecular Diagnostics, Abbott Laboratories, 100 Abbott Park Road, Abbott Park, IL 60064. The revised author line appears below.

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Both insect and mammalian life cycle stages of *Leishmania mexicana* take up glucose and express all three isoforms encoded by the *LmGT* glucose transporter gene family. To evaluate glucose transporter function in intact parasites, a null mutant line has been created by targeted disruption of the *LmGT* locus that encompasses the *LmGT1*, *LmGT2*, and *LmGT3* genes. This Δlmgt null mutant exhibited no detectable glucose transport activity. The growth rate of the Δlmgt knockout in the promastigote stage was reduced to a rate comparable with that of WT cells grown in the absence of glucose. Δlmgt cells also exhibited dramatically reduced infectivity to macrophages, demonstrating that expression of *LmGT* isoforms is essential for viability of amastigotes. Furthermore, WT *L. mexicana* were not able to grow as axenic culture form amastigotes if glucose was withdrawn from the medium, implying that glucose is an essential nutrient in this life cycle stage. Expression of either *LmGT2* or *LmGT3*, but not of *LmGT1*, in Δlmgt null mutants significantly restored growth as promastigotes, but only *LmGT3* expression substantially rescued amastigote growth in macrophages. Subcellular localization of the three isoforms was investigated in Δlmgt cells expressing individual *LmGT* isoforms. Using anti-*LmGT* antiserum and GFP-tagged *LmGT* fusion proteins, *LmGT2* and *LmGT3* were localized to the cell body, whereas *LmGT1* was localized specifically to the flagellum. These results establish that each glucose transporter isoform has distinct biological functions in the parasite.

The *Leishmania* species include human pathogens whose digenetic life cycle involves transmission of an extracellular flagellated promastigote from the gut of a hematophagous sandfly vector to a mammalian host, where the parasite multiplies within macrophage phagolysosomes as a nonmotile amastigote form. The two life cycle stages are exposed to highly contrasting nutritional environments. Sandflies feed principally on sugar-rich plant fluids and thereby subject promastigotes to high levels of sugars (1). When a sandfly ingests a bloodmeal containing *Leishmania* amastigotes, the parasites transform into promastigotes and multiply in the midgut. As the infection progresses, promastigotes colonize the insect foregut, where a subpopulation differentiates to metacyclic promastigotes that are highly motile and infective to mammals (2). Ingestion of a sugar-rich meal by the sandfly is required for successful transmission by bite (3), possibly because material released from the sugar crop (4) provides the major nutrient source in the cuticle-lined foregut (5).

On transmission to mammalian tissues, metacyclic promastigotes can survive phagocytosis by macrophages and multiply as amastigotes in a phagolysosome. This compartment is very acidic and will thus present special challenges to parasite membrane transporters (6). The phagolysosome is rich in the products of macromolecular breakdown but may not accumulate high levels of free glucose. *Leishmania* parasites down-regulate glucose transport activity (7) and up-regulate catabolism of fatty acids (8) as they transform from promastigote to amastigote, suggesting that uptake systems are developmentally regulated to maximize nutrient availability despite changing environments.

