Blockade of TNF-α rapidly inhibits pain responses in the central nervous system

Andreas Hess, Roland Axmann, Juergen Reich, Stefanie Finzel, Cornelia Heindl, Silke Kreitz, Marina Sergeeva, Marc Saake, Meritxell Garcia, George Kollias, Rainer H. Straub, Olaf Sporns, Arnd Doerfler, Kay Brune, and Georg Schett

There has been a consistent gap in understanding how TNF-α neutralization affects the disease state of arthritis patients so rapidly, considering that joint inflammation in rheumatoid arthritis is a chronic condition with structural changes. We thus hypothesized that neutralization of TNF-α acts through the CNS before directly affecting joint inflammation. Through use of functional MRI (fMRI), we demonstrate that within 24 h after neutralization of TNF-α, nociceptive CNS activity in the thalamus and somatosensory cortex, but also the activation of the limbic system, is blocked. Brain areas showing blood-oxygen level-dependent signals, a validated method to assess neuronal activity elicited by pain, were significantly reduced as early as 24 h after an infusion of a monoclonal antibody to TNF-α. In contrast, clinical and laboratory markers of inflammation, such as joint swelling and acute phase reactants, were not affected by anti-TNF-α at these early time points. Moreover, arithmetic mice overexpressing human TNF-α showed an altered pain behavior and a more intensive, widespread, and prolonged brain activity upon nociceptive stimuli compared with wild-type mice. Similar to humans, these changes, as well as the rewiring of CNS activity resulting in tight clustering in the thalamus, were rapidly reversed after neutralization of TNF-α. These results suggest that neutralization of TNF-α affects nociceptive brain activity in the context of arthritis, long before it achieves anti-inflammatory effects in the joints.

Arthritis is one of the most disabling chronic human diseases. Rheumatoid arthritis (RA) affects up to 1% of the population and is characterized by pain, swelling, and stiffness of joints, leading to a serious decay of life quality. Pain is the initial and prevailing symptom of the disease, leading to immobility, which in turn causes complications such as osteoporosis and cardiovascular disease. During the last 10 y the pharmacologic treatment of diseases such as RA has substantially improved because of the development of cytokine blocking agents (1).

Although inflamed joints express a multitude of mediators, including cytokines, chemokines, and growth factors, which contribute to the pathogenesis of arthritis, inhibition of TNF-α has emerged as a particularly successful therapeutic strategy (2, 3). The therapeutic success of TNF-α blockade in RA is unique and has been largely considered to result from rapid and efficient neutralization of joint inflammation based on breakdown of the inflammatory cytokine network in the affected joint, which results in an improvement of the signs and symptoms of the disease (4). It has, however, always been stunning, how fast the blockade of TNF-α improves the patient condition, in particular because diseases like RA are highly chronic, building up a vast amount of inflammatory tissue, and leading to irreversible damage of the cartilage and the bone. Thus, rapid resolution of this highly organized inflammatory tissue or tissue damage is very unlikely to explain the fast effect of TNF-α blockade. Even more importantly, neutralization of other inflammatory mediators, in particular IL-1, which is a central inducer of inflammation and structural damage in arthritis (5), appears to have less-pronounced and less-rapid effects on the symptoms of RA, although its role in protecting structural changes in the joints is substantial (6, 7).

These observations, and the fact that traditional antirheumatic drugs such as methotrexate achieve far slower clinical responses than TNF-α blockers, have led us to suspect that blockade of TNF-α could have additional effects that go beyond the sole inhibition of joint inflammation. We hypothesized that TNF-α blockade could influence processes in the CNS, which control the patient’s perception of the disease state. Indeed, earlier observations in mice have suggested that TNF-α can induce a depressive-like behavior in mice (8). Pain processing and sensation are key mechanisms in the CNS elicited by arthritis, and pain is the dominant symptom of disease, which by far has the strongest impact on the disease burden of arthritis. Importantly, effects on pain also drive the standard read-out parameters for measuring therapeutic response in arthritis. Thus, if TNF-α blockade has unique pain-reducing properties, its efficacy in reducing clinical disease activity of RA will be high, as such disease-activity measures are strongly influenced by pain. In consequence, other treatment strategies, lacking such CNS effects, will less strongly affect the patient’s disease state, even if showing similar anti-inflammatory or structure-sparing effects.

