MicroRNA-155 as a proinflammatory regulator in clinical and experimental arthritis

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155 in PB CD14+ cells led to down-regulation of SHIP-1 and an increase in the production of proinflammatory cytokines. Conversely, inhibition of miR-155 in RA synovial CD14+ cells reduced TNF-α production. Finally, miR-155-deficient mice are resistant to collagen-induced arthritis, with profound suppression of antigen-specific Th17 cell and autoantibody responses and markedly reduced arthritic inflammation. Our data therefore identify a role of miR-155 in clinical and experimental arthritis and suggest that miR-155 may be an intriguing therapeutic target.

Rheumatoid arthritis (RA) is a chronic autoimmune disease involving synovial inflammation and adjacent cartilage and bone destruction. RA causes progressive disability associated with early mortality primarily reflecting vascular comorbidity. RA is driven by dysregulated adaptive and innate immune pathways (1) that offer an increasingly rich therapeutic resource. Thus, cytokine inhibitors (e.g., TNF, IL-6 receptor), B-cell depletion, and T-cell costimulatory blockade are components of current standard of care. Partial, transient, or nonresponse is common, however, and clinical or radiographic remission is rarely sustained; significant unmet clinical need remains.

The prospect of targeting multiple pathways simultaneously is attractive to optimize the neutralization of complex effector immune pathways. A class of posttranscriptional regulators termed “microRNAs” (miRNAs) has been identified that appears to be critical for fine-tuning many biological processes and offers the prospect of multiply targeting RA (2, 3). miRNAs are noncoding 22- or 23-nucleotide RNAs that act via formation of an miRNA-induced silencing complex (miRISC). miRNAs block target mRNA translation or induce mRNA cleavage upon binding to mRNA recognition elements within the 3′ UTRs of target mRNA (4).

Emerging data suggest that single miRNA species (miRs) can profoundly alter the phenotype and outcome of immune responses (5–7). For example, increased lymphocyte expression of miR17-92 promotes lymphoproliferation and occurrence of autoimmune manifest in anti-DNA autoactivity (5). Moreover, miRNA dysregulation has been reported in a number of pathologic contexts (8). RA peripheral blood mononuclear cells (PBMC) express elevated levels of miR-146a, miR-155, miR-132, and miR-16, with miR-146a and miR-16 particularly associated with disease activity (9). In addition, miR-146a expression in RA CD4+ cells is positively correlated with the levels of TNF-α in both peripheral blood (PB) and synovial fluid (SF) (10). Also, miR-155 and miR-146a are up-regulated, whereas miR-124a and miR-15a are down-regulated in synovial membrane in clinical and experimental arthritis (11–14). MiR-124a regulates the cell cycle (13), and miR-155 modulates the production of metalloproteinases (11) of RA synovial fibroblasts; manipulation of miR-15a expression triggers apoptosis of these cells (14). However, few reports thus far have addressed the presence and functional impact of miRNAs in inflammatory arthritis. Because of the aberrant expression of miR-155 in RA patients (9, 11), we sought to focus on the functional contribution of miR-155 in clinical and experimental arthritis models. We report here that miR-155 is crucial for the proinflammatory activation of human myeloid cells and antigen-driven inflammatory arthritis. These data provide a powerful proof-of-concept for miR-155–based therapeutic approaches that could modulate the aberrant innate and adaptive autoimmunity associated with RA.