A gene family (*LmGT*) from *Leishmania mexicana* that encodes three distinct glucose transporter isoforms, *LmGT1*, *LmGT2*, and *LmGT3*, has been cloned recently (9). The *LmGT* glucose transporter isoforms are members of the major facilitator superfamily, a group of transport proteins (10) that mediate facilitated and active transport of various nutrients in both prokaryotes and eukaryotes. The amino acid identity among the *LmGT* isoforms is high, although discrete patches of divergence are clustered at the N and C termini and at several internal locations. mRNA derived from each of the *LmGT* genes is expressed in both promastigotes and amastigotes, although *LmGT2* mRNA is strongly up-regulated in promastigotes (9). The simultaneous presence of multiple glucose transporters might be explained by postulating discrete functions for each permease. In mammals, multiple hexose transporters exhibit different substrate specificities and have specific tissue and subcellular localizations commensurate with their biological roles. In *Leishmania*, the changing environments encountered by the parasite during its life cycle may require differential expression of glucose transporter isoforms. Moreover, the glycolytic enzymes responsible for glucose metabolism are sequestered in peroxisome-like organelles called glycosomes (11), implying a potential role for glucose translocation across the glycosomal membrane. A *Leishmania enrietti* glucose transporter isoform is localized to the flagellar membrane (12) for reasons that appear enigmatic because the glycosomes, and hence glycolysis, are probably restricted to the cell body. However, transporters and transporter-like proteins may have roles other than simply acquiring substrates for metabolism. For example, GLUT1, the archetypal mammalian glucose transporter, has a glucose-sensing function (13), and in yeast a number of transporter-like molecules have been shown to play roles in nutrient sensing (14). In the current study, we have investigated the biological function of the *L. mexicana* glucose transporters by generating a null mutant of the *LmGT1*, *LmGT2*, *LmGT3* gene cluster by targeted gene replacement (15). We have analyzed phenotypes of this glucose transporter “knockout” line, Δlmgt, and of this null mutant complemented with each of the individual glucose transporter genes. The results reveal distinct roles for each transporter isoform and suggest important functions for these permeases in both promastigotes and amastigotes.

### Materials and Methods

**Parasite Culture.** *L. mexicana* WT MNYC/BC/82/2379 promastigotes were cultured at 26°C in MEM designated HOMEM (16), RPMI medium 1640, or DMEM-*Leishmania* [DME-L (17), supplemented with 10% heat-inactivated FCS (iFCS). *L. mexicana* parasites down-regulate glucose transport activity (7) and up-regulate catabolism of fatty acids (8) as they transform from promastigote to amastigote, suggesting that uptake systems are developmentally regulated to maximize nutrient availability despite changing environments.

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form (CF) amastigotes (9) were grown at 32.5°C in either Schneider’s Drosophila medium or in DME-L containing 30 mM Mes buffer instead of Hepes. Both of these media were supplemented with 20% iFCS and were adjusted to pH 5.5.

Infection of Cultured Peritoneal Macrophages. Peritoneal exudate macrophages were isolated from female BALB/c mice by peritoneal lavage, seeded onto chamber slides at a density of 2 × 10^6 per ml, and incubated overnight in RPMI at 32°C, 5% CO₂. Attached macrophages were washed with fresh RPMI, then incubated for 6 h with stationary phase promastigotes at 2 × 10^6 per ml in RPMI at 32°C, 5% CO₂. Residual free parasites were removed by three washes with RPMI, and slides were incubated as above in RPMI until fixation.Slides were fixed with methanol, stained with Giemsa, and examined under the microscope to detect intracellular parasites.

Generation of the Δlmgt Line. LmGT alleles were replaced sequentially with puromycin acetyltransferase (PAC) and streptothricin selectable marker (SAT) genes, encoding resistance markers for the antibiotics puromycin and nourseothricin, respectively. The gene deletion construct for puromycin selection (LmGTKOPAC) was based on the Leishmania expression vector pX63PAC (18). Flanking sequences upstream (1.2 kb) of the LmGT1 ORF and downstream (1.8 kb) of the LmGT3 ORF (upstream and downstream segments, respectively; Fig. 1 A) were amplified by PCR, using primers that incorporated restriction sites suitable for subsequent insertion into the polylinker regions flanking the PAC gene in pX63PAC. The gene deletion construct for nourseothricin selection (LmGTKOSAT) was generated by removing the PAC coding region from LmGTKOPAC and replacing it with a fragment containing the SAT coding region from pCPC-SAT (19). Plasmid DNA for each construct was digested with three restriction enzymes to destroy the plasmid backbone, and the linear gene deletion constructs were gel purified by using QIAEX columns (Qiagen, Valencia, CA).