Interestingly, inflammation is considered to lower the threshold for pain perception in the CNS, leading to hyperalgesia (9). In contrast to the well-documented role of TNF-α as a proinflammatory cytokine, its role as a mediator of pain is incompletely characterized. It is known that both receptors for TNF-α are expressed in dorsal root ganglion neurons and that intra-arterial injection of TNF-α blocking agents reduces nociception elicited by joint inflammation (10). TNF-α is also involved in development of mechanical hyperalgesia following injury (11, 12). Hence, we hypothesized that TNF-α leads to a major change in pain perception in the CNS during arthritis.

To search for the rapid effects of TNF-α blockade, we visualized cerebral nociception elicited by arthritis and its reversibility upon TNF-α blockade using blood-oxygen level-dependent (BOLD) functional MRI (fMRI) as early as 24 h after initiation of the treatment. The BOLD signal reflects changes in hemodynamics linked to increased or decreased neuronal metabolic ac-


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1A.H., R.A., and J.R. contributed equally to this work.

2To whom correspondence should be addressed. E-mail: georg.schett@uk-erlangen.de.

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tivity in response to outer sensory stimulation, thus allowing localization of brain areas activated by stimuli (13, 14). As a proof-of-concept, we first evaluated whether TNF-α blockade rapidly reverses the hypernociception in patients with RA. We then performed an in-depth analysis of this rapid effect of TNF-α blockade in a mouse model of arthritis based on the over-expression of human TNF-α.

**Results**

**Reduction of Nociceptive Responses in the CNS of Arthritis Patients by TNF-α Blockade Preceding Its Anti-Inflammatory Effects.** To test whether TNF-α blockade indeed affects the CNS, we first performed a proof-of-concept study in humans. We measured CNS activity in patients with RA by assessing the BOLD signal in the fMRI before and after exposure to an infusion of 3 mg/kg of the TNF-α blocker infliximab (IFX) (15, 16). The BOLD signal was induced by compression of the metacarpophalangeal joints of the dominantly affected hand, which is a standard assessment procedure for RA. Interestingly, the size of the brain area with enhanced BOLD activity was significantly reduced as early as 24 h after administration of the TNF-α blocker IFX. Moreover, it remained low up to 6 wk after the first infusion (Fig. 1A). In contrast, BOLD signals elicited by a standardized control procedure, such as finger tapping, were not affected by TNF-α blockade (Fig. 1B). Importantly, clinical measures of disease activity, such as joint swelling and joint tenderness, composite disease activity scores (DAS), such as DAS28, and acute phase reactants, such as blood sedimentation rate, serum C-reactive protein levels, and serum IL-6 levels, did not change measurably within 24 h after TNF-α blockade but improved later during the treatment (Fig. 1C–E and Table S1). In contrast, subjective rating of pain intensity using the visual analog scale (VAS) was decreased within 24 h after TNF-α blockade (Fig. 1F). Thus, these data indeed indicate a fast effect of TNF-α blockade on the pain responses in the CNS even before a measurable anti-inflammatory effect is achieved.

**Reduced Activity in CNS Regions Involved in Pain Perception and in the Limbic System After TNF-α Blockade.** A more detailed analysis of the BOLD signal in arthritis patients showed that reduction of activated brain area size after TNF-α blockade could be attributed to the CNS regions contralateral to the affected joint (Fig. 2A–C). No significant side difference was found for finger tapping, which was used as a control procedure (Fig. 2A–C). TNF-α blockade did particularly change the activity of CNS structures, which are involved in pain perception, such as the thalamus and the secondary somatosensory cortex (Fig. 2D and E). Interestingly, centers of the limbic system were also affected, such as the cingulate and insular cortex. The cingulate cortex is particularly involved in forming and processing of emotions and memory, whereas the insular region is important for the subjective sense of the inner body, including the experience for pain and emotions (Fig. 2D). These findings suggest that not only pain responses are affected by TNF-α blockade but also CNS structures relevant for creating pain perception, emotions, and body experience.

**Nociceptive Responses in Arthritic Mice Overexpressing Human TNF-α and Their Rapid Reversal by TNF-α Blockade.** To study the effect of TNF-α blockade on CNS pain responses elicited by arthritis in more detail, we used an animal model, which is characterized by an overexpression of human TNF-α in the peripheral joints because of knocking the human TNF-α gene into the mouse genome (17, 18). These TNF-α transgenic (TNFtg) mice spontaneously develop inflammatory arthritis, which leads to progressive functional impairment. The disease itself manifests as progressive swelling of the peripheral joints associated with decreased grip strength, mirroring progressively impaired joint function (Fig. S1). We first assessed the effect of TNF-α overexpression on behavioral pain responses and their reversibility upon TNF-α blockade by IFX. Spinal reflex arches, as measured by the tail-flick test, were not affected in early arthritis of TNFtg mice but were increased in latency at later disease stages (Fig. S2A). When testing mechanical hyperalgesia using von Frey filaments, the force (threshold) needed to elicit responses was significantly lower in early (6 wk) and later stages of the disease (10 wk) (Fig. 2B). The Hargraves test, assessing thermal hyperalgesia, showed a similar pattern, as observed with mechanical hyperalgesia (i.e., increased sensitivity to nociception in early and late stages of the

