Corrections

**MEDICAL SCIENCES**

Correction for "Regulation of bone remodeling by vasopressin explains the bone loss in hyponatremia," by Roberto Tamma, Li Sun, Concetta Cuscito, Ping Lu, Michelangelo Corcelli, Jianhua Li, Graziana Colaianni, Surinder S. Moonga, Adriana Di Benedetto, Maria Grano, Silvia Colucci, Tony Yuen, Maria I. New, Alberta Zallone, and Mone Zaidi, which appeared in issue 46, November 12, 2013, of Proc Natl Acad Sci USA (110:18644–18649; first published October 28, 2013; 10.1073/pnas.1318257110).

The authors note that Fig. 1 appeared incorrectly. The corrected figure and its legend appear below.

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**Fig. 1.** Bone cells express Avprs. Immunofluorescence micrographs (A) and Western immunoblotting (B) show the expression of Avpr1α in osteoblasts and osteoclasts, and as a function of osteoblast (mineralization) and osteoclast (with Rankl) differentiation. The expression of Avp (ligand) and Avpr1α (receptor) in osteoblasts is regulated by 17β-estradiol, as determined by quantitative PCR (C) and Western immunoblotting (D). (Magnification: A, 63×.) Because Avp is a small peptide, its precursor neurophysin II is measured. Statistics: Student t test, P values shown compared with 0 h. Stimulation of Erk phosphorylation (p-Erk) as a function of total Erk (t-Erk) by Avp (10⁻⁸ M) in osteoclast precursors (preosteoclasts), osteoclasts (OC), and osteoblasts establishes functionality of the Avpr1α in the presence or absence of the receptor inhibitor SR49059 (10⁻⁸ M) (E). Western immunoblotting showing the expression of Avpr2 in preosteoclasts, OCs (F), and osteoblasts (G) isolated from Avpr1α−/− mice, as well as in MC3T3.E1 osteoblast precursors (G). Functionality of Avpr2 was confirmed by the demonstration that cells from Avpr1α−/− mice remained responsive to AVP in reducing the expression of osteoblast differentiation genes, namely Runx2, Osx, Bsp, Atf4, Opn, and Osteocalcin (quantitative PCR, P values shown) (H). Only relevant bands from Western blots are shown, with gaps introduced where empty lanes are excised to conserve space.

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PHARMACOLOGY

The authors note that an additional affiliation should be listed for Torsten Christ and Alberto Kaumann. The new affiliation should appear as Department of Experimental Pharmacology and Toxicology, University Medical Center Hamburg-Eppendorf. The corrected author and affiliation lines appear below. The online version has been corrected.

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PHYSIOLOGY

The authors note that the author name Timo Myöähänen should instead appear as Timo Myöähänen. The corrected author line appears below. The online version has been corrected.

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Regulation of bone remodeling by vasopressin explains the bone loss in hyponatremia

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Contributed by Maria I. New, September 27, 2013 (sent for review August 14, 2013)

Although hyponatremia is known to be associated with osteoporosis and a high fracture risk, the mechanism through which bone loss ensues has remained unclear. As hyponatremic patients have elevated circulating arginine-vasopressin (AVP) levels, we examined whether AVP can affect the skeleton directly as yet another component of the pituitary-bone axis. Here, we report that the two AVP receptors, Avpr1a and Avpr2, coupled to Erk activation, are expressed in osteoblasts and osteoclasts. AVP injected into wild-type mice enhanced and reduced, respectively, the formation of bone-resorbing osteoclasts and bone-forming osteoblasts. Conversely, the exposure of osteoblast precursors to Avpr1a or Avpr2 antagonists, namely SR49059 or ADAM, increased osteoblastogenesis, as did the genetic deletion of Avpr1a. In contrast, osteoclast formation and bone resorption were both reduced in Avpr1a−/− mice and in wild-type mice injected with SR49059. Collectively, the data not only establish a primary role for AVP signaling in bone mass regulation, but also call for further studies on the skeletal actions of AVP antagonists used commonly in hyponatremic patients.

Significance

In this study, we show that a primitive brain hormone, arginine vasopressin (AVP), which has hitherto been implicated in regulating water balance in mammals, has a function in skeletal homeostasis. Using genetically modified mice that are lacking one of the AVP receptors as well as pharmacologic inhibitors, we show that AVP negatively regulates osteoclasts (cells that form new bone) and stimulates osteoclasts (cells that remove old bone). Our findings explain the bone loss that is known to accompany low blood sodium levels in patients when AVP levels are high.


