Combined targeting of HER2 and VEGFR2 for effective treatment of HER2-amplified breast cancer brain metastases

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Approximately 25% of human breast cancers demonstrate amplification of the human epidermal growth factor receptor-2 (HER2) protooncogene. Patients with HER2-positive breast cancer are at a high risk of developing brain metastases, with a frequency as high as 50% in patients succumbing to advanced disease (1–4). A combination of factors explains this phenomenon, including the proclivity of HER2-positive tumor cells to colonize the brain and improved control of extracranial disease with the anti-HER2 monoclonal antibody trastuzumab (Genentech/Roche) enabling patients to live long enough to develop brain metastases (1, 5). Although systemic disease is under control in many of these patients, their associated brain metastases appear resistant to trastuzumab (2, 6, 7). Furthermore, lapatinib (GlaxoSmithKline), a small-molecular-weight HER2 kinase inhibitor, thought to penetrate the blood–brain barrier better compared with trastuzumab, has shown only a modest response in patients with recurrent or progressive breast metastases (8–10). Thus, alternative strategies are desperately needed to treat patients with metastatic disease (11).

Angiogenesis is critical to breast cancer progression and metastasis, and adversely affects prognosis (12, 13). VEGF is one of the most potent angiogenic factors (14). In patients with metastatic breast cancer, the anti-VEGF antibody bevacizumab (Genentech/Roche), in combination with paclitaxel, prolonged progression-free survival but not overall survival compared with paclitaxel alone (15). In addition, the use of bevacizumab with trastuzumab and chemotherapy in HER2-positive metastatic breast cancer has shown some promise in phase II trials (16). However, there remain no data on the efficacy of bevacizumab in the context of brain metastases because these patients have often been excluded from clinical trials due to fear of an increased risk of cerebral hemorrhage after anti-VEGF therapy. However, retrospective analysis has shown this potential risk may not be significant (17). Currently, clinical trials are underway to evaluate the efficacy of bevacizumab in patients with breast cancer who have active brain metastases, including its combination with trastuzumab in patients with HER2-positive disease (http://clinicaltrials.gov identifier CT01004172). However, the effects of dual blockade of VEGF and HER2 in brain metastases have not been examined in preclinical models.

We have previously shown that trastuzumab alone can act as an antiangiogenic mixture in a leptomeningeal metastasis model of HER2-overexpressing breast cancer through a reduction in proangiogenic factors and an increase in an antiangiogenic factor (18). These effects were transient, however, and were counteracted by the compensatory production of VEGF from host stromal cells, perhaps to rescue angiogenesis. Indeed, carcinoma-associated fibroblasts as well as immune cells can produce significant amounts of VEGF in breast cancer models (19, 20). These findings suggest that blockade of the VEGF pathway may overcome trastuzumab resistance by inhibiting the activity of the...
compensatory VEGF production by stromal cells, resulting in a significantly greater antiangiogenic effect. This hypothesis is consistent with the finding that targeting both HER2 and VEGF in orthotopic breast cancer xenografts results in better growth delay than targeting either agent alone (21, 22). Furthermore, the phase II study of bevacizumab in combination with trastuzumab and capecitabine as first-line treatment for HER2-positive locally recurrent or metastatic breast cancer showed an overall response rate of 73% (16).

Herein, we developed a model of HER2-amplified breast cancer brain metastases. Although the brain metastases were resistant to single-agent lapatinib or trastuzumab, we observed that the combination of trastuzumab, lapatinib, and an anti-murine VEGFR2 antibody, DC101 [ImClone Systems/Eli Lilly and Company (23)], resulted in a markedly delayed progression of these brain metastases. These studies point to a new therapeutic approach for patients with HER2-amplified breast cancer brain metastases.

Results

Mouse Model That Recapitulates the Differential Response of Intracranial and Extracranial HER2-Amplified Breast Cancers to HER2 Inhibitors. We used a previously established technique to monitor metastatic tumor growth sensitively and noninvasively in the brain in real-time (24). HER2-amplified BT474 cells were infected with a lentivirus leading to expression of both Gaussia luciferase (Gluc) and cerulean fluorescent protein (CFP). This facilitates noninvasive monitoring of BT474-Gluc tumor growth in the brain parenchyma after direct injection (a) by measuring Gluc activity in the bloodstream (Gluc is secreted by tumor cells into the bloodstream) and (b) through the cranial window using intravital fluorescence microscopy, respectively. Fig. 1A illustrates a brain metastatic lesion as seen with bioluminescence imaging and intravital multiphoton microscopy. The BT474-Gluc brain lesion depicted by bioluminescence represents the approximate tumor size on initiation of treatment (Fig. 1A, Left). Intravital multiphoton microscopy illustrates the brain lesion boundary (Fig. 1A, Center) and the abnormality of the tumor vasculature (Fig. 1A, Right). Blood Gluc activity correlated well with BT474-Gluc tumor volume as measured by intravital microscopy (Fig. S1A). Blood Gluc activity also correlated with tumor volume measured using MRI and bioluminescence (Fig. S1B and C). Furthermore, analysis of tumor volume ex vivo using multispectral fluorescent imaging supports the correlation between blood Gluc activity and tumor volume (Fig. S1D). Thus, blood Gluc activity was used to monitor tumor size and response to treatment.

