Promoting oligodendrogenesis and myelin repair using the multiple sclerosis medication glatiramer acetate

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The formation of oligodendrocytes (oligodendrogenesis) and myelination is regulated by several neurotrophic factors. Strategies to increase the level of these trophic molecules may facilitate repair in demyelinating conditions, such as multiple sclerosis (MS). Because leukocytes are a source of neurotrophic factors, and as glatiramer acetate (GA) generates T helper 2 (Th2) lymphocytes that are not known to be harmful, we tested the hypothesis that GA regulates oligodendrogenesis and myelin formation. First, we generated GA-reactive Th2 cells and determined that they produced transcripts for neurotrophic factors, including insulin-like growth factor-1 (IGF-1). The conditioned medium from GA-reactive T cells elevated IGF-1 protein and promoted the formation of oligodendrocyte precursor cells (OPCs) from embryonic brain-derived forebrain cells in culture. We next subjected mice to lysolecithin-induced demyelination of the spinal cord. At 7 days after the insult, the number of OPCs in the demyelinated dorsal column was higher than that in uninjured controls, and was further increased by the daily s.c. injection with GA. Increased OPC generation by GA was associated temporally with the elevation of IGF-1 and brain-derived neurotrophic factor (BDNF) in the spinal cord. Finally, the resultant remyelination at 28 days was higher in mice treated with GA during the first 7 days of injury compared with vehicle controls. These results indicate that GA promotes oligodendrogenesis and remyelination through mechanisms that involve the elevation of growth factors conducive for repair.

Regeneration | Remyelination

Multiple sclerosis (MS) is characterized by inflammation and demyelination in the CNS. The demyelinated lesions can be repaired and, indeed, the extent of remyelination is substantial in some patients (1). There is increasing interest in facilitating remyelination as its benefits extend beyond restoration of nerve impulse conduction to preventing axonal degeneration, since axon and myelin units have dynamic interactions involving survival signaling.

Much has been learned about the process of remyelination (2) and key steps include the proliferation and maturation of oligodendrocyte precursor cells (OPCs) and the appropriate interactions of oligodendrocyte processes with axons to form myelin. A large number of molecules are critically involved in the proliferation, maturation and survival of OPCs, and among these are neurotrophic factors such as platelet-derived growth factor (PDGF), insulin-like growth factor-1 (IGF-1) and brain-derived neurotrophic factor (BDNF) (3–6). Accordingly, strategies that have been used to induce remyelination in animals have included the provision of neurotrophic factors through infusion or gene therapy.

Another means to deliver growth factors to the nervous system takes advantage of the observation that leukocytes are rich sources of neurotrophic factors. Indeed, even the disease-promoting inflammatory cells of perivascular cuffs in MS lesions are immunoreactive for neurotrophic factors such as BDNF (7). However, any approach to use leukocytes to deliver neurotrophic factors for repair must balance their potential detrimental effects in exacerbating the pathology of MS.

Glatiramer acetate (GA), a medication used in relapsing-remitting MS (8), generates GA-reactive T lymphocytes that are of the T helper 2 (Th2) anti-inflammatory bias (9, 10). GA-reactive Th2 cells accumulate in the CNS, where they have been shown to produce not only anti-inflammatory cytokines but also neurotrophic factors (11). Importantly, Th2 cells have not been found to be neurotoxic, unlike the proinflammatory Th1 or Th17 lymphocytes that destroy neurons in tissue culture (12). More recently, GA has also been found to modulate monocytoid cells into those that express anti-inflammatory cytokines (13).

Here, we have tested the hypothesis that T cells exposed to GA elevate neurotrophic factors that are important for oligodendrogenesis in culture. Furthermore, we have used mice with lysolecithin-induced demyelination to address whether treatment with GA would increase OPC numbers and remyelination within the lesioned spinal cord. These studies have relevance to the biology of harnessing the benefits of inflammation to evoke repair.