The authors declare no conflict of interest.

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Results
As part of our efforts in characterizing the posterior pituitary-bone axis (5), we have shown previously that functional Oxtrs and the ligand OT are expressed both in osteoclasts and osteoblasts (22, 23). Similarly, immunofluorescence microscopy and Western immunoblotting revealed Avpr1 expression in both cell types (Fig. 1A and B). Avpr1α expression in the osteoblast decreased between weeks 2 and 3 of mineralization, but in the osteoclast, remained stable over 8 d of differentiation (Fig. 1B). Because the expression of OT and Oxt were estrogen-sensitive (23), we studied the effect of 17β-estradiol on Avp and Avpr1α expression in osteoblasts. There was a significant increase in expression of Avp within 12 h of exposure to 17β-estradiol, but protein levels of its precursor, neurophysin II, remained unchanged (Fig. 1C and D). Note that, as AVP is 9-aa long, we have measured levels in culture supernatants using ELISA (401 ± 133 pg/mL) to demonstrate production in bone marrow. In contrast, Avpr1 mRNA and protein expression were increased at 12 and 24 h, respectively (Fig. 1C and D).

To confirm the functionality of the detected Avpr1α, we measured Erk activation in response to AVP in the presence or absence of an Avpr1α inhibitor, SR49059. In osteoclast precursors, osteoclasts and osteoblasts, AVP produced a robust increase in Erk phosphorylation without changing total Erk (Fig. 1E). These responses were attenuated, but not abolished by the Avpr1α inhibitor SR49059 (Fig. 1E). That Erk activation persisted despite maximally effective inhibitor concentrations suggested that another Avpr isoform, Avpr2, was also expressed in bone cells.

To specifically examine for Avpr2 expression, we used bone marrow cells from Avpr1a−/− mice. Avpr1a−/− osteoclast precursors and mature osteoclasts expressed Avpr2 as did the kidney, but brain and liver, which express Avpr1a, did not (Fig. 1F). However, Avpr2 was expressed at lower levels in osteoclasts than in the precursors, but was equally detectable in Avpr1a+/− and Avpr1a−/− cells (Fig. 1F). Similarly, calvarial osteoblasts from Avpr1−/− mice expressed Avpr2 in addition to Avpr1a (Fig. 1B and G). Functionality of osteoblastic Avpr2 was confirmed by studying the effect of AVP on the expression of differentiation genes, namely runt-related transcription factor 2 (Runx2), osterix (Ox), bone sialoprotein (Bsp), activating transcription factor 4 (Atf4), osteopontin (Opn), and Osteocalcin. Gene expression in Avpr1−/− osteoblasts was profoundly inhibited in response to AVP, suggesting that the Avpr isoform Avpr2 was also a functional regulator of osteoblastogenesis (Fig. 1H).

We then studied specifically the role of Avpr1α in regulating bone formation and bone mass. Injection of SR49059 into wild-type mice resulted in a dramatic increase in bone volume fraction, BV/TV (Fig. 2A). This finding was accompanied by a significant increase in bone formation parameters, notably mineralizing surface, mineral apposition rate (MAR), and bone formation rate, in double-labeled femur metaphyseal bone sections (Fig. 2B). Parallel studies with the double-labeled calvarial bone confirmed a substantial increase in MAR, which is normally measured in

![Figure 1](https://example.com/figure1.png)

**Fig. 1.** Bone cells express Avprs. Immunofluorescence microscopyographs (A) and Western immunoblotting (B) show the expression of Avpr1a in osteoblasts and osteoclasts, as well as a function of osteoblast (mineralization) and osteoclast (with Rankl) differentiation. The expression of Avp (ligand) and Avpr1a (receptor) in osteoblasts is regulated by 17β-estradiol, as determined by quantitative PCR (C) and Western immunoblotting (D). (Magnification: A, 63x.) Because Avp is a small peptide, its precursor neurophysin II is measured. Statistics: Student t test. P values shown compared with 0 h. Stimulation of Erk phosphorylation (p-Erk) as a function of total Erk (t-Erk) by Avp (10−8 M) in osteoclast precursors (preosteoclasts), osteoclasts (OC) and mature osteoclasts (M) indicated by 17β-estradiol, as determined by Western immunoblotting (E). Western immunoblotting showing the expression of Avpr2 in preosteoclasts, osteoclasts (OC), and mature osteoclasts (M) isolated from Avpr1−/− mice, as well as in MC3T3-E1 osteoblast precursors (G). Functionality of Avpr2 was confirmed by the demonstration that cells from Avpr1a−/− mice remained responsive to AVP in reducing the expression of osteoblast differentiation genes, namely Runx2, OX, Bsp, Atf4, Opn, and Osteocalcin. (H) Western immunoblotting showing the expression of Avpr2 in osteoblast precursors, mature osteoclasts (OC), and osteoblasts (OB) isolated from Avpr1−/− mice.
calvaria (Fig. 2C). Additionally, resorption parameters, including osteoclast surfaces (N.oc/B.Pm), as well as serum C-telopeptide levels were significantly reduced in inhibitor-treated mice compared with controls (Fig. 2D).