We first tested the effects of anti-HER2 therapies on the growth of established BT474-Gluc tumors growing in the mammary fat pad (primary) and in the brain parenchyma (metastases). Treatment was initiated when brain metastatic tumors reached ~10 mm³ in volume [corresponding to a blood Gluc activity of 10 Relative Light Units per second (RLU/s)], as illustrated in Fig. 1A. Although BT474-Gluc tumors in the mammary...
fat pad responded well to trastuzumab or lapatinib treatment, both HER2 inhibitors failed to control BT474-Gluc tumor growth in the brain parenchyma (Fig. 1 B and C). Thus, this mouse model closely mirrors the discordant effects of therapy in patients with metastatic HER2-positive breast cancer.

**Dual Inhibition of HER2 and VEGFR2 Significantly Slows Brain Metastatic Tumor Growth and Extends Mouse Survival.** Based on our previous findings proposing VEGF as a mechanism of resistance to trastuzumab in HER2-amplified breast cancer brain metastases, we tested if the addition of an anti-VEGF pathway inhibitor to anti-HER2 therapy could control tumor growth better than anti-HER2 therapy alone (Fig. 2). Consistent with our findings above, either trastuzumab or lapatinib monotherapy slowed brain metastatic BT474-Gluc tumor growth by only a few days compared with control-treated tumors (Fig. 2 A and B, respectively). The minimal overall survival benefit of either monotherapy was consistent with its modest effect on tumor growth: trastuzumab prolonged average survival by 7 d (1.4-fold compared with control; Fig. 2A), and lapatinib prolonged average survival by 3 d (1.2-fold compared with control; Fig. 2B). At the time of euthanasia, the size of the brain metastatic tumor was consistent among mice and occupied nearly an entire hemisphere of the brain. Thus, mouse survival in our model was considered to reflect brain metastatic tumor burden. Treatment with the antimurine VEGFR2 antibody DC101 exhibited a significant delay in tumor growth, corresponding to an improvement in survival between 2.3- and 2.4-fold greater than control-treated mice (Fig. 2A and B, respectively; \( P < 0.001 \)). Intriguingly, the combination of trastuzumab (Fig. 2A) or lapatinib (Fig. 2B) with DC101 showed a substantial delay in BT474-Gluc brain metastatic growth. The combination of trastuzumab and DC101 prolonged mouse survival to a median of 47.5 d, almost threefold greater than control-treated mice (additional 31.5 d; \( P < 0.01 \) compared with DC101 monotherapy). The combination of lapatinib and DC101 prolonged mouse survival to a median of 47 d, 3.4-fold greater than control-treated mice (additional 33 d; \( P < 0.001 \) compared with DC101 monotherapy). The impressive tumor growth delay witnessed by the combination of trastuzumab and DC101 was confirmed using MRI (Fig. 3A4), bioluminescent imaging (Fig. 3B), and postmortem ex vivo multispectral brain imaging (Fig. S2), further validating the reliability of blood Gluc activity in monitoring brain metastatic BT474-Gluc tumor growth.

**Combination of Anti-HER2 and Anti-VEGFR2 Therapies Induces Tumor Necrosis Through Antivascular Effects.** To investigate the mechanism(s) for the effect of the combination treatment groups, we evaluated the histology of brain metastatic tumor tissues collected after 15 d of treatment with control IgG, trastuzumab, lapatinib, DC101, or the combination of each HER2 inhibitor with DC101. Immunohistochemical analysis revealed no obvious difference in tumor cell proliferation or apoptosis in viable tumor tissue from the six different treatment groups (Fig. S3). In addition, analysis of tumor tissue collected after 4 or 8 d of treatment revealed no difference in tumor cell proliferation or apoptosis between groups. However, we found a clear difference in the necrotic fraction of tumor tissue between treatment groups after 15 d of treatment (Fig. 4 and Fig. S4). There was a significant increase in necrotic area in tumors treated with DC101.
compared with control- or anti-HER2–treated tumors: 7–8% in control- or anti-HER2–treated tumors vs. 21 ± 1.5% in DC101–treated tumors (P < 0.001). This necrotic fraction was even higher within the combination-treated tumors: 28 ± 1.8% of the tumor area in the trastuzumab and DC101-treated mice and 27 ± 1.8% in the lapatinib and DC101-treated mice (P < 0.001 and P < 0.01, respectively, compared with DC101 monotherapy). These findings indicate an enhanced antiangiogenic effect induced by the combination treatment groups, and thus led us to investigate their effects on the tumor vasculature.

To this end, we stained brain metastatic lesions from all six treatment groups for both the endothelial cell marker CD31 and perfusion marker biotinylated-lectin, infused into the bloodstream 5 min before tumor tissue collection. HER2 inhibitors did not have a significant effect on the total or functional microvascular density (MVD) compared with control-treated tumors, which make up 5.09% and 3.88% of the viable tumor area, respectively (Fig. 5 and Fig. S5A). Mice treated with DC101 showed a decrease in the area of both total and functional viable tumor MVD to 2.75% and 1.84%, respectively, a decrease of ∼50% compared with control-treated mice (P < 0.001). Finally, the combination treatments of trastuzumab and DC101 or lapatinib and DC101 revealed a steeper reduction in the total and functional tumor MVD compared with DC101 monotherapy. The combination of trastuzumab and DC101 decreased total and functional tumor MVD to 1.30% and 0.87% of viable tumor area, respectively, an approximate 75% reduction compared with control-treated tumor tissues (P < 0.05, compared with DC101 monotherapy). Meanwhile, the combination of lapatinib and DC101 decreased total and functional tumor MVD to 1.46% and 1.06%, respectively, an approximate 72% decrease compared with control-treated tumors (P < 0.05, compared with DC101 monotherapy). A similar trend was found in tumor tissues collected after 8 d of treatment (Fig. S5 B and C). These data demonstrate that combined anti-VEGFR2 and anti-HER2