Results

MiR-155 Is Up-Regulated in RA Synovial Macrophages and Monocytes and Promotes Production of Proinflammatory Cytokines. Macrophages/monocytes and dendritic cells are a critical source of proinflammatory cytokines in RA synovium (1). However, the mechanism that underlies their chronic activation is poorly understood. miR-155 has emerged recently as an important regulator of myeloid cell biology (15). In situ hybridization revealed that miR-155 is strongly expressed in RA but not OA synovial biopsies, mainly in the lining layer and to a lesser extent in the sublining layer (Fig. 1A and B). Double immunofluorescence staining showed that miR-155 is expressed in a majority of CD68+ macrophages in the RA membrane-lining layer (Fig. 1C). In addition, miR-155 is significantly up-regulated in RA SF-derived CD14+ cells compared with those obtained from PB CD14+ cells (Fig. 1D). These observations suggest that the inflammatory microenvironment present in the RA synovial compartment may trigger the expression of miR-155 in synovial monocytes/macrophages. Indeed, ex...
miR-155 Targets Src Homology 2-Containing Inositol Phosphatase-1 in RA SF CD14+ Cells. To investigate the posttranscriptional mechanisms whereby miR-155 uniquely operates in synovial monocytes, we focused on Src homology 2-containing inositol phosphatase-1 (SHIP-1), because it has been reported to be a target of miR-155 in murine myeloid cells (19), and is a potent inhibitor of many inflammatory pathways, and served as a plausible candidate pathway (20). MiR-155 overexpression in PB CD14+ cells markedly decreased SHIP-1 mRNA expression, whereas inhibition of miR-155 significantly increased the expression of SHIP-1 mRNA in RA SF CD14+ cells (Fig. 2D). To verify that human SHIP-1 is targeted directly by miR-155, the 3′ UTR of SHIP-1 was cloned in a pMir luciferase system. Cells transfected with pMir-SHIP alone exhibited no change in luciferase expression. Transfection of pMir with sense (s) or antisense (as) SHIP in the presence of control mimic revealed only marginal effects on luciferase expression. In contrast, in the presence of miR-155 mimic, cells transfected with pMir-SHIP(s) but not pMir-SHIP(as) showed diminished expression of luciferase (Fig. 2B). These results demonstrate that miR-155 can directly target human SHIP-1 for degradation.

SHIP-1 Expression Is Down-Regulated in RA SF CD14+ Cells and RA Synovial Tissue Macrophages. Given the direct interaction of miR-155 and SHIP-1 in CD14+ cells, we next examined the expression of SHIP-1 in RA patients. Consistent with the high expression of miR-155 in RA synovial CD14+ cells (Fig. 1A), the expression of SHIP-1 mRNA in these cells was markedly down-regulated compared with that in matching PB CD14+ cells (Fig. 2C). Moreover, consistent with the distribution of miR-155 in RA synovium, SHIP-1 protein was not expressed in cells of the superficial lining of PB CD14+ cells to 25% cell-free RA SF to mimic this milieu strongly up-regulated the expression of miR-155 (Fig. 1E).

To examine possible factors responsible for this regulation, PB CD14+ cells next were incubated with candidate stimuli previously implicated in enhanced cytokine production in RA synovitis, such as Toll-like receptor (TLR) ligands (16) and protease-activated receptor 2 (PAR2) agonists (17, 18). Selective TLR but not PAR2 stimulation increased the expression of miR-155 in PB CD14+ cells (Fig. 1E).

Overexpression of miR-155 in RA SF CD14+ cells suggested that it could be involved in posttranscriptional control of the inflammatory pathways in these cells. To test this possibility, PB CD14+ cells were transfected with control or miR-155 mimics (confirmation of expression is shown in Fig. S1). Overexpression of miR-155 triggered the production of cytokines and chemokines strongly implicated in RA synovitis, namely TNF-α, IL-6, IL-1β, and IL-8. Furthermore, miR-155 significantly increased LPS-induced proinflammatory cytokine production but decreased IL-10 production compared with control mimic (Fig. 1F). Moreover, miR-155 overexpression triggered TNF-α and IL-6 production by CD14+–derived macrophage colony-stimulating factor (M-CSF) mature macrophages (Fig. S2). Next, CD14+ cells were transfected with either a control or miR-155 inhibitor (antagomir) and were incubated with RA SF to mimic again the synovial microenvironment; then cytokine production was measured. RA SF triggered TNF-α production in cells transfected with control antagomir (Fig. 1G), but this effect was abrogated in cells transfected with miR-155 antagomir. Finally, to confirm that target tissue monocytes endogenously matured in vivo could be regulated by miR-155, RA synovial CD14+ cells were isolated, immediately transfected with either control or miR-155 antagomir, and then were reactivated by LPS stimulation. Crucially, miR-155 antagomir inhibited the production of TNF-α (Fig. 1H). Together these data indicate that the chronic proinflammatory phenotype of CD14+ myeloid cells in RA synovium could be regulated by aberrant expression of miR-155.