These results from pharmacologic Avpr1α inhibition by SR49059 were closely mimicked by the genetic deletion of the Avpr1α. There was a highly significant increase in bone volume, BV/TV, observed on von Kossa staining of vertebral bone, as well as in femur metaphyseal bone (Figs. 2E and F). Notably, there were accompanying increases in trabecular thickness (Tb.Th) and trabecular number (Tb.N), and decreases in trabecular spacing (Tb.Sp). Significant increases and decreases, respectively, in osteoblast (N.ob/BV) and osteoclast (N.oc/BPm) number were noted (Fig. 2F and G). The increases in bone mass were further confirmed by 3D microtomography of femur metaphyseal trabecular bone (Fig. 2H), as well as in the femur diaphysis, where cortical thickness (Ct.Th) was significantly increased (Fig. 2I). Overall, the findings suggest a prominent role for Avpr1α in bone remodeling and bone mass regulation.

Consistent with the increase in bone formation noted in vivo, Avpr1α−/− bone marrow stromal cells showed a significant elevation in colony-forming-fibroblastoid (Cfu-f) at day 10 (Fig. 3A), consistent with the increase in bone formation noted in vivo, Avpr1α−/− bone marrow stromal cells showed a significant elevation in colony-forming-fibroblastoid (Cfu-f) at day 10 (Fig. 3A),
Fig. 3. Avpr1α and Avpr2 regulate osteoblastogenesis. Genetic deletion of Avpr1α in Avpr1α−/− mice causes a dramatic increase in osteoblastogenesis, measured by alkaline phosphatase-positive Cfu-f (10 d) (A) and von Kossa-positive Cfu-ob (2 and 4 wk) (B). For Cfu-f and Cfu-ob assays, bone marrow stromal cells were grown in α-MEM and 15% (vol/vol) FCS with ascorbate-2-phosphate (1 mM) (Materials and Methods). (C) The enhanced osteoblastogenesis was confirmed by a strong up-regulation of differentiation genes (quantitative PCR), namely Bsp, Bmp2, Osteocalcin, Runx2, Osx, and Atf4. (D) Effect of inhibitors of Avpr1α (SR49059) or Avpr2 (ADAM) on Cfu-f (i, 10 d) and Cfu-ob (ii, 4 wk) formation. (E) Ex vivo Cfu-f formation (10 d) in mice injected with AVP (4 μg per mouse, twice, 6-wk-old, killed 12 h after second injection). Cfu-ob formation (F) and expression of Runx2, Atf4, Bmp2, Opn, Bsp, Lrp6, Sost, and Ocn (G) at 2 wk in the presence of AVP in vitro. Statistics: Unpaired Student t test, corrected by Bonferroni’s; *P < 0.05; **P < 0.01, comparison between Avpr1α+/+ and Avpr1α−/− cells or betweenCtl and AVP. (Magnification: all stained images, 1×.)
with colony forming-osteoblastoid (Cfu-ob) increasing progressively until 4 wk of culture (Fig. 3B). This pronounced increase in osteoblast differentiation was demonstrated further by measuring the expression of Bmp, bone morphogenetic protein-2 (Bmp2), Osteocalcin, Runx2, Oss, and Atf4. Although the expression of all of these genes increased as a function of osteoblast differentiation in wild-type mice, this phenomenon was exaggerated significantly in Avpr1α−/− cultures (Fig. 3C). Interestingly, with the exception of Runx2, these increases contrast the inhibitory effects of Ostr deletion, reported previously (4). We suggest that Oxtr and Avpr have opposing actions on osteoblastogenesis, although the deletion of either receptor elevates Runx2 expression (4). The significance of this discordance remains unknown. Furthermore, gain-of-function studies showed that the injection of AVP into mice or exposure to AVP in vitro inhibited osteoblast formation (Fig. 3E and F) and gene expression (Fig. 3G).