Fig. 3. Imaging of metastatic breast cancer growth in the brain parenchyma and response to the combination of anti-HER2 and anti-VEGFR2 therapy. (A) MRI analysis of control or trastuzumab and DC101 treatment on BT474-Gluc metastatic tumors in the brain. MRI tumor volume was measured on days 0, 5, and 14 of treatment. (Left) Representative MRI of control- and combination-treated tumors is shown. (Right) Tumor volume, calculated using MRI, is plotted for each individual mouse (# corresponds to the tumors illustrated by the MRI scans in A). (B) Bioluminescence imaging analysis of control or trastuzumab and DC101 treatment on metastatic BT474-Gluc tumors in the brain. (Left) Images of control- and combination-treated tumors 14 d after treatment initiation are shown. (Right) Average bioluminescence signal for the three control- and three combination-treated tumors is depicted. *P < 0.05.

Fig. 4. Effects of the indicated treatments on the necrotic fraction of the tumor. The percentage of necrotic area in the total tumor area for each BT474-Gluc tumor was determined 15 d after treatment initiation using an in-house MATLAB program. **P < 0.01; *P < 0.05. DC101 vs. control, trastuzumab, or lapatinib (P < 0.001). Representative H&E-stained sections used for quantification of the necrotic area are shown in Fig. S4.
Fig. 5. Effects of the indicated treatments on blood vessels. CD31-positive blood vessel area (A) and lectin-positive perfused blood vessel area (B) in the viable regions of BT474-Gluc tumors 15 d after treatment initiation. Tissues were analyzed by an in-house MATLAB program. *P < 0.05; **P < 0.01. DC101 vs. control, trastuzumab, or lapatinib (P < 0.001). (C) Representative images of CD31-positive endothelial cells in the viable tumor tissue. (Scale bar: 100 μm.) (Representative images of lectin-positive blood vessels in the viable tumor tissue are shown in Fig. S5A).
therapies lead to a marked decrease in MVD and increased tumor necrosis.

**Delivery of HER2 Inhibitors, NK Cell Infiltration, or Direct Tumor Cell Cytotoxicity Does Not Mediate the Reduction in Metastatic Tumor Growth Caused by the Combination of HER2 and VEGFR2 Inhibitors.** We next investigated the possibility that the anti-VEGFR2 antibody augmented the capacity for the HER2 inhibitors to suppress signaling in the brain metastases. After 3 d of lapatinib, but not trastuzumab, treatment, we observed a significant decrease in phosphorylation of HER2 as well as downstream AKT and ERK in BT474-Gluc brain metastases (Fig. S6A). Consistent with this, after 3 d of lapatinib treatment, we found its concentration to be ~14.5 μM within the brain metastatic tumor (Fig. S6B). Although this concentration is one-third of that reached in tumors growing in the mammary fat pad, it is at least 500-fold greater than the in vivo IC₅₀ for this cell line (25). Interestingly, however, after 15 d of lapatinib treatment, this suppression of HER2 activation was no longer present (Fig. S7B). Western blot and immunohistochemical analysis showed that neither combination treatment group (either trastuzumab and DC101 nor lapatinib and DC101) significantly altered the activation status of HER2 after 15 d of treatment, although a trend toward a modest decrease in HER2 activation was suggested (Fig. S7).

In addition to direct signaling inhibition, another mechanism proposed for trastuzumab’s efficacy is antibody-dependent cell cytotoxicity, mediated by natural killer (NK) cells (26). Although the mechanism of trastuzumab in slowing BT474-Gluc mammary fat pad tumors was not mediated by NK cells (Fig. S8A), we found a significant increase in NK cells within the brain metastatic tumor after DC101 treatment alone or with trastuzumab (Fig. S8B). Therefore, we tested if NK cell depletion using the NK1.1 antibody (clone PK136; BioXCell) could alter tumor response to this combination therapy. Treatment with the NK1.1 antibody for 7 d resulted in a >99% and 95% reduction in NK cells in the peripheral blood and spleen of nude mice, respectively (Fig. S8C). Untreated BT474-Gluc tumor growth in the brain parenchyma of nude mice was not altered due to the absence of NK cells, and the effect that the combination of trastuzumab and DC101 had on tumor growth and survival in nude mice lacking NK cells was no different from that in WT NK cell-competent nude mice (Fig. S8D). These findings are consistent with clinical data showing a lack of association between Fcγ receptor genotypes and trastuzumab efficacy (27). Finally, DC101 had no direct effect, either alone or in combination with trastuzumab or lapatinib, on the growth of BT474-Gluc cells in vitro (Fig. S9). Taken together, these data indicate that the enhanced benefit of the anti-HER2 and anti-VEGFR2 combination therapy in this model is not primarily due to enhanced inhibition of HER2 activity, increased NK cell-mediated cytotoxicity, or direct cytotoxicity.