Results

T Cells Exposed to GA Elevate Their Production of Growth Factors in Vitro. Cells obtained from the lymph nodes of GA-pretreated mice (Fig. S1) were incubated with APCs and GA, resulting in a proliferative response (Fig. S2) that suggested the presence of T cells reactive to GA. Cell conditioned medium from these GA-reactive cells was then examined for the accumulation of IFN-γ, a Th1 cytokine, and for IL-5, a Th2 cytokine. Increasing culture periods with GA resulted in loss of detectable IFN-γ and steady accumulation of IL-5 (Fig. S2), indicating that the GA-reactive T cells were of the Th2 phenotype, as reported by others (9–11). With this confirmation, we examined the capacity of GA-reactive T cells to produce growth factors. From several mouse donors previously treated with GA, lymph node cells were restimulated with GA and RNA was harvested. Fig. 1 shows that increases in transcripts encoding IGF-1 and PDGFaa were evident after 3 days of GA treatment. Transcripts for BDNF was variable, with three cultures elevating this trophic factor in response to GA whereas three did not.

The cell-conditioned medium collected from T cells over 3 days of culture was analyzed for IGF-1 protein content. IGF-1

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protein was significantly elevated in T-cell cultures treated with GA, either collected at first in vitro restimulation with GA (day 3), or after restimulation a week after (day 10) (Fig. S2).

Conditioned Medium from GA-Reactives T Cells Increases the Number of OPCs in Culture. We evaluated whether growth factors produced by GA-reactive T cells (Fig. 1) were sufficient to regulate the formation of OPCs from neural precursor cells. Single cells from the anterior entopeduncular area (AEP) of embryonic day 15 mice were exposed to conditioned medium collected from GA-reactive T cells for 72 h. Fig. 2 and Fig. S3 show that there were significantly more OPCs and Ki67-positive cells when the embryonic brain cells were incubated with medium from GA-reactive T cells, compared with medium from control T cells (not restimulated with GA). These results show that GA-reactive T cells secrete factors conducive for oligodendrogenesis.

GA Treatment of Mice with Lysolécin Injury Increases the Number of OPCs in the Lesioned Spinal Cord. We next investigated whether GA could increase OPC numbers in vivo. To better simulate conditions in patients, we injected mice daily with s.c. GA. Previous studies have shown that significant demyelination (14, 15) and newly generated OPCs (16) could be found by day 7 after lysolecithin administration, so we chose this time point for analysis. Fig. S4 of MBP immunoreactivity shows that demyelination of the dorsal column of the spinal cord was extensive at day 7. In the area of demyelination, a dense accumulation of Iba1+ microglia/macrophages was readily observed. Moreover, OPCs identified by PDGFRα immuno-labeling and as discrete cells could be discerned (Fig. S4). We blindly enumerated the number of OPCs in the dorsal column across 10 sections of the spinal cord, each spaced 100 μm apart, and centered around the epicenter of injury. Fig. 3 shows that in mice subjected to lysolécin injury and treated s.c. with PBS vehicle, the average total number of OPCs in the dorsal column summed across 10 sections of individual mice averaged 257. This represents an increase from uninjured mice (average sum of 141 OPCs across 10 sections). Impressively, daily treatment with s.c. GA for 7 days after lysolécin injury resulted in a further increase of OPC numbers, where an average of 390 OPCs across 10 sections was found per mouse (Fig. 3).

The increase of OPC numbers in the dorsal column induced by GA compared with vehicle occurred not only at the lesion epicenter, but also rostrally or caudally across 300–400 μm of either side (Fig. 3) containing a demyelinating lesion. Overall, these results demonstrate that the daily s.c. injection of GA for 7 days after lysolécin injury results in an elevation of the injury-induced increase of OPCs in the spinal cord and that this increase occurs locally in the dorsal column that contains a demyelinating injury.

Alterations of Cytokine and Growth Factor Levels in Spinal Cords of GA-Treated Mice. We determined whether GA treatment of lysolécin-injured mice led to alterations of inflammatory cytokines or growth factors in vivo. Coronal spinal cord blocks from around the injury site were harvested from mice treated s.c. with GA or vehicle daily for 7 days after lysolécin injury, and

Fig. 1. GA-reactive T cells in tissue culture produce growth factors. Real-time PCR analyses and GADPH-normalized levels show that compared with non-GA exposed cells (control), treatment of T cells with GA resulted in an increase in levels of IGF-1 and PDGFaa. Each circle, square, or triangle within a given growth factor dataset represents a separate culture. *, *P < 0.05; ***, *P < 0.01, compared with non-GA exposed controls (t test).