L. mexicana promastigotes were grown in culture to ~ 8 × 10^6 per ml, washed in cold Zimmerman medium with glucose (ZMG) (20) and resuspended in cold ZMG at 1 × 10^6 per ml. Four hundred-microliter aliquots were electroporated with ~5 µg of linear DNA (0.45 kV, 500 µF by using a Bio-Rad Gene Pulser II apparatus) in 0.2 cm of electrode gap cuvettes, transferred to 5 ml of HOMEM/10% iFCS, and incubated at 26°C. After 24 h, cells were pelleted and resuspended in 5 ml of fresh HOMEM/10% iFCS with puromycin at 10 µg/ml or nourseothricin at 25 µg/ml. When living cells were observed (after ~10 days in a typical experiment), clones were derived by limiting dilution on multiwell plates, without allowing further expansion for substrate saturation curves of LmGT2 and LmGT3, incubations with [14C]D-glucose were performed for 20 s, after pilot studies indicated that uptake was linear for at least this period over the range of glucose concentrations used. For LmGT1, uptake assays were performed with [3H]glucose between 0 and 50 s, and the initial rate data were fitted to a straight line by linear regression.
Infection of Sandflies with Promastigotes. Three- to five-day-old Lutzomyia longipalpis sandflies were fed through a chick-skin membrane on a mixture of heparinized mouse blood containing $1\times 10^6$ procyclic promastigotes per ml, obtained from 1- to 2-day-old logarithmic cultures. Blood-engorged sandflies were separated and maintained at 28°C with 30% sucrose solution. At various times after feeding, the flies were anesthetized with CO$_2$, and their midguts were dissected and examined microscopically for the presence and location of promastigotes. The number of midgut promastigotes was determined by placing individual midguts into a microcentrifuge tube containing 30 μl of PBS, pH 7.4, homogenizing each gut by using a Teflon-coated microtissue grinder, and counting released promastigotes on a hemacytometer.

Preparation of Peptide Antiserum. Synthesis of a peptide (CSSLS-GNRAE) encompassing the COOH-terminal nine amino acids of LmGT1 and LmGT2 and containing a NH$_2$-terminal cysteine residue, coupling to keyhole limpet hemocyanin, and generation of rabbit polyclonal antisera were performed by Alpha Diagnostic International (San Antonio, TX). Crude antiserum was affinity-purified by using the cognate peptide coupled to Affigel 15 (Bio-Rad), following the manufacturer's instructions. Affinity-purified antiserum was used at a 1:50 dilution.

Fluorescence Microscopy. The Δlmgt cells expressing LmGT1 or LmGT2 were processed for immunofluorescence microscopy by using affinity-purified primary antibody and fluorescein isothiocyanate-coupled goat anti-rabbit secondary antibody, as described (22). For expression of proteins consisting of GFP fused to the COOH terminus of LmGT2 or LmGT3, each transporter ORF was subcloned into the pX-GFP expression vector (23). Fluorescence images of Δlmgt cells expressing LmGT2-GFP and LmGT3-GFP were obtained as described (24). Fluorescence and differential interference contrast images were obtained and deconvolved by using the DeltaVision Image Restoration System (Applied Precision Instruments, Issaquah, WA).

Results

Targeted Replacement of the LmGT1, LmGT2, LmGT3 Gene Cluster. To determine the biological function(s) of the L. mexicana glucose transporters LmGT1, LmGT2, and LmGT3, knockout parasites were created by double-targeted gene replacement of the LmGT locus (Fig. 1A). A clonal line representing the doubly disrupted null mutant of the LmGT locus, designated Δlmgt, was examined by Southern blot analysis to demonstrate that the correct homologous integration had occurred. Genomic DNA from the Δlmgt line (−/−, Fig. 1B) did not hybridize to a probe from the LmGT2 ORF that hybridizes to all three LmGT genes (9), whereas DNA from WT L. mexicana or from the heterozygous knockout line (+/+ and +/−, respectively, Fig. 1B) did hybridize to this probe, demonstrating that all three LmGT ORFs had been eliminated from the Δlmgt line. Probing the Southern blot with the paraflagellar rod gene, PFR2 (25), revealed hybridization to DNA from all three cell lines (Fig. 1C). Furthermore, hybridization with the upstream probe demonstrated that a 14-kb EcoRI fragment present in DNA from WT parasites was reduced to a predicted band of 2 kb, whereas hybridization with the downstream probe revealed that a 10-kb SmalI band in WT DNA was reduced in size to a predicted band of 7 kb (Fig. 1A and data not shown). Together, these results confirm that the correct homologous integrations had occurred at the 5′ and 3′ sides of the LmGT locus for both rounds of homologous gene replacement.