Fig. 1. MiR-155 is up-regulated in RA synovial macrophages and monocytes and promotes production of proinflammatory cytokines. (A and B) Expression of miR-155 in RA and OA biopsies. (A) Representative staining. (B) Quantitative evaluation of miR-155 expression, *P < 0.05 RA vs. OA. (C) Representative photograph of double staining for miR-155 (green) and macrophage marker CD68 (red) in RA lining layer. Double-positive cells are orange or yellow. (D) Expression of miR-155 in paired PB and SF CD14+ cells from RA patients (n = 4), *P < 0.05 PB vs. SF. (E) TLR ligands and RA SF up-regulate miR-155 expression in PB CD14+ cells (n = 3). Cells were stimulated with TLR ligands or PAR2 agonist or 25% SF for 24 h as described in Materials and Methods. *P < 0.05 stimulated vs nonstimulated. (F) PB CD14+ cells overexpressing miR-155 produce proinflammatory cytokines. CD14+ cells were transfected with control or miR-155 mimic (20 nM). After 24 h, LPS (100 ng/mL) was added to some wells for a further 16 h (n = 5). (G and H) PB CD14+ cells and RA SF CD14+ cells (n = 2 and 3, respectively) were transfected with control or miR-155 antagomir. After 24 h cells were exposed to SF (F) or LPS (G) for a further 24 h. *P < 0.05 as indicated. Data are means ± SEM or box-and-whisker diagrams. Cm, control mimic; miR155sant, miR-155 antagomir; miR155m, miR-155 mimic, C ant, control antagomir; PAR2, PAR2 agonist, (SLIGKV-NH2). In the situ scoring system is described in SI Materials and Methods.
expression was present in CD68+ macrophages in OA but not in RA synovium (Fig. 2D). In contrast, SHIP-1 was expressed in RA CD14+ cells (Fig. 2E). Double immunofluorescent staining revealed that such SHIP-1 expression was present in CD68+ macrophages in OA but not in RA synovium (Fig. 2E). In contrast, SHIP-1 was expressed in RA CD3 and CD20 lymphocyte subsets, suggesting some lineage specificity in the regulation of SHIP-1 (Fig. S3). These data indicate that a functional proinflammatory miR-155/SHIP-1 pathway may be operational in synovial cells in RA patients.

miR-155 Is Crucial for the Development of Collagen-Induced Arthritis.

We next examined the contribution of miR-155-regulated pathways to the development and progression of arthritis in collagen-induced arthritis (CIA) in mice. WT or miR-155−/− C57BL/6 mice were immunized with type II collagen in complete Freund’s adjuvant and challenged 21 d later to provoke arthritis. Consensurate with our observations in human synovium, miR-155 expression was elevated in arthritic joints in WT mice by day 34 (Fig. 3A). Although ~50% of WT littermates developed arthritis, commensurate with the numbers expected on the B6 background (21), miR-155−/− collagen-immunized mice did not develop clinical evidence of arthritis (Fig. 3B and C). Furthermore, in contrast to WT controls, miR-155−/− mice developed neither synovial inflammation nor cartilage and bone destruction (Fig. 3D). Control WT and miR-155−/− mice administered PBS developed no symptoms of arthritis. Serum IL-6, VEGF, IL-2, and IFN-γ and the chemokines CXCL1 and CXCL9 were decreased significantly in miR-155−/− mice compared with WT mice (Fig. 4D). Other cytokines either were unchanged [IL-12, FGF-β, IL-10, IL-1α, IL-1β, monocyte chemotactic protein 1, and chemokine (C-C motif) ligand 3] or were below the limit of detection (TNF-α, GM-CSF, IL-4, IL-13, IP-10, and IL-17). Synovial TNF-α expression was significantly reduced in miR-155−/− mice compared with WT mice (Fig. 4B). This reduction was associated with higher expression of SHIP-1 and lower expression of some proinflammatory mediators, including TNF-α and IL-1β in miR-155−/− bone marrow-derived macrophages (Fig S4). To investigate further the mechanism of disease suppression, we measured the development of antigen-specific Th17, Th1, and Th2 cells in draining lymph nodes (DLN) as well as the production of collagen-specific autoantibodies in the serum of WT and miR-155−/− mice. CD4+ cells from miR-155−/− DLNs produced significantly less IL-17 and IFN-γ than did CD4+ cells from WT DLN (Fig. 4C and D). Moreover, CD4+ cells from miR-155−/− deficient mice produced significantly less IL-17 and IFN-γ than did CD4+ cells from WT mice. There was no signifi-
miR-155 has been implicated in the differentiation and activation of cells of both the innate and the adaptive immune systems (3). It regulates myeloid cell differentiation and, putatively, their response to TLR ligands in mice (15, 23–26). It mediates a proposed regulatory role in T-cell homeostasis (27) and in the development of B cells in germinal centers (6, 28). Recently, expression of miR-155 has been demonstrated in the context of several autoimmune diseases, including RA. Up-regulated expression of miR-155 in PBMC (9) and synovial membrane cells (11) of RA patients has been observed. However, given the redundant and highly cell-specific effects mediated by microRNA species, the precise functional implications of miR-155 expression in the immunopathogenesis of arthritis remained obscure.