Fig. 2. Conditioned medium (CM) from GA-reactive T cells promotes the development of OPCs in vitro. OPCs were identified by PDGFRα labeling whereas cycling cells were tagged by Ki67. Values are mean ± SEM. ***, *P < 0.05; ***, *P < 0.001, compared with control non-T-cell-exposed medium. *, *P < 0.05; **, *P < 0.01, compared with control T-cell CM (one-way ANOVA with Bonferroni multiple comparisons).

Fig. 3. GA treatment increases injury-induced rise of OPC numbers. (A) The number of PDGFRα-positive OPCs were counted in the dorsal column of 10 sections, each spaced 100 μm apart and centered around the lesion epicenter. The sum per mouse was then plotted in A, where each value is from a separate mouse. There were 7 uninjured mice, 8 lysolécin-vehicle mice, and 11 lysolécin-GA animals. ***, *P < 0.01; ***, *P < 0.001 (one way ANOVA with Bonferroni multiple comparison test). (B) Data of OPC numbers (mean ± SEM) in discrete sections around the lesion epicenter are displayed, to indicate that the increase in OPCs induced by GA is widespread across several sections containing demyelination. *, *P < 0.05, one way ANOVA with Bonferroni multiple comparison test. All analyses were blinded.
RNA was collected and subjected to real-time PCR. Fig. 4 shows that lysolecithin injury led to the rise of levels of the Th2 cytokine, IL-5; and IL-12, a proinflammatory cytokine expressed by APCs including macrophages and important in the generation of Th1 cells (IFN-γ was not detected). Analysis of the ratio of IL-5 to IL-12 from individual mice showed that this ratio was highly elevated by GA compared with vehicle treatment.

For growth factors (Fig. 4), we found that IGF-1 transcript increased after lysolecithin injury in vehicle treated mice compared with uninjured controls, and this was elevated further after injury by GA treatment. BDNF did not rise after injury from uninjured controls, but GA treatment significantly raised BDNF levels.

**GA Treatment Increases Indices of Remyelination 28 Days After Lysolecithin Injury.** In correspondence with the documented remyelination that occurs after lysolecithin-induced demyelination (14–16), we found that the extent of eriochrome cyanine staining was greater in the dorsal column at 28 days of vehicle-treated mice compared with that at 7 days after lysolecithin injury (Fig. 5). An initial extensive demyelination at 7 days, which resulted in a large area of the dorsal column being devoid of intact myelin, was progressively repaired by day 28. Impressively, in mice treated with daily s.c. GA for the first 7 days of injury, eriochrome cyanine staining at 28 days indicated that an even greater degree of remyelination had occurred compared with the profile in vehicle-treated mice (Fig. 5). Much of the initially demyelinated area of the dorsal column in the GA-treatment group contained patches of blue profiles (Fig. S5), reminiscent of “shadow plaques” that have been used to describe remyelination in the human CNS.

We used the pixel intensity of eriochrome cyanine staining to document the area of each dorsal funiculus that was devoid of myelin. Such analyses were conducted across 10 sections spanning...
ning 1 mm and centered around the lesion epicenter. Fig. 5 demonstrates that the remaining demyelination at day 28 was more extensive in vehicle-treated mice compared with that in the GA-treated group, and this was evident not only around the lesion epicenter, but also rostrally and caudally.