![Fig. 1. Rapid reversal of pain induced BOLD signals after TNF-α blockade in patients with RA. (A and B) Area of BOLD signal based on fMRI scans elicited by joint compression (A) and finger tapping (B) in patients (n = 5) with RA before, 1, 14, and 42 d after administration of TNF-α blocking agent IFX. (C) Area of BOLD signal (C–E) maps of BOLD activity in a single patient before (Top) as well as 1 (Middle) and 42 d (Bottom) after administration of IFX. Asterisks indicate significant difference to baseline (P < 0.05).](www.pnas.org/cgi/doi/10.1073/pnas.1011774108)
The amplitude of the BOLD signal was significantly increased at early disease stages in TNFtg mice (Fig. S2C). However, in later stages, the amplitude was significantly reduced rather than enhanced in TNFtg mice, both in early and late disease stages, suggesting a desensitization to visceral nociception in TNFtg mice (Fig. S2D). The Rotarod test measuring motor activity was not impaired at an early disease stage but significantly decreased at later stages (Fig. S3A). Similar data were obtained when the suspended-tail test was applied, which showed a significant decrease in overall activity in TNFtg mice but only in later stages of disease (Fig. S3B).

To test whether this nociceptive sensitization is reversible, we treated TNFtg mice with the TNF-α blocker IFX. Whereas the latency of spinal pain reflex arches, as measured by the tail-flick test, were normalized 72 h after TNF-α blockade (Fig. S2E), increased sensitivity to mechanical nociceptive stimuli in TNFtg mice was completely reversed as early as 24 h after injection (Fig. S2F). This profound and rapid antinociceptive effect of TNF-α blockade was further substantiated by the Hargreaves test, which normalized within 24 h after TNF-α blockade (Fig. S3A). This pattern was completely reversed upon TNF-α blockade as early as 24 h after injection. Aside from higher amplitudes, brain activation was more widespread in TNFtg mice, affecting many more brain regions than in WT mice. In the latter, the nociceptive response was localized to several well-defined brain areas (vertical pattern in Fig. 4A, white circles). This spreading of brain activation in TNFtg mice was partially reversed upon blockade of TNF-α (Fig. S3D).

**Mapping of Nociceptive Brain Responses Elicited by Arthritis and Their Modulation by TNF-α Blockade.** Comparative fMRI analysis of the brain-activity patterns elicited by heat stimulation of the hind paw revealed a significant increase of the BOLD activity in TNFtg mice in the so-called pain matrix (19, 20) of the brain compared with the WT mice (Fig. 3A–C). TNF-α blocked to a fast and complete reversal of the increased brain activity in TNFtg mice to the level of WT mice within 24 h. When quantifying the BOLD activity by measuring the area size and peak amplitude of the BOLD signal, we observed a consistent and significant increase in the somatosensory cortex of TNFtg mice, which was completely abolished after TNF-α blockade (Fig. 3C). Smaller, but still significant, increases (Fig. 3C) were also observed in the thalamus, as well as in the association cortex in TNFtg compared with WT mice, which were again completely reversed upon blockade of TNF-α (Fig. 3D).

On the basis of these observations, we aimed at precisely mapping the interaction of the temporal structure of the BOLD signals in arthritis (Fig. 4A). Again, amplitudes of BOLD signals were generally higher in TNFtg mice than in WT mice. Moreover, even low-intensity subthreshold stimulation led to increased brain activity in TNFtg mice, which was not found in WT mice (red color in Fig. 4A at S1/S2 and peaks in Fig. 4B). This pattern was completely reversed upon TNF-α blockade as early as 24 h after injection. Aside from higher amplitudes, brain activation was more widespread in TNFtg mice, affecting many more brain regions than in WT mice. In the latter, the nociceptive response was localized to several well-defined brain areas (vertical pattern in Fig. 4A, white circles). This spreading of brain activation in TNFtg mice was partially reversed upon blockade of TNF-α. In addition, the increased BOLD activity was maintained even during the intervals between stimulations and did not completely regress to baseline levels (horizontal red lines in Fig. 4A; red line in Fig. 4B is permanently high above baseline). TNF-α blockade rapidly reversed this prolonged activation, and the BOLD signal in the time periods between the stimulations dropped dramatically, at times below the baseline level. This effect was particularly pronounced in somatosensory cortical and limbic regions of the brain (asterisks at dark blue intervals in Fig. 4A and green line Fig. 4B).