**Dual HER2 Targeting with Trastuzumab and Lapatinib Delays Brain Metastatic Tumor Growth Better Than Either Monotherapy.** Preclinical and clinical evidence suggests the combination of trastuzumab and lapatinib acts synergistically compared to either agent alone. One report attributes this to the marked downregulation of the protein survivin, leading to enhanced tumor cell apoptosis following the combination of the two agents (28). Another report shows incomplete inhibition of the HER2 kinase after lapatinib monotherapy, and the addition of trastuzumab better suppresses HER3 phosphorylation by blocking ligand-independent interactions between HER2 and HER3 (29). In agreement, we found better growth suppression of BT474-Gluc cells in vitro with the combination compared with either monotherapy (Fig. S10A). Furthermore, these preclinical results are supported by clinical data in both the preoperative primary and metastatic settings (30, 31). Because these findings are limited to extracranial sites (primary and metastasis), we asked if the same tumor growth delay would occur in the brain metastasis setting. Of note, we consistently observed increased phosphorylation of HER2 in BT474-Gluc cells growing in the brain compared with the mammary fat pad (Fig. S10B). Although short-term (3 d) treatment with lapatinib significantly reduced HER2 phosphorylation in the brain, it could do so only to the level of that observed in the untreated mammary fat pad (Fig. S10B), and this effect disappeared after 15 d of treatment (Fig. S7B). We hypothesized that more pronounced HER2 inhibition with the combination of trastuzumab and lapatinib would be beneficial to these brain metastases with increased HER2 activation.

Consistent with the data from extracranial disease, we show a beneficial effect of combining trastuzumab and lapatinib compared with either monotherapy in our model of brain metastasis (Fig. 6A). Once again, we observed very little effect on tumor growth with either trastuzumab or lapatinib monotherapy, resulting in a 1.2-fold and 1.3-fold increase in median overall survival, respectively (a median of 31 d and 32 d, respectively, compared with 25 d in control-treated mice). However, the combination of two anti-HER2 agents shows a significant delay in tumor growth, leading to an increase in median survival 1.8-fold greater than in control-treated mice (44 d compared with 25 d; P < 0.001 compared with either monotherapy).

**Triple Combination of Trastuzumab, Lapatinib, and DC101 Produces a Significant Delay in Tumor Growth and Improvement in Mouse Survival in Our Model of HER2-Positive Breast Cancer Brain Metastasis.** We next examined the efficacy of combining two anti-HER2 therapies with DC101. Remarkably, we found a more dramatic tumor growth delay in mice treated with the triple combination compared with the combination treatment of any two drugs (Fig. 6B). This delay in tumor growth was accompanied by a dramatic increase in survival. In this particular experiment, control mice lived for a median of 22.5 d. Mice treated with the combination of trastuzumab and lapatinib lived 2.2-fold longer than control mice (a median of 49 d). Mice treated with the combination of trastuzumab and DC101 or lapatinib and DC101 lived 2.6-fold or 2.3-fold longer, respectively, than control-treated mice (a median of 58 d or 52.5 d, respectively). Meanwhile, mice treated with the triple combination lived fivefold longer than control-treated mice (a median of 113 d; P < 0.001 compared with any double-treatment group). It is important to note that two of nine mice receiving triple-combination therapy were euthanized after a significant loss in weight despite their brain metastatic tumors being relatively small. The reason for this is not clear, but toxicity associated with the three-drug regimen could be involved. Additionally, although the triple combination had a dramatic effect on overall survival and the growth of the brain metastases, all tumors eventually escaped from therapy.

**Discussion**

Studies of HER2-positive breast cancer metastasis to the brain have been severely hampered by the lack of clinically relevant laboratory models (11, 32–34). We describe here a metastatic model that faithfully replicates the clinical response to treatment: failure of a HER2-dependent tumor growing in the brain to respond to anti-HER2 therapy. Although our direct brain parenchymal tumor implantation model lacks several earlier steps in the metastatic cascade (35), it allows for consistent tumor volume at treatment initiation as well as real-time evaluation of therapeutic efficacy using imaging and blood surrogate marker evaluation. The blood surrogate marker correlated not only with tumor volume, as estimated with intravital microscopy using the transparent cranial window model, but with tumor volume measurements made by MRI, bioluminescence, and ex vivo multispectral imaging.

The reason for the differential tumor growth and response to anti-HER2 therapy when tumors are growing at these two different sites, as observed both in our laboratory model and in...
patients, has been elusive. Surprisingly, we witnessed increased phosphorylation of HER2 in tumor cells growing in the brain compared with the mammary fat pad. This increased HER2 activity could explain a mechanism of resistance, because the reduction in phosphorylation with lapatinib treatment reached a level only equal to that of untreated mammary fat pad tumors. In addition, the effect of lapatinib disappeared after 15 d of treatment. Increased HER2 activity in the brain metastatic tumors could also explain the reason why the combination of trastuzumab and lapatinib is more beneficial than monotherapy. Future studies will be needed to test whether more intense and/or prolonged HER2 inhibition is associated with better response.