A functional role of Avpr2 in osteoblastogenesis was also explored using Avpr1α−/− cells and the inhibitors of the two receptors Avpr1α and Avpr2. Exposure of wild-type cells to the Avpr1α inhibitor SR49059 or deletion of Avpr1α resulted in enhanced Cfu-f and Cfu-ob formation (Fig. 3D), whereas no further increases were noted when Avpr1α cells were treated with SR49059. However, a dramatic increase in Cfu-f was observed when Avpr1α−/− cells were exposed to the Avpr inhibitor ADAM (Fig. 3D). This suggests that both Avpr isoforms work in tandem to inhibit osteoblastogenesis.

We next probed the effects of Avpr activation on osteoclasts in loss- and gain-of-function studies using Avpr1α−/− cells, and by administering either the Avpr antagonist SR49059 or AVP. Bone marrow cell or spleen cultures from Avpr1α−/− mice in the presence of Rankl showed a dramatic reduction in osteoclast formation compared with wild-type mice (Fig. 4A). This was accompanied by reduced expression of certain osteoclast genes, namely Cfms, Rank, Nfatc1, and Integrin β3 (Int β3) in ex vivo bone marrow cell cultures (Fig. 4B). When osteoclasts were seeded on hydroxyapatite-coated coverslips, there was a striking attenuation in bone resorption, as evaluated by von Kossa staining (Fig. 4C). These findings were consistent with the significant decrease in osteoclast surface, N.ocr/B.Pm, noted when wild-type mice were injected with SR49059 (Fig. 2C). Collectively, the data suggest that Avp is a proresorptive hormone in mice, as the loss of Avpr signaling inhibits osteoclast formation and bone resorption.

We tested this putative proresorptive action directly by injecting AVP into wild-type mice. A profound increase in osteoclastogenesis was observed ex vivo (Fig. 4D). This increase was accompanied by a significant up-regulation in bone marrow cell cultures of osteoclast differentiation genes, namely Cfms, Rank, Trap, Int β3, Calcr, Csk, Clc7, and Atpase6 (Fig. 4E). Overall, these findings suggest that Avp enhances bone resorption by stimulating the formation and function of osteoclasts, although we cannot demonstrate a direct effect of AVP on bone in mice for technical reasons. These actions are in stark contrast to OT, which stimulates osteoclastogenesis, but inhibits the function of mature osteoclasts, with little change in overall bone resorption (4).

**Discussion**

The G protein-coupled receptor superfamily of receptors, to which the Avprs belong, is much more widely distributed than originally thought. The clearest example is ACTH, which is part of a widely distributed melanocortin receptor system that regulates a diverse array of functions, including pigment production, apatite, and sexual function (24, 25). These receptors are evolutionarily primitive in that lower organisms, such as coelenterates, express, for example, the TSH receptor (24). In fish, the TSH receptor is not only present in the thyroid gland, but also in heart muscle and brain. Oxtrs also have a widespread distribution in the central nervous system to control apatite, sexual function, and behavior (26). Similarly, in lower vertebrates, such as birds, amphibians, and fish, Avpr1α, expressed in the central nervous system, determines sexual and reproductive behavior (27). In mammals, Avpr1α regulates the vasculature, Avpr1β is involved in aggression, social memory, and stress responsiveness, and Avpr2, localized primarily to the kidney, controls body water (27). Here, we show in parallel with Oxtrs, that Avpr1α and Avpr2 are expressed in osteoblasts and osteoclasts. However,

![Loss of function](image)

**Fig. 4.** Avpr1α regulates osteoclast formation and function. Genetic deletion of Avpr1α in Avpr1α−/− mice causes a dramatic reduction in osteoclastogenesis in bone marrow and spleen cell cultures (A), which is accompanied by a reduction in the expression (quantitative PCR) of differentiation genes, namely Cfms, Rank, Nfatc1, and Int β3 (B), and by a reduction in bone resorption by mature osteoclasts plated on dentine slices (von Kossa-stained) (C). Gain-of-function experiments evaluating the stimulatory effect of AVP injections (4 µg per mouse, twice, 6-wk-old mice killed 12 h after second injection) on TRAP-positive osteoclast formation in bone marrow cell cultures treated with Rankl (30 ng/mL) (D), and on osteoclast genes, namely Cfms, Rank, Trap, Int β3, Calcr, Csk, Clc7, and Atpase6 (E). Statistics by Unpaired Student t test; **P < 0.01 vs. vehicle (−), no AVP) treated at 3 and 7 d of culture. (Magnification: A, Upper, and D, 10×; A, Lower, and C, 2×.)
Avpr action on the skeleton, which is to inhibit bone formation, is opposed to Otrx's anabolic action.