Transport Properties of Δlmgt Knockout Promastigotes. To determine the transport phenotype of the Δlmgt null mutants, uptake assays were performed on both WT and Δlmgt cells by using the glucose analog $[^{3}H]$2-deoxy-D-glucose (Fig. 2A). WT cells exhibited robust uptake, whereas the Δlmgt cells were incapable of taking up the glucose analog. In contrast, both cell lines incorporated $[^{3}H]$adenosine at identical rates (Fig. 2B), indicating that the null mutants were competent for transport of adenosine. These results imply that LmGT1, LmGT2, and LmGT3 are probably the only functional glucose transporter genes in L. mexicana.

Growth Properties of Δlmgt Promastigotes. To determine the role of the LmGT transporters in parasite growth, we examined the growth of WT and Δlmgt promastigotes in DME-L medium (17) with and without 25 mM glucose and with and without 5 mM proline, an amino acid that can be used as an alternative energy source by Leishmania promastigotes (26). In medium containing both glucose and proline, Δlmgt parasites grew less rapidly and to a lower cell density than WT cells, doubling from $1 \times 10^7$ to $2 \times 10^7$ cells per ml in $\sim$60 h compared with $\sim$20 h for WT cells and attaining a density of $\sim$2.5 $\times 10^7$ cells per ml compared with $\sim$5 $\times 10^7$ cells per ml for WT cells (compare WT(+, +) to Δlmgt(−, +, +), Fig. 3). The growth rate of Δlmgt cells was unaffected by withdrawal of glucose from the culture medium (compare Δlmgt(+, +, +) to Δlmgt(−, −, +), Fig. 3B). In contrast, WT parasites grew more slowly in medium without glucose compared with medium containing glucose (compare WT(+, +, +) to WT(−, −, +), Fig. 3A). Notably, WT cells in glucose-deficient medium had the same growth rate as Δlmgt cells growing in either the presence or absence of glucose. Thus, either removal of glucose from the medium [WT(−, +, +), Fig. 3A] or elimination of glucose transport capacity [Δlmgt(+, +, +), Fig. 3B] slowed parasite growth to the same extent compared with WT cells grown in the presence of glucose.
summary, glucose uptake promotes growth of WT promastigotes but is not essential for survival.

Similarly, we have determined that removal of proline from the medium further slows growth of both WT and Δlmgt parasites in either the presence or absence of glucose (compare +P and −P curves in Fig. 3). Hence, consistent with previous observations by others (27), both glucose and proline can support growth of promastigotes and, in the absence of glucose uptake, proline can function as a major source of energy. Although Δlmgt cells grow very slowly in the absence of both glucose and proline, they are still viable and probably use other amino acids as energy sources (28), albeit inefficiently.

Ability of Individual LmGT Isoforms to Promote Growth of Promastigotes. To assess the role of each LmGT isoform in supporting growth of promastigotes, we complemented the Δlmgt null mutant individually with the LmGT1, LmGT2, or LmGT3 ORF and monitored growth in RPMI containing 25 mM glucose. A region of each gene containing the ORF was subcloned into the Leishmania expression vector pX63NEO (29) and transfected into the Δlmgt line to generate the Δlmgt[pGT1], Δlmgt[pGT2], and Δlmgt[pGT3] lines, each of which was complemented with the gene for one isoform. Growth curves (Fig. 4) revealed that the LmGT2 gene was able to complement growth of the null mutant to the same level as WT parasites. LmGT3 was able to partially restore the growth rate, but LmGT1 provided very little increase in growth over the level of the Δlmgt null mutant.