Finally, we documented the area of demyelination per section and summed these across 10 sections spanning 1 mm per mouse to obtain an index of the lesion volume. Fig. 5 reveals that the volume of remaining demyelination at day 28 was significantly less in the GA-treatment group (average of 0.26 ± 0.03 mm³ × 10⁻³ (mean ± SEM) compared with vehicle controls (average of 0.55 ± 0.10 mm³ × 10⁻³ (P < 0.05, Mann–Whitney unpaired T test). This was not the result of differing lesion volumes at day 7 after the lysolecithin insult, which was found to be similar in both groups (average of 0.98 ± 0.21 mm³ × 10⁻³ vs. 0.72 ± 0.20, vehicle vs. GA, mean ± SEM, n of 7 each, P > 0.05). Comparing across time points, the lesion volume at day 28 in the vehicle group was 56% of that at day 7 whereas that in the GA group at day 28 was 36% of that at day 7.

Overall, studies of indices of remyelination suggest that GA treatment in the first 7 days after lysolecithin administration favors a more pronounced repair over the ensuing 28 days.

Discussion

Summary of Results. We have determined that in culture, GA-reactive T cells produced growth factors described for oligoden-
drogenesis, and that the conditioned medium from GA-reactive T cells increased the number of OPCs that were formed from embryonic brain cells. After demyelination of the spinal cord in mice, the daily treatment for 7 days with s.c. GA elevated the injury-induced rise of OPCs correspondent with an mRNA profile in the injured tissue that was anti-inflammatory and proreparative. Although the size of the demyelination at day 7 was comparable between the two groups, suggesting that GA did not modulate grossly the initial demyelinating events, determinations at day 28 show increased remyelination when mice were exposed to GA during the first 7 days of injury.

Growth Factors Regulate the Generation of OPCs in Culture, in Development, and in Remyelination. Recovery after demyelination requires the presence of OPCs and the ability of these cells to divide, migrate, mature, and enwrap axons in response to signals in demyelinated lesions. The generation and maturation of OPCs are dependent on several growth factors. In tissue culture studies, the proliferation of OPCs is stimulated by PDGF, among others. PDGF increases the proliferation of OPCs and their maturation into oligodendrocytes (3, 4). IGF-1 has strong ca-
pacity to facilitate both OPC proliferation and maturation (5), and is a survival factor for oligodendrocytes.

Developmental studies have also highlighted the importance of growth factors. Transgenic mice overexpressing PDGF pre-

tended with hyperproliferation of precursor cells whereas PDGF null mice had severely reduced number of OPCs (6). IGF-1 transgenic mice have larger brain and higher myelin content than controls (17). Conversely, IGF-1 null mice have lower number of OPCs at 1–3 weeks of age, and mice with deficient IGF receptor 1 signaling have retarded developmental myelination (18).

Growth factors are also critical during remyelination. The increase of several growth factors within the demyelinated CNS has been reported (19). IGF-1 transgenic mice remyelinated more readily compared with wildtype after demyelination of the corpus callosum; moreover, type 1 IGF receptor null mice did not remyelinate adequately after insult, corresponding with the failure of OPCs to accumulate, proliferate or survive in mutants (20). It must be noted that although we have focused discussions to PDGF, BDNF, and IGF-1, a myriad of molecules has been described to influence myelin formation, including the chemo-
kine GRO-α, hormones, notch signaling and LINGO.

Specific growth factors have been infused or genetically expressed in animal models of demyelination in attempts to promote remyelination. IGF-1 treatment of rats subjected to EAE increased the numbers of proliferating oligodendrocyte precursors (21). For BDNF, a single dose in rabbits subjected to cervical spinal cord ventral root avulsion and replantation promoted remyelination of axons that regenerated from the replantation (22).

The provision of a particular growth factor may sometimes result in proliferation of cells but not their subsequent maturation. This is highlighted by Woodruff et al. (23) where the transgenic increase of PDGFaa led to a rise in numbers of OPCs after lysolecithin demyelination; however, there was a trend toward lower remyelination in the PDGF transgenics, which the authors speculated to be due to the prolonged mitogenic stimulus.

These results indicate that although single growth factor treatment may be adequate to affect repair, this may be subop-
timal or counterproductive in some cases. The concomitant provision of multiple growth factors, with proliferative and/or differentiating capacity, would appear to be required to enhance repair.