We now demonstrate that miR-155 is up-regulated significantly in the RA synovial compartment, particularly in CD68+ macrophages in the membrane-lining layer and in SF CD414+ cells and that this up-regulation is associated with the lower expression of the miR-155 target, an inhibitor of inflammation, namely SHIP-1, both in SF CD414+ cells and in macrophages in RA synovial tissue. It coincides with the recent report showing that synovial tissue from RA patients is highly enriched in miR-155 compared with synovia from patients with OA (11), although the latter did not provide cellular specificity. Moreover, CD14+ monocytes and macrophages overexpressing miR-155 exhibited decreased SHIP-1 expression that may lead to increased production of proinflammatory cytokines, including IL-6, which is an inducer for the development of the autoreactive Th17 cells, and TNF-α, an established mediator of chronic inflammation. Both IL-6 and TNF are validated clinical therapeutic targets in RA. Recent studies show that SHIP-1 is essential for the suppression of macrophage activity of macrophages on inflammatory and IL-17–driven responses (29). Interestingly, SHIP-1 is readily detectable in T and B cells in RA synovium (Fig. S3), possibly reflecting a differential role of SHIP-1 in the regulation of macrophage and lymphocyte activation (29). Thus, the miR-155/SHIP pathway may be partially responsible for excessive proinflammatory activation of myeloid cells in inflammatory arthritis. We cannot, however, exclude a role for other miR-155-regulated targets (3) in the modulation of macrophage cytokine responses, and future studies will be important to define the range of pathways that might mediate such effects; candidate pathways include at least suppressor of cytokine signaling 1 (26).

Consistent with the foregoing discussion, we report here that miR-155 deficiency is protective against development of CIA. At a mechanistic level, reduced disease was associated with impaired production of proinflammatory cytokines, development of the autoreactive Th17 cells, production of anti-collagen II antibodies, and lack of articular inflammation. The generation of autoantibodies is prerequisite to the progression of CIA. B-cell–deficient mice are protected against the development of CIA (30). We have

Fig. 4. miR-155−/− mice exhibit reduced development of T- and B-cell responses during CIA. Groups are as in Fig. 3. CLi/miR-155−/− mice show (A) reduced systemic cytokine and chemokine concentrations and (B) reduced expression of TNF-α mRNA in articular tissue. (C and D) CLi-miR-155−/− mice show a reduced differentiation of Th17 and Th1 cells in DLN. Representative staining (C) and quantitative evaluation (D) are shown. (E and F) miR-155 is required for the production of collagen-specific antibodies. Quantitative evaluation of antibody activity in isotypes IgG2a (E) and IgG1 (F) is shown. Data are means ± SEM, n = 15–17; *P < 0.05, miR-155−/− vs. WT mice (CIA protocol). CLi, type II chicken collagen.
unraveled an unequivocal role for miR-155 in the generation of collagen II-specific IgGs. These data reflect a role for miR-155 in antibody class switching and B-cell differentiation into plasma cells, as was demonstrated recently (6, 28). However, it should be noted that miR-155 is dispensable for induction of arthritis following passive transfer of collagen-specific autoantibodies (Fig. S5). In line with this observation, we detected no role for miR-155 in regulating cytokine production in response to immune complexes in vitro; these data suggest that the effects of miR-155 in myeloid lineage activation may be somewhat stimulus dependent.