**Rewiring of the Pain Matrix in the CNS by Chronic Arthritis and Its Reversibility by TNF-α Blockade.** Higher-order changes in brain function may not only be reflected in changes of response properties of single brain structures. Instead, dynamic changes of the complex network connectivity (i.e., within the pain matrix)
might occur. Graph theory has proven to have a high impact on the quantitative analysis of complex networks (i.e., brain network organization) (19, 20). We therefore wondered if graph theoretical analysis explaining the functional connectivity of CNS regions provides insight into the dynamic of rewiring of the pain matrix in arthritis and after neutralization of TNF-α. After removing the global (mainly task-driven) signals, cross-correlation of the fMRI time profiles of all structures, which were significantly activated, revealed distinct patterns of interactions within the pain matrix (Fig. S4A, lower triangular half). There was an increased degree of connectivity, clustering, and modularity in arthritic TNFtg mice compared with WT mice (Fig. S4A and Table S2). These changes largely resulted from the formation of a tight cluster of the thalamus, the periaqueductal gray, and the amygdala (Camada-Kawai plots, Fig. S4B). TNF-α blockade led to a rapid partial dissolution of this cluster, resulting in a more diffuse network, indexed by an overall decreased connectivity, clustering, and modularity similar to that observed in WT mice.

Discussion

In this study we show that neutralization of the proinflammatory cytokine TNF-α rapidly affects CNS pain responses elicited by arthritis. Administration of TNF-α blockers rapidly abrogates CNS activity linked to nociceptive stimuli in patients with arthritis, as well as in mice with TNF-α-driven arthritis. This effect occurs immediately after antibody administration and well before anti-inflammatory effects of TNF-α blockade are observed. Our data suggest that TNF-α appears to trigger hypernociception because of changed pain processing in the CNS and a strong linkage to activation of limbic areas involved in pain experience, emotions, and body sensation.

Cytokine blockade has dramatically changed the treatment of inflammatory diseases, particularly of inflammatory arthritides, such as RA. Despite RA being the embodiment of a chronic disease, patients with RA experience a very rapid improvement of disease-related symptoms upon blockade of TNF-α. Although, it takes several weeks to observe a consistent and objective reduction of joint inflammation (16), many patients report on a rapid effect of TNF-α blockade on their subjective disease state. However, the basis of this rapid effect has not been addressed convincingly so far, although CNS effects of TNF-α blockade have always been suspected to play a role in this process. We were stunned by the fact that TNF-α blockade triggered a profound and significant change of CNS activity in RA patients as early as 24 h after its administration, compared with baseline CNS activity. In contrast, joint swelling, as well as composite DAS (DAS28) did not change significantly within the first 24 h, although they improved later. These observations suggest that functional CNS changes clearly precede the improvement of inflammatory signs of arthritis in human patients. Nociceptive signals elicited by joint compression did induce the activity in CNS structures, which are typically involved in pain perception, such as the thalamus and the somatosensory cortex, but also in parts of the limbic system, such as the cingulum and the insular region, which are relevant for the inner body sensation as well as negative emotional experience, including pain experience. Suppression of neuronal activity in these centers may likely explain the rapid improvement of the symptom state of patients with RA after TNF-α blockade, because the perception of nociceptive stimuli in the CNS is impaired and activation of CNS centers involved in shaping the emotional state of the individual—as well as body perception—is blocked.

In murine arthritis triggered by TNF-α overexpression, we observed that behavioral measurements of nociceptive responses showed a sensitization to somatic nociception (hyperalgesia) during arthritis. Moreover, investigation of arthritic mice by fMRI unequivocally demonstrated an altered pattern of brain activation, as measured by an enhanced BOLD activity. Even low-impact subthreshold stimulation led to increased BOLD signals, which could not be depicted in nonarthritic controls. In support of this notion, TNF-α blockade caused a significant and rapid reduction of these nociceptive BOLD signals in the somatosensory and the association cortex, as well as the thalamus, within 24 h. Upon neutralization of TNF-α, both thermal and mechanical hyperalgesia induced by arthritis were completely reversed within 24 h, an observation that supports the
results obtained in RA patients. Within this short period no apparent effect on clinical analog parameters (such as paw swelling) or histopathologic signs of arthritis (such as synovitis) could be observed. This result indicates that the CNS effect of TNF-α blockade precedes its anti-inflammatory effect.