Our data provide a strong rationale for the development of anti-VEGF therapy for brain metastases from HER2-positive breast cancer. Murine VEGFR2 blockade significantly reduced tumor MVD and increased tumor necrosis and survival on its own. In combination with HER2 inhibition, VEGFR2 blockade showed an even more dramatic response. Moreover, the combination of VEGFR2 blockade with dual inhibition of HER2 showed the most impressive benefits, although there is the potential for increased toxicity. Although the exact mechanism for the triple combination is not entirely clear, we propose that the antiangiogenic benefit seen with an anti-HER2 agent and VEGFR2 blockade is synergizing with the direct tumor cell cytotoxicity induced by two anti-HER2 agents. A clinical trial studying the efficacy of carboplatin and bevacizumab in progressive breast cancer brain metastasis is currently underway (http://clinicaltrials.gov identifier NCT01004172). In this trial, patients with HER2-positive disease will also be treated with HER2 inhibitors, which may provide clinical evidence for the approach presented here. Some of these drug combinations have been previously tested in patients with HER2-amplified breast cancer, and there is a growing body of experience with the use of anti-VEGF therapies for primary brain tumors (12, 36). Therefore, our findings could be readily translated into the clinic to increase survival in patients with brain metastatic HER2-amplified breast cancer whose systemic disease is responding to therapy. Moreover, future studies will be needed to determine how these brain metastases eventually escape from this triple-combination therapy.

A recent study by our group confirmed better growth inhibition in primary breast tumors with the standard dose of DC101 (40 mg/kg) compared with lower doses, owing to a superior antiangiogenic effect (37). However, the study also reported an added benefit of immune therapy only when combined with a low dose of DC101 (10 mg/kg), but not with the standard dose, mediated through “normalization” of the tumor vasculature. Future studies must be done to determine the optimal dose of anti-VEGFR2 therapy when combined with anti-HER2 therapies in the brain metastatic setting to achieve the greatest tumor growth delay.

Finally, it is important to mention that treatment of breast cancer brain metastases with anti-VEGF pathway therapy did
not result in an increased invasive phenotype, similar to what is observed after anti-VEGF therapy in primary brain tumors, especially after tumor hypoxia (36). Several theories could explain this phenomenon. Because BT474 cells are not invasive in nude mice, even when growing in the mammary fat pad, this phenomenon could be limited to this particular breast cancer cell line. To address this issue, future studies should investigate the invasiveness of a more aggressive breast cancer cell line when growing in the brain microenvironment after anti-VEGF therapy. Another reason for the lack of invasiveness in response to anti-VEGF therapy in our model could be because breast cancer cells do not disseminate after colonizing the brain. Could it be that the brain is the last metastatic stop for breast cancer cells, even in a hypoxic environment? If so, is this due to the unique brain microenvironment? Tracking the dissemination of breast cancer cells after growth in the brain would address this hypothesis.

Materials and Methods

Cell Lines, Infections, and Culture. BT474 cells were transduced with an expression cassette encoding Gluc and CFP separated by an internal ribosomal entry point (IREBP; Champion Dental Products) to 71-estradiol pellet (Innovative Research of America) the day before implantation of tumor cells and every 60 d thereafter. For the mammary fat pad model, 1 ± 10⁶ BT474-Gluc cells were suspended in a PBS and Matrigel Matrix High Concentration (BD Biosciences) at a 1:1 ratio before injection. For injection into the head, the brain of the mouse was fixed with a stereotactic apparatus and the skull over the left hemisphere of the brain was exposed via skin incision. Using a high-speed air-turbine drill (CH4201S; Champion Dental Products) with a burr tip size of 0.5 mm in diameter, three sides of a square (~2.5 mm in length, each side) were drilled through the skull until a bone flap became loose. Using blunt tweezers, the bone flap was pulled back, exposing the brain parenchyma. Roughly 10 μL of a BT474-Gluc cell suspension, at a concentration of 2.5 × 10⁶ cells/mL, was injected into the brain parenchyma using an insulin syringe. The bone flap was then placed back into position in the skull and sealed using histocompatible cyanoacrylate glue, and the skin atop the skull was sutured closed. All animal procedures were performed according to the guidelines of the Public Health Service Policy on Human Care of Laboratory Animals and in accordance with a protocol approved by the Institutional Animal Care and Use Committee of Massachusetts General Hospital.

Tumor Size Monitoring and Survival Analysis. Tumor size was measured twice a week by measuring the activity of secreted Gluc in the blood. Measurement of blood Gluc was performed as described previously (24). Briefly, blood was drawn from a slight nick in a tail vein of the mouse. Thirteen microliters of a mixture of PBS and Matrigel Matrix High Concentration (BD Biosciences) at a 1:1 ratio before injection. For injection into the head, the brain of the mouse was fixed with a stereotactic apparatus and the skull over the left hemisphere of the brain was exposed via skin incision. Using a high-speed air-turbine drill (CH4201S; Champion Dental Products) with a burr tip size of 0.5 mm in diameter, three sides of a square (~2.5 mm in length, each side) were drilled through the skull until a bone flap became loose. Using blunt tweezers, the bone flap was pulled back, exposing the brain parenchyma. Roughly 10 μL of a BT474-Gluc cell suspension, at a concentration of 2.5 × 10⁶ cells/mL, was injected into the brain parenchyma using an insulin syringe. The bone flap was then placed back into position in the skull and sealed using histocompatible cyanoacrylate glue, and the skin atop the skull was sutured closed. All animal procedures were performed according to the guidelines of the Public Health Service Policy on Human Care of Laboratory Animals and in accordance with a protocol approved by the Institutional Animal Care and Use Committee of Massachusetts General Hospital.