Growth of Δlmgt Promastigotes in Sandflies. Glucose is thought to be an important nutrient for the promastigote in the sandfly and has been proposed to be essential for development of infectious parasites in the insect (3). To determine whether the glucose uptake is important for parasite growth in the sandfly, we infected L. longipalpis with both WT and Δlmgt knockout parasites and monitored the number of parasites in the insect midgut as a function of time (Fig. 5). WT parasites established initial infections with ~10^3 parasites per midgut, which then dropped to ~10^3 parasites during excretion of the blood meal on day 5, followed by repopulation of the midgut, including the anterior midgut, by day 9. In contrast, Δlmgt parasites grew significantly less efficiently in the midgut, especially after excretion of the bloodmeal, and the repopulation occurred later (day 15) and to a lesser extent than for WT parasites.

Development of Metacyclic Promastigotes in WT and Δlmgt Parasites. To determine whether deletion of the glucose transporter genes affects transformation of L. mexicana promastigotes into infectious metacyclic forms whose abundance is increased in stationary-phase promastigote cultures (2) and in the anterior of the sandfly during a natural infection (30), we monitored metacyclogenesis in stationary-phase cultures by Percoll density gradient enrichment (31) followed by morphological quantitation (32). WT parasites produced 7.8% and 10.4% metacyclic forms in two cultures, whereas Δlmgt parasites produced 2.5% metacyclic forms in both cases. Furthermore the meta 1 transcript, which is up-regulated in metacyclic parasites (33), was induced 3.6-fold in WT stationary-phase compared with mid-logarithmic phase parasites, whereas the transcript was induced 1.7-fold in stationary-phase Δlmgt cells. By these criteria, metacyclogenesis appears to be impaired but not abrogated in glucose transporter null mutants.

Growth of the Δlmgt Null Mutant and Individual Complemented Lines as Amastigotes. To determine whether glucose transporters play a significant role in the amastigote stage of the life cycle, we infected macrophages with WT and Δlmgt null mutant cells (Fig. 6). Primary cultures of mouse peritoneal exudate cells, which are comprised chiefly of macrophages (34), were exposed to infection by stationary-phase WT and Δlmgt cells. After 6 h of exposure similar numbers of intracellular parasites were observed (~10–15 parasites per 100 macrophages), indicating that each cell line possessed a similar capacity to infect macrophages (data not shown). However, 48 h after infection, only 1% of macrophages examined harbored Δlmgt amastigotes, whereas 26% of macrophages harbored WT parasites. Six days after infection, 1% of macrophages were infected with Δlmgt cells, whereas 35% were infected with WT parasites, and 182 WT parasites were observed per 100 macrophages examined, compared with two Δlmgt parasites per 100 macrophages. These

![Fig. 4](image-url) Growth in RPMI medium of promastigotes of WT, Δlmgt null mutant, and Δlmgt null mutants complemented with each of the LmGT genes (Δlmgt[pGT1], Δlmgt[pGT2], Δlmgt[pGT3]).

![Fig. 5](image-url) Infection of L. longipalpis sandflies by WT (filled circles) and Δlmgt (open circles) promastigotes. At each time point after infection, midguts were dissected from 10–12 sandflies, and parasites were quantitated. Similar results were obtained from four independent experiments.

![Fig. 6](image-url) Growth of WT, Δlmgt, Δlmgt[pGT1], Δlmgt[pGT2], and Δlmgt[pGT3] lines in murine peritoneal macrophages. Filled bars represent percent of infected macrophages, and open bars represent parasites per 100 macrophages for the same fields. Primary peritoneal macrophages were infected with stationary phase promastigotes, and the number of intracellular amastigotes (n = 3, average ± SD) was quantitated 6 days after infection.
of the corresponding proteins to be expressed in amastigotes. Efficient restoration of growth of amastigotes was not due to failure of the parasites to grow (Fig. 7B). Consequently, L. mexicana apparently require glucose to survive as amastigotes, and the nonviability of the Δlmgt null mutants, either inside macrophages (Fig. 6) or as CF amastigotes (Fig. 7A), is likely due to their failure to take up this essential nutrient.