Disruption of Th17 pathway components (31, 32) and myeloid-cell activation (33) protects mice against several rodent models of arthritis. Our data suggest that miR-155 seems to be indispensable for the development of antigen-dependent induction of arthritis and subsequent cytokine-driven articular inflammation. Here we postulate that a modified cytokine milieu produced by myeloid cells may be one of the potential mechanisms responsible for impaired development of collagen-specific Th17 cells during arthritis in miR155-deficient mice, particularly given the potent stimulatory effect of miR-155 on human myeloid cells to produce IL-6 and IL-1β, whose involvement in Th17 differentiation is established (Fig. 1F). In line with this reasoning, we noted a decrease in systemic IL-6 concentrations in miR-155−/− mice. In agreement with our data, Connell et al. (34) recently reported that miR-155 plays a role in the development of Th17 cells during experimental autoimmune encephalomyelitis.

The role of IFNγ-producing Th1 cells in the development of CIA (1) is unclear, with net protective effects postulated on the basis of IFNγR−/− and IL-12p35−/− models (32, 35). Nevertheless, IFN-γ plays an important role in proinflammatory macrophage activation (36). miR-155−/− mice elaborated significantly less IFN-γ-producing cells than the WT mice during CIA. Serum levels of IL-2, a known Th1 expansion factor, were decreased in miR-155−/− mice and possibly could be responsible for an impaired differentiation of Th1 cells in miR-155−/− mice. Although a relative increase in Th2 differentiation caused by v-maf musculoaponeurotic fibrosarcoma oncogene (c-maf) activation was established in miR-155−/− deficient mice under steady-state conditions (7), we did not observe any change in IL-4 production in our experimental model. This result could be explained by strong skewing properties of the adjuvant used in experimental arthritis which differentiates Th1/Th2 differentiation. Also, it has been suggested that miR-155 is required for homeostatic proliferation of regulatory T cells, and miR-155−/− deficient mice show reduced numbers of forkhead box P3-positive cells (27). However, as reported here and elsewhere (34), this deficit did not render miR-155−/− mice susceptible to the development of autoimmunity as occurs in CIA and experimental autoimmune encephalitis.

Consistent with a crucial role of miR-155 in the regulation of proinflammatory cytokine production by human synovial myeloid cells, miR-155−/− mice showed reduced expression of articular TNF-α, reduced levels of VEGF and chemokines including CXCL1 (IL-8 homolog) and CXCL9, which play critical role in articular inflammation, neoangiogenesis of hyperplastic synovium, and recruitment of inflammatory cells into the joint cavity. The central role of cytokines in the effector biology of RA is shown unequivocally in the successful clinical application of biologic inhibitor agents (e.g., agents that neutralize TNF and IL-6 receptor).

The miRNA network is emerging as an important contributor factor in the pathophysiology of RA. The potential of a single miRNA species to modulate many distinct disease-regulatory pathways simultaneously renders miRNAs particularly attractive candidate targets. Therefore, a better understanding of the complex nature of miRNA regulatory interactions with multiple inflammatory pathways is bound to prove important for identifying future targets and developing therapeutic strategies. In this report, we identify a clinically relevant, functional miR-155/SHIP-1 pathway that may be responsible for some of the excessive inflammatory response observed in articular tissues in RA patients.

Materials and Methods

Cell Culture. CD14+ cells from 5F or PB of RA patients and CD14+ cells from PB of healthy control subjects were purified using CD14 MACs MicroBeads (Miltenyi Biotech). Cells were stimulated with LPS (100 ng/mL), CpG, CpG control (both 5 μg/mL), Pam3CSK4 (300 ng/mL), or poly I:C (50 μg/mL) (all from InvivoGen) for 24 h. Also, PB and SF CD14+ cells were transfected with either miR-155 mimic or miR-155 antagonist (each at 20 or 40 nM) or, as controls, with scrambled mimic and antagonist (20 or 40 nM) (Thermo Scientific, Fermo, Denmark). Cells and supernatant were collected after 48 h. LPS (100 ng/mL) was added to some cultures 24 h after transfection for a further 16 h of stimulation.