Beneficial Aspects of Neuroinflammation. One means to deliver multiple growth factors into the CNS takes advantage of the observation that leukocytes have the capacity to generate a range of growth factors (7). For example, activated T cells, B cells, and macrophages express nerve growth factor and BDNF in vitro, and inflammatory infiltrates of T cells, NK cells, and macrophages in the CNS of animals afflicted with experimental autoimmune encephalomyelitis (EAE) were immunoreactive for neurotrophin-3 and BDNF (24). These reports highlight that there are beneficial aspects to neuroinflammation, which should not be unexpected since a main role for an inflammatory response in other tissues is to enhance repair. In accordance, Schwartz and colleagues (25) have noted that even autoreactive T cells could prevent neuronal death in neurotrauma.

There is increasing evidence that inflammation aids re-
myelination. The depletion of macrophages (26) or T cells (27) impairs remyelination after lysolecithin-induced demyelina-
tion. Conversely, the promotion of an inflammatory response by creating a stab injury to the spinal cord (28) or by injecting zymosan along with OPC transplants into the retina (29) increased remyelination.

The benefits of inflammatory cells are attributed in part to their production of neurotrophic factors described above but other aspects of an inflammatory response may also help. For instance, proteases released by inflammatory cells may help remove nonpermissive molecules, such as proteoglycans that hinder recovery (14). The challenge of harnessing beneficial neuroinflammation is how to avoid its potential detriments, given the extensive literature that persistent neuroinflammation can harm the CNS.

Harnessing the Benefits of Inflammation with GA. GA has been in clinical use to treat MS for over 15 years where it has been shown to be safe, to reduce relapse rate and to produce longterm stabilization of disease in a significant proportion of patients (8). The mechanisms of GA in MS are multiple, and include the generation of Th2 cells and Type 2 monocytes (9, 13). These cells are not known to harm the CNS and may elicit CNS outcomes differently from other inflammatory cell types. When Th2 cells were contrasted with proinflammatory Th1 cells, it was found that Th2 cells interacted with glia to elevate neurotrophins; in contrast, the interaction of Th1 cells with glia did not affect neurotrophins but enhanced the levels of proinflammatory molecules (30). In our study, we were unable to reproducibly detect T cells at sites of demyelination by immunohistochemistry.
or flow cytometry (data not shown), even though the PCR data (Fig. 4) suggested a Th2:Th1 bias in GA-treated mice. Future studies will need to examine representation of Th2 versus Th1 T cells, or Type 2 monocytes, in detail in the demyelinated spinal cord after GA-treatment.

In accordance with potentially useful outcomes of GA for the CNS, GA has been found to confer neuroprotection in animal models of optic nerve injury, EAE, ALS, Parkinson’s, and Alzheimer’s disease (31). GA has also been studied in the context of neural repair. Immunoglobulins harvested from GA-primed mice and injected into mice with viral-induced demyelination of the spinal cord promoted remyelination (32); whether similar outcome could result from daily s.c. injections to mimic the mode of treatment of MS patients with GA was not addressed. Treatment of EAE mice using GA has been reported to increase the extent of neurogenesis (33) and the number of OPCs (34) in the CNS. Overall, these studies support the repair potential of GA that is suggested by our study.

Conclusion

We have determined that the MS medication, GA, promotes oligodendrogenesis and indices of remyelination in mice subjected to lyssolecithin-induced demyelination. This demonstration of the myelination capacity of GA injections in a non-inflamatory animal model, suggests the direct effect on myelin formation is not solely due to the reduction of a harmful inflammatory milieu. The results herein indicate that it is possible to employ GA to harness the benefits of neuroinflammation for repair. The data also suggest that the long term benefits observed in patients on GA treatment (8) may in part be due to remyelination, and this deserves further evaluation.