Interestingly, we even observed a decrease in the BOLD signal below the normal levels observed in nonarthritic WT mice. This effect was again particularly pronounced in somatosensory cortical and limbic regions of the brain. One explanation for this decrease of the BOLD signal below the baseline level could be an increased activity of the cerebral inhibitory network developed during this chronic pain state (21). In this context, it is worth mentioning that brain network analysis detected a tight cluster during this chronic pain state (21). In this context, it is worth mentioning that brain network analysis detected a tight cluster of thalamic structures, the periaqueductal gray, and limbic areas, such as the amygdala in TNFtg mice. This finding supports the concept of an increased efficacy of the cerebral inhibitory network in arthritis. This rewiring may have developed as an adaptation to chronic pain preventing the abnormally increased flow of nociceptive signals to higher CNS structures, especially to the cortex. Importantly, TNF-α blockade rapidly induced partial dissolution of this cluster; this happened within 24 h after TNF-α blockade, in parallel with the reduction of the nociceptive BOLD activity and the restoration of the normal nociceptive behavior.

In summary, these data show that TNF-α leads to more intensive, widespread, and prolonged brain activity upon nociceptive stimulation, which is reversible upon neutralization of TNF-α. This reversal of pain sensitization occurs rapidly and is not primarily linked to the anti-inflammatory effects of TNF-α blockade. In patients with RA, a very similar rapid and profound down-regulation of nociceptive brain activity can be observed, which did precede the alleviation of joint inflammation. Although subclinical anti-inflammatory effects of TNF-α blockade within the first 24 h after exposure to TNF-α blockade cannot be ruled out completely, the lack of a drop in acute phase reactant and cytokine levels within this short time interval does at least not support such a concept. Our findings emphasize that changes of nociceptive responses in the brain precede the anti-inflammatory effect of TNF-α blockers. Importantly, these findings may be a good explanation for the rapid improvement of the symptom state induced by TNF-α blocking therapy. These data also suggest that cytokine therapy targeting TNF-α may achieve its (fast or rapid) clinical benefit remote from the inflamed joint through influencing CNS processes. Our findings also extend previous observations on the effects of TNF-α in the CNS (i.e., the regulation of expression of clock genes), which may explain the high prevalence of fatigue in inflammatory diseases (22). Finally, our findings also raise the possibility that the assessment of responsiveness of BOLD activity in the CNS of RA patients and patients with other forms of inflammatory arthritis may be used as a surrogate marker for predicting clinical responses to therapeutic intervention in a much faster way than it is currently realized.

Materials and Methods

Mice. The human TNF-α and TNFtg mice (strain Tg197) were previously described (17). Treatment with IFX (Cenntocor), a neutralizing chimeric human-murine monoclonal antibody directed against human TNF-α, was done by a single intravenous injection of a dose of 10 mg/kg antibody 24 h before testing (15). Clinical evaluation was performed weekly, starting at 4 wk after birth. Arthritis was evaluated in a blinded manner, as described previously (18). All animal experiments were approved by the local ethics committee of the University of Erlangen-Nuremberg.

Tests for Nociceptive Behavior. The tail-flick test was conducted using a tail-flick analgesia meter (Columbus Instruments), Hargreaves test by IITC Plantar Analgesia Meter, and measurement of paw-withdrawal latency upon heat stimulation. The mechanical nociception test was performed by applying an ascending series of von Frey hairs (23). In visceral nociception tests, mice were tested by the tail-suspension test and locomotor activity was tested by Ugo Basile 7750 accelerating Rotarod (Ugo Basile) (24).

Functional MRI in Mice. WT and TNFtg mice (n = 10 per group, 10 wk) were anesthetized with isoflurane and placed on a cradle inside the magnetic resonance tomograph (Bruker BioSpec 47/40, quadrature head coil) (25). The contact heat stimuli sequences (40, 45, 50, and 55 °C, plateau for 5 s, ramp 15 s) were presented at the right hind paw with 3-min and 25-s intervals, three times, using a custom-made computer-controlled Peltier heating device. A series of 750 sets of functional images (matrix 64 × 64, field of view 15 × 15 mm, slice thickness 0.5 mm, axial, 22 slices) were sampled using gradient echo-based Echo Planar Imaging Technique (single shot: TR = 4,000 ms,
Disease-modifying antirheumatic drugs and having active disease with joint tenderness and swelling, received the TNF-α blocker IFX at a dose of 3 mg/kg as an intravenous infusion. Mean (± SD) age was 56.3 ± 8.2 y and mean (± SD) disease duration was 15 ± 3.3 y. The number of tender and swollen joints, joint pain (VAS ranging from 0 to 10), overall disease activity calculated by the DAS based on 28 joints (DAS28) (27), C-reactive protein, and IL-6 levels were assessed at baseline and 1, 14, and 42 d after the infusion.