CIA. CIA was induced in B6 WT or miR-155−/− male littermates at 14 wk of age according to a previously described method (21). Further information is given in SI Materials and Methods.

Note Added in Proof. While this manuscript was in revision, Bluml et al. (37) showed similar resistance of miR-155−/− mice to antigen-induced arthritis commensurate with our own observations.

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Synovial tissue specimens and 3βμSSC at RT for 5 min. Next, sections were blocked 1o f4> cells fl After depar-

Male microRNA species 155-negative (miR155 Collagen-induced SSC at 50 °C for two washings of 5 min each, in 0.2

After depar-

Animals. Male microRNA species 155-negative (miR155-negative) mice on a C57BL/6 (B6) background were purchased from Jackson Laboratories (1). To get WT control littermates, miR-155<>/miR-155 mice were backcrossed with B6 mice. Both strains then were bred in-house in a pathogen-free facility. Experiments were carried out according to the guidelines of the UK Home Office.

In Situ Hybridization on Paraffin-Embedded Tissues. After depar-

Some sections were incubated with fluorescein-conjugated anti-DIG antibody (Roche) followed by incubation with monocl-

RA and OA tissues were evaluated using a numerical score based on the number of positive cells in the lining, sublining, and perivascular areas of the section (three different fields in each section), with a score of 0 indicating no positive cells; 1 indicating <10% positive cells; 2 indicating 10–50% positive cells; and 3 indicating >50% positive cells.

Induction and Scoring of Experimental Arthritis. collagen-induced arthritis. On day 0, mice were injected intradermally with a type II chicken collagen/ Freund’s complete adjuvant emulsion (200 μg) (MD BioSciences). On day 21, type II chicken collagen/PBS, pH 7.4 (0.4 μg per well). Serum samples (100 μL) serially diluted in PBS were incubated for 1 h, followed by the addition of biotin anti-mouse IgG2a or IgG1 (BD) and then ExtrAvidin peroxidase (Sigma). Each 1-h incuba-

To establish the transfection efficiency, CD14+ cells were transfected with miR mimic labeled with Dy547 using the N-TER nanoparticle siRNA transfection system (Sigma). After 48 h cells were collected and analyzed by flow cytometry (FL-2). To determine at the antigen-specific T-lymphocyte response, draining (popliteal) lymph nodes cells from WT and miR-155<>/miR-155 mice that underwent the CIA protocol were analyzed as described previously (3). Briefly, cells were stimulated with phorbol 12-myristate 13-acetate/ionomycin for 4 h; GolgiStop was added during the final 3 h. The cells then were incubated with anti-mouse CD16/32 antibodies (BD Biosciences) to minimize non-specific Fe-staining, followed by allophycocyanin (APC)- or phycoerythrin (PE)-conjugated anti-CD4 (BD Biosciences) and PE- or APC-conjugated anti–IL-17, anti–IL-4, or anti–IFN-γ antibodies as appropriate (all from BD Biosciences).

Quantitative PCR. Total RNA was isolated using the miRNeasy kit (Qiagen). The miScript Reverse Transcription Kit (Qiagen) was used for cDNA preparation. TaqMan miRNA assays (Applied Biosystems) or miScript primer assay (Qiagen) were used for se-

For histologic assessment, rear paws were fixed for 3 wk in 10% neutral buffered formalin and were decalcified for 3 wk in 10% nitric acid. Paws then were cut into 7-μm sections and stained with H&E.

Collegen antibodies-induced arthritis. Collagen antibodies-induced arthritis (CAIA) was induced in B6 WT or miR-155<>/miR-155 male mice at 10 wk of age according to a previously described method (2). Briefly, on day 0 mice received the anti-collagen II antibody mixture (8 mg per mouse i.v.) (MD Biosciences). On day 3, 100 μg of Escherichia coli LPS (055:B5; Sigma) was administered i.p. Mice were monitored for clinical signs of disease using microcaliper measurements of paw swelling. A clinical severity score was used in which 0 indicated no reaction, 1 indicated swollen digit(s), 2 indicated erythema, 3 indicated swollen paw/ankle, and 4 indicated loss of function. Mice were killed on day 14.