Quantification of Blood Vessel and Lecitin Perfusion Area. Before euthanasia, mice were injected with 2.5 μg of biotinylated-lactin per gram of body weight into the heart and allowed to circulate for 5 min. For immunofluorescence staining, mouse brains were fixed for 8h in 4% (v/v) formaldehyde in PBS at 4°C, followed by incubation in 30% sucrose in PBS at 4°C for 48 h and subsequent mounting in freezing media [Optional Cutting Temperature (OCT); Tissue-Tek]. Brains were sectioned every 20 μm. For endothelial cell identification, tissue sections were incubated overnight at 4°C with an anti-CD31 antibody (2.5 μg/mL, clone 2H8, Millipore), followed by incubation with a 1:200 dilution of an Alexa 649-conjugated anti-Armenian hamster antibody for 1 h at room temperature. For identification of perfused blood vessels, tissue sections were incubated for 1 h at room temperature with a 1:200 dilution of Alexa 546-conjugated streptavidin. Tissues were mounted with 4′-6-diamidino-2-phenylindole–containing mounting media (Vectorshield; Vectorlabs). Quantification of the stained area was performed using an in-house MATLAB program.

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Supporting Information

Kodack et al. 10.1073/pnas.1216078109

SI Materials and Methods

Cranial Window, Intravital Multiphoton Microscopy, and Tumor Volume Calculation. Cranial windows were implanted into nude mice as previously described (1). Intravital imaging of normal and angiogenic vessel morphology was performed using custom-built multiphoton laser scanning microscopy instrumentation. To contrast-enhance functional blood vessels, angiography was performed after i.v. injection of 0.1 mL rhodamine-dextran (10 mg/mL, 2,000 kDa). For intravital tumor volume calculation, BT474-Gaussia luciferase (Gluc) tumors were measured twice a week by intravital fluorescent microscopy. Using an in-house MATLAB (MathWorks) program, an ellipse was fitted to the cerulean fluorescent protein (tumor) signal and the short and long axes were calculated. Tumor volume was calculated based on a half-ellipse, using the following equation:

\[ \text{Tumor volume} = \left( \text{long axis} \times \text{short axis} \right) \times \left( \frac{\text{long axis} + \text{short axis}}{2} \right) \times \pi / 6 / 2 \]

MRI. MRI-based measurement of tumor growth was performed similar to the technique used by Kamoun et al. (2). All experiments were performed on a 9.4-T magnet (Magnex Scientific Ltd.) equipped with a gradient coil with a 60-mm inner diameter (Resonance Research) and interfaced with an MRI console (Bruker BioSpin). The gradient coil has a maximum strength of 1,500 mT/m and a rise time of 100 \( \mu \)s. Imaging was performed using a home-built mouse head birdcage coil. Mice were positioned on a custom-made mouse cradle and anesthetized with 1.5% (vol/vol) isoflurane in a 50:50 O\(_2\)/medical air mixture with total flow rate of 1,200 mL/min. T2-weighted RARE images were acquired to assess the tumor volume. The image acquisition parameters were as follows: echo time/repetition time = 10/3,000 ms; rapid acquisition relaxation enhanced factor = 16, four averages; field of view = 1.92 cm; matrix = 128 \( \times \) 128 (in-plane resolution = 150 \( \mu \)m); 0.5-mm slice thickness; and 11 image slices. Tumor volume was determined manually from the T2 hyperintense tumor regions of the brain.

Bioluminescence Imaging. Individual animals were anesthetized, and bioluminescence imaging (BLI) was performed immediately after retroorbital injection of coelenterazine (CTZ) solution (at a concentration of 4 mg/kg of body weight). An In Vivo Imaging System (IVIS Lumina II; Caliper Life Sciences) was used for bioluminescence image recording. The image acquisition time was 30 s at a precise 30-s time point after retroorbital injection. (Because Gluc has flash kinetics, imaging at a consistent time after CTZ injection is critical.) Postprocessing and quantification were performed using Living Image Software 3.0 (Caliper Life Sciences) coupled to the IVIS system. Image analysis was done similar to the technique used by Chung et al. (3).

Ex Vivo Multiphoton Fluorescent Imaging. The same mice used for the MRI analysis, bearing BT474-Gluc tumors treated with either control IgGs or the combination of trastuzumab and DC101, were killed immediately after the last imaging sequence. The entire brain was collected and fixed in 4% formaldehyde. After 24 h, the brain was sectioned into 1-mm slices. The tumor area was revealed in each section by spectral unmixing using the Maestro System (Cambridge Research and Instrumentation). Tumor volume was calculated by integration of the tumor areas, knowing that each section is 1 mm thick.

Histology and Immunostaining. Before embedding tissues in paraffin, mouse brains were fixed for 8 h in 4% formaldehyde in PBS at 4 \( ^\circ \)C, followed by washes in PBS and storage at 4 \( ^\circ \)C. Phosphorylation of human epidermal growth factor receptor-2 (HER2) was visualized with an anti–phospho-HER2 antibody (Tyr1221/Tyr1222, clone 6B12; Cell Signaling Technology) diluted at 1:520 in SignalStain Diluent (Cell Signaling Technology) and incubated overnight, followed by incubation with SignalStain Boost IHC Detection Reagent (HRP, rabbit; Cell Signaling Technology) for 1 h at room temperature (RT). Identification of proliferating cells was done using a Ki67 antibody (clone MIB-1; Dako) supplied at the working concentration and incubated for 1 h at RT, followed by incubation with mouse EnVision polymer (Dako) for 30 min at RT. Identification of apoptotic cells was accomplished using an antibody to cleaved caspase 3 (clone 5A1E; Cell Signaling Technology) diluted 1:50 and incubated overnight. Liquid DAB+ (Dako) was used to detect HRP-conjugated anti-mouse and antirabbit secondary antibodies, followed by mounting in Permount.

