Corrections

PHARMACOLOGY

The authors note that an additional affiliation should be listed for Emanuela Galliera. This author’s affiliations should appear as “Department of Biomedical, Surgical and Dental Sciences, University of Milan, I-20133 Milan, Italy; and Istituto di Ricerca e Cura a Carattere Scientifico (IRCCS) Galeazzi Orthopaedic Institute, I-20161 Milan, Italy.” The corrected author and affiliation lines appear below. The online version has been corrected.


Neuroscience

The authors note that the author name Benoit Lebonté should instead appear as Benoit Labonté. The corrected author line appears below. The online version has been corrected.

Georgia E. Hodes, Madeline L. Pfau, Marylene Leboeuf, Sam A. Golden, Daniel J. Christoffel, Dana Bregman, Nicole Rebusi, Mitra Heshmati, Hossein Aleyasin, Brandon L. Warren, Benoit Labonté, Sarah Horn, Kyle A. Lapidus, Viktorija Stelzhammer, Erik H. F. Wong, Sabine Bahn, Vaishnav Krishnan, Carlos A. Bolaños-Guzman, James W. Murrough, Miriam Merad, and Scott J. Russo

Applied Physical Sciences

The authors note that on page 19269, right column, fifth full paragraph, line 4, “200 ms” should instead appear as “200 μs.”

Houxun Miao, Lei Chen, Eric E. Bennett, Nick M. Adamo, Andrew A. Gomella, Alexa M. DeLuca, Ajay Patel, Nicole Y. Morgan, and Han Wen

www.pnas.org/cgi/doi/10.1073/pnas.1423575112

www.pnas.org/cgi/doi/10.1073/pnas.1423579112

www.pnas.org/cgi/doi/10.1073/pnas.1422166112
Individual differences in the peripheral immune system promote resilience versus susceptibility to social stress

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Edited by Bruce S. McEwen, The Rockefeller University, New York, NY, and approved September 25, 2014 (received for review August 7, 2014)

Depression and anxiety disorders are associated with increased release of peripheral cytokines; however, their functional relevance remains unknown. Using a social stress model in mice, we find preexisting individual differences in the sensitivity of the peripheral immune system that predict and promote vulnerability to social stress. Cytokine profiles were obtained 20 min after the first social stress exposure. Of the cytokines regulated by stress, IL-6 was most highly up-regulated only in mice that ultimately developed a susceptible behavioral phenotype following a subsequent chronic stress, and levels remained elevated for at least 1 mo. We confirmed a similar elevation of serum IL-6 in two separate cohorts of patients with treatment-resistant major depressive disorder. Before any physical contact in mice, we observed individual differences in IL-6 levels from ex vivo stimulated leukocytes that predict susceptibility versus resilience to a subsequent stressor. To shift the sensitivity of the peripheral immune system to a pro- or antidepressant state, bone marrow (BM) chimeras were generated by transplanting hematopoietic progenitor cells from stress-susceptible mice releasing high IL-6 or from IL-6 knockout (IL-6−/−) mice. Stress-susceptible BM chimeras exhibited increased social avoidance behavior after exposure to either subthreshold repeated social defeat stress (RSDS) or a purely emotional stressor termed witness defeat. IL-6−/− BM chimeric and IL-6−/− mice, as well as those treated with a systemic IL-6 monoclonal antibody, were resilient to social stress. These data establish that preexisting differences in stress-response IL-6 release from BM-derived leukocytes functionally contribute to social stress-induced behavioral abnormalities.

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uman studies indicate that psychosocial stressors increase peripheral cytokine production, a potentially important factor in the development of depression or anxiety (1–6). Subsets of patients with major depressive disorder (MDD) and posttraumatic stress disorder have higher levels of multiple inflammatory markers, including the cytokine interleukin 6 (IL-6) (3, 4, 6–8), which meta-analyses indicate is consistently elevated across studies (2, 9–11). Although systemic injection of proinflammatory cytokines induces “sickness behavior” reminiscent of depressive symptoms (12), a causal relationship between peripherally derived cytokines and stress-related disorders awaits confirmation. To directly address whether systemic inflammation functionally contributes to stress vulnerability, we used two social stress paradigms: repeated social defeat stress (RSDS) (13, 14) and a purely emotional stressor (witness defeat) (15). As in humans (16, 17), chronic social subordinations in mice lead to depression-like behavior, including social avoidance, in a subset of mice termed susceptible, whereas resilient mice resist the development of such behavior (18–20). We hypothesize that preexisting differences in the sensitivity of an individual’s peripheral immune system dictate their subsequent vulnerability or resilience to social stress.