Functional MRI in Humans. All anatomical and fMRI data were acquired on a 3T scanner (Magnemot Trio; Siemens) using a standard eight-channel phased-array head coil. The ethics committee of the University Clinic of Erlangen approved all procedures and written informed consent was obtained from all patients. For anatomic datasets, we used a T1-weighted MPRAGE-sequence with a field-of-view (field-of-view = 256 mm, matrix size = 256 × 256, voxel size = 1.0 × 1.0 mm², slices = 176, slice thickness = 1 mm, TR = 1,900 ms, TE = 1.13 ms). For each subject, two experiments with different stimulation conditions (first, finger-tapping and second, compression of the metacarpophalangeal joints) were performed. In each experiment, 93 whole-brain images were obtained with a gradient-echo-echo-planar scanning sequence (TR = 3,000 ms, TE = 30 ms, flip angle 90°; field-of-view, 220 mm², acquisition matrix 64 × 64, 36 axial slices, slice thickness 3 mm, gap 0.75 mm).

Functional MRI Analysis. Functional analysis was performed for mice and humans using Brain Voyager QX (Version 10.3) and our own software Magna2D (25), as previously described (28). In summary, after preprocessing (motion correction using sinc interpolation) across the entire animal’s global signal (human: FWHM = 4 mm, mouse: 0.469 mm) and temporal smoothing (FWHM = 3 volumes smoothing), general linear modeling analysis was performed with separate predictors for each stimulus. The statistical parametric mapping obtained were corrected for multiple comparisons, false-discovery rate threshold at z-score level of 3.3, and different groups of activated voxels were labeled as belonging to certain brain structures based on (I) the mouse atlas from Paxinos (29) or (II) the mouse atlas of the human brain (30). The voxels, which were significantly activated by the above criterion, were counted as the activated volume per given brain region. The mean corresponding peak activity was determined for each stimulation temperature for mice and for tapping and compression in humans, averaged over all activated brain structures and finally over all subjects of one experimental group, respectively, to provide a global group comparison.

Graph Theoretical Analysis. Functional connectivity patterns were computed as cross-correlations of the residuals after the global signal mean was removed by linear regression and represented as correlation matrices. Similarly across these correlation matrices was calculated as Spearman rank correlation. Modularity and clustering coefficients, as well as several centrality measures, were computed from the weighted pattern of positive correlations present between brain regions (31). The network modularity index was obtained by optimally subdividing the network into modules, such that most connections are made within modules and only few connections exist between modules (32, 33). Clustering was computed from the weighted graph-theoretical connectivity matrix (34) and scaled relative to a population of 100 random networks with identical degree distribution. Node-strength centrality was measured as the sum of each node’s positive cross-correlations. The networks were visualized using a force-based algorithm after Kamada-Kawai for achieving that all edges are of more or less equal length, and there exist few crossing edges as possible (35).

Statistical Data Analysis. Data are presented as mean ± SEM. Group mean values were compared by the Student’s t tests in a task-specific single-test framework to assess significant differences between the different mouse strains.

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Mapping the immunological homunculus

Betty Diamond and Kevin J. Tracey
The Feinstein Institute for Medical Research, Manhasset, NY 11030

During the first century, the Roman physician Cornelius Celsus defined four cardinal signs of inflammation: redness, swelling, heat, and pain. These signs and symptoms occur during infection by invasive pathogens or as a consequence of trauma. Today, we understand the molecular basis of these physiological responses as mediated by cytokines and other factors produced by cells of the innate immune system. Cytokines are both necessary and sufficient to cause pathophysiological alterations manifested as the four cardinal signs. Importantly, this knowledge has enabled the development of highly selective therapeutic agents that target individual cytokines to prevent or reverse inflammation. For example, selective inhibitors of TNF, a major inflammatory cytokine, have revolutionized the therapy of rheumatoid arthritis, inflammatory bowel disease, and other autoimmune and autoinflammatory diseases affecting millions worldwide. Now, in PNAS, Hess et al. (1) use functional MRI to monitor brain activity and report that patients with rheumatoid arthritis who receive anti-TNF develop significant changes in brain activity before resolution of inflammation in the affected joints.