Materials and Methods

Assessment of Neurotrophic Factors Produced by GA-reactive T Cells in Vitro. Responder and APC populations (see SI Text) freshly harvested from mice were cocultured (3 million each, in 3 mL medium) in 6-well plates in the absence or presence of 10 μg/mL GA. Three days after, total RNA was extracted from cells with 1 mL of TRIzol solution (Life Technologies BRL). The mRNA expression of IGF-1, BDNF, PDGFaa, and the housekeeping gene, 18s rRNA, was analyzed by real-time PCR. All PCR primer sets were purchased from SA Biosciences (IGF-1, PPM03387A; BDNF, PPM03006A; and PDGFaa, PPM03103A). For 18s rRNA the following forward (F) and reverse (R) primers were used: F: GTACACCGTT-GAACCCCAT, R: CCATCCAATCGGTAGCG. Value of growth factor expression from each PCR was normalized to 18s rRNA. Then, the same PCR reactions were analyzed by comparing the ratio of normalized growth factor value for a given T-cell culture exposed to GA versus a sister culture that is not, the relative fold change in transcript was thus obtained for that culture.

To verify that protein for growth factors was also elevated in GA-reactive T cells, conditioned media collected for cytokine analyses described earlier were subjected to the measurement of a selected growth factor, IGF-1, using ELISA (Biosource). Furthermore, the cell conditioned medium was analyzed for their capacity to promote OPCs in culture as described in SI Text.

Lyssolecithin-Induced Demyelination in the Spinal Cord of Mice and Treatment with GA. C57BL/6 adult 2-3-month-old male mice were anesthetized i.p. with a mixture of ketamine (85 mg/kg) and xylazine (15 mg/kg). A 1.5 μL solution of 1% D-lysophosphatidylcholine (lyssolecithin; Sigma) was injected over 1.5 min into the dorsal column of the T3 spinal cord segment using a 32 G needle, with 5 mL Hamilton syringe (Hamilton). The needle was left in the spinal cord for an additional 2 min to prevent backflow of the lyssolecithin. Muscles, connective tissue and skin were then sutured above the injection site. Animals received daily injections of 2 mg per mouse of GA dissolved in saline s.c. for 7 days starting from the day of lyssolecithin administration. This dose was used by Aharoni et al. in mice to increase neurogenesis (33) or document OPC numbers (34) in EAE. Control animals were treated with daily s.c. injections of vehicle (PBS) after extraction lyssolecithin in the brain.

Mice were killed for the evaluation of OPC numbers and cytokine/growth factor content in the spinal cord at day 7 after demyelination, and for the evaluation of myelin reformation at day 28, as described in detail in SI Text.

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amino acids, 1 mM sodium pyruvate, 50 

supplemented with 2 mM L-glutamine, 0.1 mM nonessential 

absence of further GA treatment. Feeding medium was RPMI 

coculture, cells were incubated with 1 

APCs, but in the absence of GA. 

GA-reactive T cells. These were contrasted with control T cells 

after coculture with APCs and GA are referred henceforth as 

days 3 and 5 and interleukin (IL)-2 (10 ng/mL) was added on 

days before harvest, or was restimulated with GA on irradiated 

described earlier was then added to the APCs that were pre-

reactive T cells (Fig. S1), or they were left as control cells in the 

GA (described further below) in culture to enrich for GA-

cells) were then exposed to antigen-presenting cells (APCs) and 

mononuclear cells. These “responder” cells (predominantly T 

cells) were then exposed to antigen-presenting cells (APCs) and GA (described further below) in culture to enrich for GA-reactive T cells (Fig. S1), or they were left as control cells in the absence of further GA treatment. Feeding medium was RPMI medium 1640 containing 1% mouse homologous serum and supplemented with 2 mM L-glutamine, 0.1 mM nonessential amino acids, 1 mM sodium pyruvate, 50 μM β-mercaptoethanol and penicillin/streptomycin. GA (pure, research grade) was provided by Teva Pharmaceutical Industries and was used diluted in PBS. 