Western Blotting. Tumor tissues were resuspended in 1% Nonidet P-40 lysis buffer (15 mM Tris, 10 mM EDTA, 10 mM EGTA) with protease and phosphatase inhibitors, and incubated on ice for 10 min. After cell lysis, lysates were centrifuged at 14,000 rpm (~18,000 \( \times \)g) for 10 min at 4 \( ^\circ \)C. The protein concentration of the supernatant was measured before boiling in Laemmli buffer. Western blot analysis was conducted after separation by SDS/PAGE electrophoresis and transferred to nitrocellulose membranes. Immunoblotting was performed according to the antibody manufacturers’ recommendations. The anti–phospho-HER2, anti-phospho Akt, anti-Akt, anti-phospho-ERK, and anti-ERK antibodies were obtained from Cell Signaling Technologies, and the anti-HER2 antibody was obtained from Calbiochem (EMD Chemicals).

Fluorescence-Activated Cell Sorting and Quantification of Tumor Natural Killer Cells, and Depletion of Natural Killer Cells. For the quantification of natural killer (NK) cells within the tumor, tumor-bearing mice were intracardially perfused with PBS. Tumor tissues were harvested, minced, and digested at 37 \( ^\circ \)C for 60 min with complete DMEM containing collagenase type 1A (1,500 U/mL), hyaluronidase (1,000 IU/mL), and DNase (2 mg/mL). The digestion mixtures were filtered through 70-\( \mu \)m cell strainers. Single-cell suspensions were incubated with rat anti-mouse CD16/CD32 monoclonal antibodies and then stained, washed, and resuspended in cold buffer (1% BSA, 0.1% NaN\(_3\), in PBS). 7-Aminoactinomycin D (eBioscience) was added to the stained tubes (5 \( \mu \)L per tube) to exclude dead cells. Flow cytometry data were acquired on an LSRII flow cytometer (Becton Dickinson Biosciences) and analyzed with FACSDiva software (Becton Dickinson Biosciences). Appropriate fluorochrome-conjugated, isotype-matched control IgGs were used in all experiments. The anti-mouse antibodies CD45-phycocerythrin (PE)-Cy7 and NK1.1-PE (Becton Dickson Biosciences) were used. For the depletion of NK cells in mice, the NK1.1 antibody (clone PK136; BioXCell) was administered at a concentration of 4 mg/kg of body weight twice a week by i.p. injection. The control for the NK1.1 antibody was 4 mg/kg of body weight of a nonspecific mouse IgG (Jackson ImmunoResearch Laboratories, Inc.).

In Vitro Cell Growth Assay. A total of 3,000 BT474-Gluc cells were seeded in RPMI + 5% FBS in a 96-well plate and allowed to adhere overnight. Cells were treated the following day, and growth was...
measured 5 d after treatment addition. Cell growth was measured using the CellTiter-Glo Luminescent Cell Viability Assay (Promega).

**Statistical Analysis.** Data are expressed as the mean ± SEM unless otherwise noted. The principal statistical test was a one-way ANOVA, and Tukey’s posttest was used to compare all pairs of columns. A t test (two-tailed with unequal variance) was used when only two variables were present in the analysis. Significant differences in tumor growth were accomplished by determining the time it took (in days) to reach a specific blood Gluc activity (specific for each experiment and illustrated on the tumor growth curve graph) (Fig. 2A and B, Fig. 6A and B, and Fig. S8D). The survival curves were estimated using the Kaplan–Meier method, and the median survival day was used when determining statistical difference. Statistical significance is defined throughout the main text and figure legends. GraphPad Prism was used for all statistical analysis.


![Graph showing correlation of blood Gluc activity and tumor volume measurements](image)

**Fig S1.** Correlation of blood Gluc activity and tumor volume measurements. (A) Correlation of blood Gluc activity and tumor volume, measured using intravital microscopy. BT474-Gluc cells were injected into the brain parenchyma of mice with a cranial window. Tumors were imaged through the cranial window using a fluorescent microscope, and tumor length and width were obtained, from which volume was calculated. Blood was collected from the mice at the time of imaging. (Left) Blood Gluc activity is plotted against tumor volume, and linear regression analysis was performed using Prism software. Representative pictures are shown (Right), and inserted numbers correspond to the blood Gluc activity (Upper, blue) and the calculated tumor volume (Lower, yellow) (n = 8 mice, 44 separate measurements). Scale bar: 1 mm. (B) Correlation of blood Gluc activity and tumor volume, measured with MRI; representative images are shown in Fig. 3A (control-treated mice only; n = 4 mice, 9 separate measurements). (C) Correlation of blood Gluc activity and tumor volume, measured with bioluminescence imaging; representative images are shown in Fig. 3B (control-treated mice only; n = 5 mice). (D) Correlation of blood Gluc activity and tumor volume, measured by ex vivo multispectral fluorescent microscopy; representative images are shown in Fig. S2 (control-treated mice only; n = 5 mice).
**Fig. S2.** Representative images of control-treated (Upper) or trastuzumab and DC101-treated (Lower) brain sections. Spectral unmixing reveals tumor (ce-rulean fluorescent protein, blue) and normal (autofluorescence, green) tissue. Each section is 1 mm thick. Images were obtained with the Maestro System.