Cell Culture. To establish the transfection efficiency, CD14+ cells were transfected with miR mimic labeled with Dy547 using the N-TER nanoparticle siRNA transfection system (Sigma). After 48 h cells were collected and analyzed by flow cytometry (FL-2). To determine at the antigen-specific T-lymphocyte response, draining (popliteal) lymph nodes cells from WT and miR-155<>/miR-155 mice that underwent the CIA protocol were analyzed as described previously (3). Briefly, cells were stimulated with phorbol 12-myristate 13-acetate/ionomycin for 4 h; GolgiStop was added during the final 3 h. The cells then were incubated with anti-mouse CD16/32 antibodies (BD Biosciences) to minimize non-specific Fe-staining, followed by allophycocyanin (APC)- or phycoerythrin (PE)-conjugated anti-CD4 (BD Biosciences) and PE- or APC-conjugated anti–IL-17, anti–IL-4, or anti–IFN-γ antibodies as appropriate (all from BD Biosciences).

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Luciferase Activity Assay. The human SHIP1 miRNA target site Δ < 0.05 was considered significant. UTR reverse: TAGTGGATCCTAATTGTGCAG-
UTR forward; CTTGCACTGGGCTTCTTAATG-
which were cloned in both sense and antisense orientations downstream of the luciferase gene in pMIR-REPORT luciferase vector (Ambion). These constructs, named “pMIR-SHIP1-miR155” and “pMIR(A/S)-SHIP1-miR155,” were sequenced to confirm inserts and were used for transfection of HEK293 cells. HEK293 cells were cultured in 96-well plates and transfected with 0.1 µg of pMIR-SHIP1-miR155, pMIR(A/S)-SHIP1-miR155, or pMIR-REPORT, together with 0.01 µg of pRL-TK vector (Promega) containing Renilla luciferase and 40 nM of mir-155 or scrambled miRNA (Thermo Scientific Dharmacon). Transfections were done using Effectene (Qiagen) according to the manufacturer’s instructions. Luciferase activity was measured using the Dual-Luciferase Reporter Assay (Promega) 24 h after transfection.

Human SHIP-1 Staining. Immunofluorescence was performed on formalin-fixed RA and synovial tissues with a double immunofluorescence procedure using a primary antibody against CD3 (mouse monoclonal anti-human CD3, VP-C316; Vector Laboratories), anti-CD68 (mouse monoclonal anti-human CD68, M0876; DakoCytomation), anti-CD20 (mouse monoclonal anti-human CD3, VP-C316; Vector Laboratories), and primary antibodies followed by incubation with secondary biotinylated antibodies and Avidin D fluorochrome conjugates (fluorescein, A-2001, or Dy-Light-594, SA-5594; Vector). Brieﬂy, 5-µm sections of paraffin-embedded tissue were deparaffinized in xylene and rehydrated through graded alcohols to water. To block endogenous peroxidization, sections were incubated with 0.5% H2O2/methanol. Next, epitope retrieval was performed in citrate buffer, pH 6.0. Sections were incubated with 5% horse serum and primary antibodies followed by incubation with secondary biotinylated antibodies and Avidin D fluorochrome conjugates. Slides were mounted in medium containing DAPI (H-1200; Vector) and were scanned on a Zeiss Axiovert S 100 fluorescent microscope.

TaqMan Low-Density Arrays. TaqMan low-density arrays (TLDA) are 384-well microfluidic cards that allow multiplex PCR. Primer sets for a range of inflammatory cytokines and chemokines were generated and used to assess transcriptional fold changes using 18S as an internal control. Macrophages generated from murine miR-155−/− and WT bone marrow precursors were treated with PBS or ultrapure LPS (100 ng/mL) (Invivogen Ltd) for 6 h. Macrophage cDNA (1 µg per sample) was generated via the Qiagen RNeasy and Affi nityscript cDNA kits, then was loaded onto TLDA plates after 1:1 dilution with TaqMan PCR Master Mix No AmpErase (ABI Ltd), and was run on a 7900HT TaqMan reader using SDS software. ΔCt values were generated via the 18S control, and ΔΔCt values were obtained by normalizing the mean ΔCt with the untreated control. Relative quantitative values were calculated as 2^−ΔΔCt. Results are expressed as fold change over untreated control.