Characterization of Glucose Transport by Each Isoform. To define the glucose-transport characteristics of each LmGT isoform, transport assays were performed with the ΔlmgtpGT1, ΔlmgtpGT2, and ΔlmgtpGT3 lines. Substrate saturation curves for uptake of [14C]glucose were generated for each line and revealed apparent $K_m$ values for D-glucose of 1.22 ± 0.22 mM, 109 ± 28 μM, and 208 ± 40 μM for LmGT1, LmGT2, and LmGT3, respectively ($n = 3$). Thus LmGT1 is a lower-affinity glucose transporter than LmGT2 or LmGT3.

Subcellular Localization of the LmGT1, LmGT2, and LmGT3 Transporters. To determine whether the isoforms might also differ in subcellular localization, we defined the distribution of each protein by deconvolution fluorescence microscopy. In the first approach, an antiserum was raised against the peptide CSSLS-GNRAE that encompasses the last nine amino acids of LmGT1 and LmGT2, which differ from those of LmGT3 (9). A negative control sample of Δlmgt cells showed only diffuse staining over the surface of the cells using this antiserum. The immunofluorescence images (Fig. 8A and B) are published as supporting information on the PNAS web site, www.pnas.org. Clearly indicate that LmGT1 is localized primarily in the flagellum, whereas LmGT2 is targeted to the pellicular plasma membrane (12) that surrounds the cell body.

To define the location of LmGT3 and to provide another reagent to confirm the location of LmGT2, we also prepared fusion constructs in which GFP (37) was fused to the COOH terminus of each of these two isoforms. Deconvolution fluorescence images of cells expressing LmGT2-GFP (Fig. 8C) and LmGT3-GFP (Fig. 8D) revealed that both of these proteins are located largely on the pellicular plasma membrane that surrounds the cell body and found only at very low levels on the flagellum. Furthermore, studies on the uptake of [3H]2-deoxy-D-glucose by using the ΔlmgtpGT2-GFP and ΔlmgtpGT3-GFP lines confirmed that these fusion proteins are functional glucose transporters (data not shown) and thus are not significantly disrupted by fusion to GFP.

Discussion

In many organisms, including kinetoplastid parasites, multiple glucose transporters are coexpressed, and functional characterization of individual glucose transporter isoforms has depended on separate expression in heterologous systems such as Xenopus oocytes. However, a “gold standard” for assessing the biological function of each gene in an organism is to generate a “knockout” line and to examine the phenotype of this null mutant. To assess the phenotype of L. mexicana without glucose transporters and to functionally evaluate each of the three LmGT isoforms in a null background, we have generated a LmGT knockout line. Δlmgtp promastigotes were unable to take up radiolabeled glucose, confirming that members of the LmGT family are responsible for glucose uptake in promastigotes. The reduced growth rate, but continued viability, of Δlmgtp promastigotes supports existing biochemical evidence that glucose is a major but not exclusive source of metabolic energy for Leishmania promastigotes. These results are further confirmed by the observation that Δlmgtp promastigotes grow poorly in the sandfly L. longipalpis and thus establish an important, albeit not essential, role for the parasite glucose transporters in the infection of the insect vector.

Functional characterization reveals important differences between the three LmGT isoforms in relation to glucose transport and ability to support parasite growth in both promastigotes and amastigotes. Expression of individual LmGT genes in Δlmgtp promastigotes revealed that expression of LmGT2 is sufficient to completely restore the WT growth phenotype. This result is consistent with the observation that LmGT2 mRNA is strongly up-regulated in WT promastigotes (9), and LmGT2 may be the isoform that is responsible for most of promastigote glucose uptake.

Growth of WT and Δlmgt Parasites as CF Amastigotes. To determine whether amastigotes require glucose uptake for viability, thus potentially explaining the failure of Δlmgt parasitoids to survive inside macrophages, we have grown both WT and Δlmgt null mutant parasites as axenic CF amastigotes present in WT or LmGT3 complemented null mutants. Furthermore, when macrophages were infected with WT L. mexicana expressing each isoform fused to GFP, fluorescence from each isoform was detected in the membranes of intracellular amastigotes (data not shown). Hence the failure of LmGT1 and LmGT2 to efficiently restore growth of amastigotes was not due to failure of the corresponding proteins to be expressed in amastigotes.