**Fig. S3.** Effect of the indicated treatments on cell proliferation and apoptosis. Immunohistochemical analysis of Ki67 (A) or cleaved caspase 3 (B) staining in tumor tissues harvested 15 d after the initiation of treatment and paraffin-embedded. (Scale bars: 50 μm.)
Fig. S4. Representative H&E-stained sections of BT474-Gluc brain metastatic tumor tissues used for the quantification of the necrotic area in Fig. 4. The borders of total tumor area and necrotic regions are highlighted in dotted blue and yellow lines, respectively. (Scale bars: 250 μm.)
Fig. S5. Effect of the indicated treatments on the tumor vasculature. (A) Representative images of lectin-positive endothelial cells in the viable tumor tissue 15 d after the start of treatment. (Scale bar: 100 μm.) (B and C) Effects of the indicated treatments on blood vessels in BT474-Gluc tumors 8 d after treatment initiation. (B) Percentage of CD31-positive blood vessel area within the viable tumor regions for each tumor was calculated using an in-house MATLAB program. (C) Before euthanasia, mice were injected with 2.5 μg of biotinylated-lectin per gram of body weight into the heart and allowed to circulate for 5 min. The area of lectin-positive blood vessels in the viable tumor regions for each tumor was calculated using an in-house MATLAB program. ***p < 0.001. DC101 vs. trastuzumab and DC101 or lapatinib and DC101, not statistically different.
Fig. S6. Effect and delivery of the indicated treatments. (A) Effect of trastuzumab and lapatinib treatment on HER2, AKT, and ERK phosphorylation in BT474-Gluc tumors growing in the brain 3 d after treatment initiation. Tumor tissues were collected 2 h after the last treatment. Western blotting illustrates levels of HER2, AKT, and ERK phosphorylation and corresponding total protein levels, as well as for GAPDH as a loading control. (B) Concentration of lapatinib in BT474-Gluc mammary fat pad and brain metastatic tumors (not statistically different). The dotted line marks the reported in vitro IC\textsubscript{50} of lapatinib for BT474 cells.
Fig. S7. Effect of anti-HER2 treatments alone or in combination with anti-VEGFR2 treatment on HER2 phosphorylation (p-HER2). Brain metastatic BT474-Gluc tumor tissues were harvested 15 d after the initiation of treatment with trastuzumab monotherapy and in combination with DC101 (A) or lapatinib monotherapy and in combination with DC101 (B). Tissues were either snap-frozen for protein lysis and Western blotting (Upper) or paraffin-embedded for immunohistochemistry (Lower). (Scale bars, 50 μm.)
Fig. S8. Role of NK cells in mammary fat pad and brain metastatic breast cancers treated with anti-HER2 and/or anti-VEGFR2 agents. (A) Effect of the depletion of NK cells on control-treated (dotted black line) and trastuzumab-treated (dotted red line) BT474-Gluc mammary fat pad growth. Nude mice were treated with the NK1.1 antibody at the time of treatment initiation. Tumor growth curves for control-treated (solid black line) and trastuzumab-treated (solid red line), NK cell-competent mice are shown (n = 6 mice). (B) FACS analysis of NK1.1-positive and CD45-positive cells within brain metastatic BT474-Gluc tumors 14 d after treatment initiation. *P < 0.05. (C) Effect of the NK1.1 antibody (clone PK136) on the presence of NK cells in the peripheral blood (Left) and spleen (Right). P < 0.01. (D) Effect of the depletion of NK cells on control-treated (black), trastuzumab-treated (red), DC101-treated (blue), and trastuzumab and DC101-treated (magenta) BT474-Gluc brain metastatic growth. Nude mice were treated with the NK1.1 antibody at the time of trastuzumab and/or DC101 treatment initiation. A tumor growth curve plot (Left) and Kaplan–Meier survival plot (Right) are shown for each treatment group. Tumor growth and survival curves for control-treated, NK cell-competent mice are shown in brown (n = 8–9 mice). **P < 0.01; ***P < 0.001.
Fig. S9. Effect of the combination of anti-HER2 and antimurine VEGFR2 agents on BT474-Gluc growth in vitro. BT474-Gluc cells were seeded in triplicate in a 96-well plate; allowed to adhere; and treated with the indicated concentrations of control IgG, DC101, trastuzumab, and/or lapatinib. After 5 d of treatment, cell growth was measured using the CellTiter-Glo Luminescent Cell Viability Assay. There was no significant difference between rat IgG and DC101 in the absence of an HER2 inhibitor, in the presence of trastuzumab, or in the presence of lapatinib. The graph is a representative of three separate experiments.

Fig. S10. Effect of two anti-HER2 therapies compared with one and the differences between tumor cells growing in the mammary fat pad or brain. (A) Effect of the combination of two anti-HER2 therapies on BT474-Gluc growth in vitro. BT474-Gluc cells were seeded in triplicate in a 96-well plate; allowed to adhere; and treated with the indicated concentrations of control IgG, trastuzumab, and/or lapatinib. After 5 d of treatment, cell growth was measured using the CellTiter-Glo Luminescent Cell Viability Assay. **P < 0.01; ***P < 0.001. The graph is representative of three separate experiments. (B) Effect of lapatinib on HER2, AKT, and ERK phosphorylation in BT474-Gluc tumors growing in the brain (Left) or mammary fat pad (MFP; Right). Tumor tissues were collected 3 d after treatment initiation, and Western blotting was performed to quantify phosphorylation (p) and corresponding total protein levels.