Statistical Analysis. Statistics were performed using the Mann–Whitney test or ANOVA followed by Tukey’s test or student’s t-test using SPSS Software. All data are expressed as mean ± SEM. P < 0.05 was considered significant.

Fig. S1. Efficient transfection of human CD14+ cells with microRNA mimic. Peripheral blood CD14+ cells were transfected with miR-155 mimic or control scramble mimic labeled with Dy547 (both 20 nM) using the N-TER nanoparticle siRNA transfection system or were left untransfected. After 48 h, cells were analyzed by flow cytometry, or RNA was isolated and quantitative PCR for miR-155 and endogenous control snU6B was performed. (A) Seventy-nine percent of CD14+ cells were considered transfected with labeled mimic. (B) Expression of miR-155 normalized to snU6B in cells transfected with miR-155 and scramble mimic is shown. Cm, control scramble mimic; miR-155m, miR-155 mimic. Data are means ± SEM of three experiments. *P < 0.05, control vs. miR-155 mimic.

**Fig. S2.** miR-155 promotes spontaneous production of proinflammatory cytokines by human macrophages. CD14⁺-derived macrophage colony-stimulating factor differentiated macrophages were transfected with miR-155 mimic (miR-155m) or control scramble mimic (Cm) (both 20 nM) using Dharma FECT 3 transfection reagent (Thermo Scientific). Cell supernatants were analyzed 48 h later for IL-6 and TNF-α. Data are means ± SEM of two experiments.

**Fig. S3.** SHIP-1 is expressed in both T and B cells in RA synovium. (A and B) Synovial specimens from RA (n = 5) and OA (n = 5) patients were incubated with anti-human SHIP-1 and either anti-human CD3 (A) or anti-human CD20 antibody (B), or appropriate isotype controls. Representative staining from one of five specimens with similar results is shown; blue, DAPI staining; red, CD3⁺ or CD20⁺; green, SHIP-1⁺; yellow, SHIP-1⁺/CD3⁺ or SHIP-1⁺/CD20⁺ cells. Magnification is as indicated.
Fig. S4. The expression of SHIP-1 and proinflammatory cytokines/chemokines in miR-155−/− macrophages. (A) SHIP-1 is up-regulated in miR-155−/− macrophages. RNA from WT and miR-155−/− bone marrow-derived macrophages stimulated with LPS (100 ng/mL) for 24 h was isolated, and expression of mouse SHIP-1 was measured with quantitative PCR. Data are means ± SEM of two independent experiments. (B) miR-155−/− macrophages show lower expression of proinflammatory cytokines/chemokines. WT and miR-155−/− bone marrow-derived macrophages were incubated with medium or LPS (100 ng/mL). After 6 h, total RNA was isolated, and mouse cytokine/chemokine TLDA was performed.

Fig. S5. miR-155 is dispensable for the development of CAIA. On day 0, WT and miR-155−/− mice (n = 8 per group) were injected with the anti-collagen II antibody mixture (8 mg per mouse; i.v.) or PBS. On day 3, 100 μg of E. coli LPS was administered i.p. Mice were killed on days 3, 6, and 14. (A) miR-155 expression is up-regulated in articular tissue of mice. (B–D) The magnitude of CAIA is similar in miR-155−/− mice and in WT animals. Incidence of disease (B), mean rear paw clinical score (C), and paw size (D) were not significantly different. (E) miR-155−/− and WT macrophages do not differ in the production of TNF-α after stimulation with immunocomplexes. Macrophages were incubated with PBS (control) or immunocomplexes (IC; 25 μg/mL) for 24 h followed by TNF-α ELISA. Data are means ± SEM or box-and-whisker diagrams. (n = 8). *P < 0.05, PBS vs. collagen type 2 antibodies or collagen type 2 antibodies plus LPS. CII Abs, collagen type 2 antibodies.