To accomplish this, the authors measured blood oxygen level-dependent (BOLD) signals in the brain after compressing the metacarpal phalangeal joints of the arthritic hand. They observe enhanced activity in the brain regions associated with pain perception, including the thalamus, somatosensory cortex, and limbic system, regions known to process body sensations and emotions associated with the pain experience (1). Brain activity was significantly reduced within 24 h after treatment with TNF inhibitors, a time frame that preceded any observable evidence of reduced signs of inflammation in affected joints. Clinical composite scores, comprising measurements of C-reactive protein, a circulating marker of inflammation severity, were not improved until after 24 h. This suggests that selective inhibition of TNF has a primary early effect on the nervous system pain centers.

The authors further explore this result in genetically engineered transgenic mice that overexpress TNF and develop spontaneous arthritis. Administration of anti-TNF significantly improved the development of mechanical hyperalgesia in the standardized von Frey filament test system and also improved the response to thermal hyperalgesia in the Hargreaves test. This clinical improvement occurred within a short period after administration of anti-TNF, before any significant improvement in joint swelling or grip strength, and before histopathological improvement of synovitis. As in the human subjects, there was significant abolishment of BOLD signals in the somatosensory cortex. Brain activation was more extensive in the transgenic mice and affected more brain regions compared with WT mice, and this spreading of brain activation was down-modulated by administration of the TNF inhibitors. Administration of anti-TNF reversed the prolonged activation of BOLD activity that was observed during the intervals between stimulations, and was particularly pronounced in the somatosensory cortex and limbic system.

These intriguing findings provide a glimpse into the future, when it is likely that sensory and motor brain regions associated with immune responses will be mapped as an immunological homunculus (2). Of particular significance here is that this approach bridges two major fields of medicine, neurology and immunology.

Reflexes Regulate Immune Responses
Recent advances at the intersection of these fields have provided direct evidence that neural circuits reflexively control the onset and resolution of inflammation, and previously undescribed molecular mechanisms have been defined (3). The fundamental principle of neurological reflexes, as originally established by Sir Charles Scott Sherrington in the early 20th century, is that neural circuits cooperate to maintain the organism’s homeostasis. He elucidated the role of the nervous system in integrating reflexes to provide stable physiological systems. He famously remarked that “Existence of an excited state is not a prerequisite for the production of inhibition; inhibition can exist apart from excitation no less than, when called forth against an excitation already in progress, it can suppress or moderate it.” These principles were first defined in relatively accessible organ systems, including cardiovascular, gastrointestinal, and respiratory, which enabled the mapping of reflex units. For example, by recording signals to the heart, it was possible to define a reflex that controls heart rate: increased heart rate activatesafferent arcs that in turn elicit firing of efferent arcs in the vagus nerve, which slows heart rate.

More recently, these principles have been applied to mapping the somewhat more elusive immune system. The combined use of neurophysiological stimulating techniques and current molecular strategies to measure immune response established the “inflammatory reflex” (3).

Cytokines activate afferent signals in the vagus nerve, which culminate on an efferent arc that inhibits further cytokine production. The molecular basis of this cytokine-inhibiting action is attributable to acetylcholine, the principal neurotransmitter of the vagus nerve, interacting with α7 nicotinic acetylcholine receptor subunits expressed by cytokine-producing cells. Signal transduction through this receptor–ligand interaction down-regulates the activity of NF-κB, a primary pathway for regulating cytokine transcription and synthesis. Accordingly, α7 KO mice rendered deficient in this pathway are exquisitely sensitive to inflammation caused by pathogen-associated molecules and experimental arthritis. The efferent arm of the inflammatory reflex, termed the cholinergic antiinflammatory pathway, has enabled the development of experimental nerve stimulators and highly selective pharmacological agents that suppress cytokine-mediated disease in standardized preclinical models.