Fig. 7. Growth of WT L. mexicana and Δlmgt null mutants as axenic CF amastigotes. (A) WT (circles) and Δlmgt (triangles) parasites were inoculated into Schneider’s medium supplemented with 20% IFCS and adjusted to pH 5.5 followed by incubation at 32.5°C, and aliquots were withdrawn and counted (n = 3, average ± SD) at various times. (B) CF amastigotes of WT L. mexicana, growing in DME-L medium containing 30 mM Mes buffer, pH 5.5, and supplemented with 20% IFCS (CF-DME-L), were pelleted, washed, and inoculated at a density of 1 × 10^6 cells per ml into fresh CF-DME-L constituted with 11 mM glucose and dialyzed IFCS (diamonds), or CF-DME-L deficient in glucose and constituted with dialyzed IFCS (squares).
uptake. Expression of LmGT3 was sufficient to partially restore the growth of Δlmgt, but LmGT1 gave only very limited growth restoration, although both of these transcripts are present constitutively throughout the life cycle. These observations are consistent with the functional characterization of the three LmGT isoforms, because LmGT2 and LmGT3 are glucose transporters with relatively high affinity, whereas LmGT1 by comparison is a lower-affinity glucose transporter.

In addition to the functional differences described above, subcellular localization studies indicate that LmGT1 is a flagellar protein, whereas both LmGT2 and LmGT3 are pellicular plasma membrane isoforms. These results are similar to those previously obtained with the two related glucose transporter isoforms from L. enriettii, in which ISO1 was demonstrated to be a flagellar transporter and ISO2 a pellicular plasma membrane permease. It bears a large NH2-terminal hydrophilic extension that is predicted to have an intracellular orientation and might play a role in restricting cell surface localization, as is the case for the ISO1 glucose transporter in L. enriettii (12).

The distinct subcellular distributions of the different isoforms suggest the possibility of biologically significant functional specializations that may be associated with the divergent targeting of each permease. It is noteworthy that in other organisms, axoneme-containing organelles such as flagella and cilia are often involved in environmental sensing (39). Hence, one possible explanation for the flagellar localization of LmGT1 is that it might function as a glucose sensor. Furthermore, in other organisms such as Saccharomyces cerevisiae (40), Neurospora crassa (41), and humans (13), glucose transporter-like proteins have been shown to function as glucose sensors. Whether LmGT1 functions as a glucose sensor may be an important topic of future investigations.

A significant role for glucose transporters in amastigotes is revealed by the observation that the Δlmgt line was unable to sustain infection in murine peritoneal exudate cells. These results imply that glucose transporter expression is essential for amastigote viability and were unanticipated, given that glucose uptake (7) and catabolism (42) are thought to be down-regulated in amastigotes. Furthermore, the failure of Δlmgt null mutants to grow as CF amastigotes and the inability of WT parasites to survive as CF amastigotes in medium devoid of glucose further imply that glucose is an essential nutrient for amastigotes and that glucose transporter null mutants are not viable as amastigotes for this reason. The ability of LmGT3, but not of LmGT1 or LmGT2, to fully restore WT viability suggests that LmGT3 may subsume an essential role in the parasitophorous vacuole. This environment is not well understood, but free glucose levels in macrophage phagolysosomes may be low. Thus LmGT3 might function in the amastigote to scavenge sparse glucose from the lumen of the parasitophorous vacuole. Regardless of their precise roles in amastigote biochemistry, the apparent requirement of amastigotes for functional glucose transporters raises the possibility that interference with parasite glucose transporter function might be of therapeutic value in L. mexicana infections and that these permeases could be targets for drug development. Although it is clear that glucose is a nonessential nutrient for promastigotes in axenic culture, glucose transport may be critical to Leishmania development in the sandfly host. The diet and digestive physiology of the sandfly is not well understood, but a substantial body of evidence suggests that sugars in the sandfly diet are important in the development of Leishmania promastigotes into infectious forms (reviewed in ref. 43). The significantly reduced numbers of parasites in the anterior midgut of sandflies infected with the Δlmgt null mutants would very likely reduce the competence of these vectors to transmit an infection to the vertebrate host.

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