Spleens from another group of mice with no prior treatment were taken to obtain mononuclear cells to serve as APCs. Cells were gamma irradiated (Cs-137) using a GammaCell 1000 (Nordion International Inc.) with 3000 Rad for 15 min; this protocol essentially prevents cells from proliferating but they retain the ability to process and present antigen. The APC population was then incubated with 10 μg/mL of GA for 30 min. An equal number of responder cell populations obtained as described earlier was then added to the APCs that were pre-loaded with GA and the resulting coculture was incubated for 3 days before harvest, or was restimulated with GA on irradiated APCs on day 7 and then analyzed 3 days after (Fig. S1). For the latter long-term cultures, feeding medium was replenished on days 3 and 5 and interleukin (IL)-2 (10 ng/mL) was added on these days to improve the survival of cells. The T cells obtained after coculture with APCs and GA are referred henceforth as GA-reactive T cells. These were contrasted with control T cells derived from responder populations that were cocultured with APCs, but in the absence of GA. 

Verification of the Generation of GA- Reactive T Cells in Vitro. Cells were plated into 96-well plates where a single well contained 250,000 cells each of responder and APC populations, in the presence (1–20 μg/mL) or absence of GA. Two days after coculture, cells were incubated with 1 μCi [3H]thymidine. One day after, the resulting radioactivity incorporated into DNA was counted. 

Development and Proliferation of OPCs in Culture. Brains from day 15 embryos were removed, and the anterior entopeduncular area (AEP) was dissected away from the rest of the brain. The AEP was then mechanically dissociated with a Pasteur pipette as described in ref. 1. The single cell suspension of brain cells was plated in media hormone mix (1) at 40,000 cells per mL onto poly-L-ornithine coated glass coverslips placed in a 24-well plate in the absence of growth factors. After 2 h, 700 μL of the medium was removed and replaced with T cell-conditioned medium collected from the coculture in 96-well plate of responder and APC populations, with (10 μg/mL) or without GA, as described above. After 24 or 72 h of culture with the T-cell-conditioned medium, cultures were fixed with 4% paraformaldehyde for 20 min. The cells were then processed for immunocytochemistry directed against PDGFRα (R&D Systems, goat anti-mouse PDGFRα, 1:10) followed by staining for Ki67 (Novoceastra, rabbit anti-Ki67, 1:200). Incubation with the appropriate fluorophore-conjugated donkey secondary antibodies (Jackson Immunoresearch) and Hoechst 33258 (Sigma) followed. The coverslips were mounted onto glass slides and the entire surface of each coverslip was scored for PDGFRα-labeled cells and the number that colabeled with Ki67. 

Tissue Processing for Counting of OPC Numbers. Seven days after lyssolecithin injection into the spinal cord, mice were deeply anesthetized and perfused transcardially with PBS followed by 4% paraformaldehyde solution in PBS. Spinal cords were exposed, dissected out, and immersed in 4% paraformaldehyde solution for overnight fixation followed by 30% sucrose for 3 days. The T1 through T5 spinal cord segments were separated as one block using a scalpel blade, and 10-μm sections were cut on a cryostat. Tissue sections were collected onto slides in a series of one every 10 in a sequence, so that two neighboring sections on the same slide were 100 μm apart from each other. After staining for MBP or eriochrome cyanine, the lesion epicenter was identified by dramatic myelin loss in the dorsal column and by virtue of significant disruption of the pia lining due to the passage of the needle into the spinal cord to deposit lyssolecithin. Each series of 10 sections spanning 1 mm of the spinal cord, and centered around the lesion epicenter, was then analyzed. 

OPCs were detected as described in ref. 2, using a rabbit anti-rat PDGFRα (1:100, from William Stallcup) (3) as this was found in preliminary experiments to be superior in marking OPCs in situ compared with the goat anti-mouse PDGFRα described above for cell culture experiments. The number of PDGFRα+ cells was quantified within the dorsal column of the spinal cord tissue sections. All 10 sections spaced 100 μm apart and centered around the lesion epicenter were tabulated. Counting was done in blinded specimens. 

In cases where other markers were used, we used a mouse monoclonal anti-MBP (SMI-94, 1:500, Covance) to label myelin, and a rabbit anti-Iba-1 to label the microglia/macrophage population (Wako Chemicals). 