Sensing Inflammation and Infection
Less is known about the mapping of the afferent or sensory arc of the inflammatory reflex, and there is significant interest in the question of how the nervous system monitors the state of inflammation. The seminal work of Watkins et al. (4) demonstrated that the fever-causing activities of the pyrogenic cytokine IL-1 required an intact vagus nerve, because administration of IL-1 to the abdomen failed to produce fever if the vagus nerve had been cut. These researchers proposed a critical role of the glomus cell, specialized to detect...
pH and other chemical changes in the internal milieu, in sensing IL-1 (5). Glomus cells express IL-1 receptors, and it is a plausible hypothesis that IL-1 binding to glomus cells stimulates the release of dopamine, which, in turn, triggersafferent signaling in the vagus nerve. Ascending action potentials culminate on the nucleus tractus solitarius and are then relayed to higher brain regions. Sensory neural circuits can also be activated by LPS, and Toll-like receptor 4 (TLR4), the principal endotoxin receptor, is expressed by neurons in the nodose ganglia (6). These neurons have been implicated in transmitting afferent signals in the vagus nerve. Another mechanism by which the nervous system can sense the presence of injury is dependent on the expression of TLR4 in microglial cells, which is required for the development of neuropathic pain in a murine model of nerve transection (7). This TLR4 pathway, which may be activated even in the absence of infection by HMGB1 released from injured cells (8), stimulates TNF release in the development of chronic pain states. Together, these results indicate that the nervous system is hardwired to monitor the presence of cytokines and molecular products of invaders.

The presence of TNF can also be sensed by sensory neurons, which express type I and type II TNF receptors (9). Local or regional administration of TNF elicits pain, partially by inciting tissue damage and activating release of other molecules capable of activating sensory neurons. TNF receptor–ligand interaction on sensory neurons may activate pain fibers directly, however, serving as a pathway that may provide the brain with pain-related information as described here. Others have shown that administration of anti-TNF antibodies into the cerebrospinal fluid of rats subjected to experimental arthritis significantly attenuated pain-related behavior and the development of inflammation in the joints (10). Moreover, intrathecal administration of antibody was significantly more effective compared with systemic anti-TNF, suggesting that the principal effects of anti-TNF are mediated through direct interaction with neurons. Considered in light of the current paper, it is plausible that local accumulation of TNF in the region of sensory pain circuit neurons drives activation of the brain’s pain centers. Thus, it is possible that the rapid early improvement from anti-TNF results from acutely neutralizing TNF and blocking its ability to stimulate pain fibers. Predictably, this would not require any significant improvement in the severity of inflammation in the arthritic joints in order for benefit to be perceived.

The current study also raises another possibility, that anti-TNF antibodies traverse the blood–brain barrier and directly alter the activity of brain neurons in the pain centers. Until quite recently, it has been dogmatic that antibodies do not enter the brain, but this has been overturned by studies in a murine model of the autoimmune disease lupus (11). This work identified antibodies that bind to dsDNA (a common antibody phenotype in this syndrome) and to the NMDA receptor expressed on brain neurons. These antibodies readily access the brain, because the blood–brain barrier can be opened during relatively mild stress caused by administration of endotoxin, cytokines, or even epinephrine. On entering the brain, the interaction of these antibodies with neurons in the hippocampus and other regions mediates cell damage and cognitive or behavioral dysfunction (12).

In light of the present study, it is interesting to determine whether TNF inflammatory mediators produced during arthritis might open the blood–brain barrier to enable anti-TNF direct access to brain neurons. TNF expressed by glial cells in brain regulates synaptic scaling, a mechanism that adjusts the strength of neuronal synapses under conditions of prolonged alterations of electrical activity (13). A testable hypothesis emerges from this reasoning: The systemic inflammation associated with rheumatoid arthritis opens the barrier to anti-TNF, which inhibits glial cell-derived TNF and alters the strength of synapses in the brain’s pain regions. This might explain the observed reversal of prolonged activation of BOLD activity during intervals between stimulations in the arthritic mice.

**Immunological Homunculus**

Advances in brain imaging modalities and molecular biology are mapping brain networks and circuits with high resolution and precision. These maps reveal sensory and motor circuits that are somatotopically and functionally organized, and form the basis for understanding the neural circuits that coordinate physiological responses to the external and internal environment. Early advances were achieved by mapping the relatively accessible domains of body sensations and skeletal muscle motor function. Now, knowledge of the inflammatory reflex as a prototypical circuit that regulates inflammation, combined with the ability to visualize brain activity in patients and mice with inflammation, makes it possible to make additional advances. It is interesting to consider that in addition to defending the host from infection, the immune system functions as a sensory organ that transmits information in real time to the central nervous system about the tissue response to injury and infection. This presents important possibilities for understanding fundamental mechanisms that maintain physiological homeostasis. Progress in this field is all but certain to reveal the identity of other circuits that reflexively regulate the immune system and to produce maps that will guide understanding of the neurological basis of immunity and physiology.