More fascinating, however, is that the ligands for the aforementioned G protein-coupled receptors, the so-called "pituitary hormones," have an equally wide cellular distribution. ACTH is produced by macrophages, so that its anabolic action in skeletal homeostasis may in fact be exerted locally and independently of its pituitary origin (28). Macrophages also produce a TSHβ-variant, which is not suppressed by thyroid hormone and may help protect the skeleton from the effects of pituitary TSH deficiency in hyperthyroidism (6). In contrast, FSHβ has not been detected in bone cells, although OT is produced in abundance by osteoclasts and osteoblasts (22, 23). Furthermore, Otrx and OT expression is upregulated in osteoclasts, particularly in osteoclasts, is regulated by estrogen (22, 23). Although estrogen stimulates bone formation, in part, by increasing Otrx/OT expression, it is likely also to enhance the expression of Avp and Avpr1a, which in turn will oppose the feed-forward loop that we have predicted for Otrx/OT interactions (23). Thus, our demonstration of regulated Avp/Avpr expression in bone not only adds to the emerging complexity of the pituitary-bone axis, but also suggests that short-range circuits have evolved that use locally produced pituitary hormones to conduct skeletal surveillance.

The findings shed new light on the long-standing dogma that has centered round the mechanisms underlying the osteoporosis that accompanies hyponatremia (10–16). It is quite possible that the bone loss seen in the patient with SIADH had arisen from a 30-fold elevation in serum AVP levels, although the accompanying high aldosterone may have contributed (19). Hyperaldosteronism has been linked to bone loss in rodents (29, 30). The use of spirinolactone in patients with secondary hyperaldosteronism because of heart failure has also been shown to reduce fracture risk (31). Nonetheless, the complementary cell biological, pharmacological, and molecular data we provide compels us to argue for a role for AVP in causing hyponatremia-induced bone loss.

Although we have yet to prove this hypothesis in people, our study does offer a strong rationale for the routine evaluation of skeletal health in patients with chronic hyponatremia, as well as for the potential of using an antiresorptive or anabolic agent for osteoprotection. However, as Avpr2 is expressed osteoblasts and osteoclasts, and is active functionally in regulating bone mass, it is possible that highly selective Avpr2 inhibitors, such as tolvaptan (32), when used as aquaretics in the therapy of chronic hyponatremia, could in fact themselves offer osteoprotection. Our data prompt future clinical studies to examine the skeletal actions of Avpr2 inhibitors in people receiving them.

Materials and Methods

Avpr1a−/− mice, originally on a C57BL/6J × 129Sv background, were crossed for 10 generations to eliminate the mixed background. For histomorphometry, the mice were injected with xylene orange (90 mg/kg, i.p.) and calcine (15 mg/kg, i.p.) 10 and 2 d before being killed. Lumbar vertebras, femora, and calvaria were dissected, processed, and analyzed for bone formation and resorption parameters, as described previously (7). Additionally, 10-μm longitudinal sections were stained by von Kossa for bone volume measurement, and 12-μm calvarial sections were analyzed for MAR. Blood (~1 ml) was collected for measurement of T-celtopeside levels (ELISA), and 3D microtomography was performed as described previously (7). The protocols were approved by the Institutional Animal Care and Use Committee, Faculty of Medicine, University of Bari and Main Scool of Medicine.

Mouse cells were isolated from Avpr1a−/− and wild-type mouse bone marrow-adherent fraction of 5- to 6-d-old pups. Cells were cultured in phenol red free α-MEM (Gibco) with 15% (vol/vol) charcoal-stripped FCS in the presence of ascorbate-2-phosphosphate (1 mM, Sigma) for mRNA and protein analysis and von Kossa staining. To detect Avpr1a expression, immunofluorescence was performed using a rabbit anti-Avpr1a (Santa Cruz) and anti-αv β3 conjugated secondary antibodies (Chemicon International) associated with 60 μg/ml fluorescein-labeled phalloidin (Sigma Aldrich). Western blotting was performed using rabbit polyclonal anti-Avpr1a and anti-Avpr2, goat polyclonal anti-neuropilin II, mouse monoclonal pERK, rabbit polyclonal tERK (Santa Cruz), and IRDye-labeled secondary antibodies (680/800CW) (LI-COR Biosciences). For immunoblotting, Li-Cor Odyssey infrared imaging system was used. Quantitative PCR was performed as described previously (4).

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Tamma et al.