Analyses of Cytokine and Growth Factors in the Spinal Cord. After lyssolecithin deposition and daily s.c. injection of GA or saline vehicle for 7 days, as described above, mice were killed and the spinal cord was removed and flash frozen. A 0.8 mm transverse block centered on the needle track was then taken. Although our results consistently show demyelination only in the dorsal column, the entire transverse block of the spinal cord was taken as we were not confident that we could remove only the dorsal columns. RNA was isolated, and real-time PCR analyses for IGF-1 and BDNF were performed as described above, with normalization to GAPDH within each sample. Real-time PCR analyses were also performed for IL-5 and a proinflammatory cytokine associated with differentiation of Th1 cells, IL-12. Primer sets were obtained from SA Biosciences (IL-5, PPM030114A; and IL-12, PPM03020E). GAPDH sequences are F: TTCACCACCATGGAGAAGG and R: GGCATGGACTGTGGTCATGA. 

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**Determination of the Extent of Remyelination.** After lysolecithin injury and the daily s.c. injections of GA or vehicle for the first 7 days, mice were left unmanipulated in order for the expected remyelination to ensue. At 28 days from lysolecithin injection, mice were killed and spinal cord cryostat sections were processed, as described above. Transverse sections were stained with a modified eriochrome cyanine protocol as described in ref. 4. Using the Image-Pro Plus Program (Media Cybernetics), we measured the area within each dorsal column that did not contain eriochrome cyanine staining; this provided an index of demyelination per section based on pixel intensity. We summed this across 10 sections to obtain the volume of demyelination per mouse across 1 mm of spinal cord.

Fig. S1. This schematic explains how GA-reactive cells were generated or analyzed.

Cells or CM* are taken for:
- T cell proliferation
- Cytokine profile
- Growth factors
- Generation of OPCs in culture

*CM: Conditioned medium from cells

T cell
APC
A. T Cells from lymph nodes of GA-primed mice can be restimulated by adding GA in culture

![Graph showing T-cell proliferation](image)

B. Cells exposed to GA become increasingly Th2 polarized

![Graph showing cytokine levels](image)

C. IGF-1 protein is also made by GA-reactive T cells

![Graph showing IGF-1 protein levels](image)

**Fig. S2.** GA-reactive T cells in tissue culture are Th2 polarized and produce growth factors. (A) Exposure to GA resulted in an increase of T-cell proliferation (Cpm = radioactive cpm). (B) Both Th1 (IFN-γ, white bars) and Th2 (IL-5, black bars) cytokines were detected at 3 days of culture. With increasing time in culture, GA-reactive T cells expressed undetectable IFN-γ but increasing levels of IL-5, indicating their progressive Th2 differentiation. The results in A and B (mean ± SEM, n of 4 wells) were reproduced in three experiments. (C) Cells were restimulated with GA (+) from days 1–3 (3d GA+) or received a second restimulation at days 8–10 (10d GA+), or no GA was added (GA−). Analyses of conditioned medium in triplicates show that GA promoted the level of IGF-1 protein. Values are mean ± SEM. ***, P < 0.001; **, P < 0.01; *, P < 0.05, compared with non-GA exposed cells of the same culture duration (t test).
Conditioned medium (CM) from GA-reactive T cells promotes the development of OPCs in vitro. Embryonic day 15 mouse brain cells were exposed to CM from control T cells or CM collected from GA-reactive T cells. Cultures were then stained for OPCs using an antibody to PDGFRα, and colabeled for Ki67. The composite merged images are also displayed (original magnification, 40×.)
Fig. S4. Characteristics of lesion 7 days after lysolecithin-induced demyelination. The top row shows sections from an uninjured mouse whereas the middle row displays 7-day-old lysolecithin lesions, focusing on the dorsal column of the spinal cord (traced in white dotted lines in some panels). The demyelinated area is shown by the lack of MBP immunoreactivity (arrow in middle left image) and a correspondent intense Iba-1 immunoreactivity reflecting microglia/macrophage in the lesion (original magnification, 10×). The bottom panels are high magnification micrographs (original magnification, 40×) of the corresponding areas indicated by the asterisk in the middle row.
Fig. S5. A high magnification view of the lesion in the dorsal column shows a large area devoid of eriochrome cyanine myelin stain in a vehicle-treated mouse and comparably more myelin profiles with GA treatment.