Results
Leukocyte-Derived IL-6 Is Predictive of Susceptibility Versus Resilience to Stress. We examined cytokine and chemokine profiles 20 min after the first exposure to an aggressor in the RSDS paradigm (Table 1 and Fig. S1A). Although acute social stress regulated a number of cytokines and chemokines, IL-6 was the only cytokine significantly elevated in animals that later developed a susceptible phenotype compared with both control and resilient mice. Furthermore, IL-6 levels strongly negatively correlated with social interaction behavior following subsequent RSDS (Fig. S1B). IL-1β was also elevated in susceptible mice compared with resilient mice, however, neither differed from controls. The anti-inflammatory cytokine IL-10 and the chemokines chemokine (C-X-C motif) ligand 1 (CXCL1) and chemokine (C-C motif) ligand 2 (CCL2) were similarly elevated in susceptible and resilient animals compared with controls, indicating a general effect of defeat (Table 1). IL-9 was not significantly regulated by stress but did correlate with subsequent social avoidance behavior (Fig. S1C).

Significance

Depression and anxiety have been linked to increased inflammation. However, we do not know if inflammatory status pre-dates onset of disease or whether it contributes to depression symptomatology. We report preexisting individual differences in the peripheral immune system that predict and promote stress susceptibility. Replacing a stress-naive animal’s peripheral immune system with that of a stressed animal increases susceptibility to social stress including repeated social defeat stress (RSDS) and witness defeat (a purely emotional form of social stress). Depleting the cytokine IL-6 from the whole body or just from leukocytes promotes resilience, as does sequestering IL-6 outside of the brain. These studies demonstrate that the emotional response to stress can be generated or blocked in the periphery, and offer a potential new form of treatment for stress disorders.


Conflict of interest statement: This work was supported by a research grant from Janssen Pharmaceuticals.

This is a PNAS Direct Submission.

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This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1415191111/-/DCSupplemental.
Table 1. Cytokines and chemokines regulated by stress 20 min after the first defeat

<table>
<thead>
<tr>
<th>Cytokine</th>
<th>IL-1β pg/mL</th>
<th>IL-6 pg/mL</th>
<th>IL-9 pg/mL</th>
<th>IL-10 pg/mL</th>
<th>Cxcl1/KC pg/mL</th>
<th>Ccl2/Mcp1 pg/mL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>8.73 ± 1.09</td>
<td>3.36b ± 0.96</td>
<td>131.2 ± 17.7</td>
<td>4.24bc ± 0.37</td>
<td>95.25bc ± 5.32</td>
<td>20.58bc ± 1.88</td>
</tr>
<tr>
<td>Susceptible</td>
<td>11.75 ± 2.47</td>
<td>92.23 ± 8.31</td>
<td>160.8 ± 9.04</td>
<td>10.37 ± 2.05</td>
<td>575.6 ± 170.8</td>
<td>40.67 ± 7.44</td>
</tr>
<tr>
<td>Resilient</td>
<td>6.49b ± 0.68</td>
<td>10.83a ± 2.78</td>
<td>130.5 ± 10.1</td>
<td>8.45 ± 1.34</td>
<td>547.1b ± 161.5</td>
<td>49.44a ± 6.93</td>
</tr>
</tbody>
</table>

Animals were behaviorally phenotyped 10 d after RSDS by measuring social interaction (Fig. S1A). Univariate ANOVA analysis was used to determine significant differences in circulating cytokine levels between control, susceptible, and resilient mice. Correlations to the SI ratio were examined in animals that underwent RSDS. Subjects with nondetectable levels of cytokines/chemokines or levels that varied more than 2 SD from the mean were excluded from analysis. Data are displayed as mean ± SEM. Correlations are listed in row R, and sample numbers are listed in row N.

*aSignificant difference from control.

bSignificant difference from susceptible.

cSignificant difference from resilient.

Given the 27-fold change in IL-6 detected in susceptible animals, we next examined an in vivo time course of plasma IL-6 levels before and throughout exposure to RSDS in a second group of animals. There were no basal differences in IL-6 before RSDS, however we again observed that susceptible animals had significantly higher serum IL-6 within 20 min of their first defeat and levels remained elevated 48 h after the last defeat (Fig. 1A and Figs. S2A and S3A). Because baseline levels of IL-6 were low, we confirmed that there were indeed no basal group differences via a high-sensitivity ELISA in a separate cohort of mice (Fig. S4A and B).

To validate that the increased levels of IL-6 found in susceptible mice were clinically relevant, we examined IL-6 in serum from patients with treatment-resistant MDD using a high-sensitivity human IL-6 ELISA. Severity of depression was quantified using the Quick Inventory of Depressive Symptomatology (QIDS) scale (Fig. 1B). Patients with MDD had higher serum IL-6 than healthy controls (Fig. 1C and Table S1). In a second cohort of patients with treatment-resistant MDD, we examined the effects of antidepressant treatment on circulating levels of IL-6. In this cohort, severity of depression was quantified using Hamilton Depression Scale scores (Table S1). Again levels of IL-6 were elevated in patients with treatment-resistant MDD, and standard antidepressant treatment did not lower circulating levels of IL-6.

IL-6 levels were also examined in mice that experienced 10 d of RSDS followed by 35 d of Imipramine treatment (Fig. 1D and Figs. S4 and S5). Alveolar macrophages were elevated in animals that developed a susceptible phenotype (Fig. 1E and Table S3). The number of leukocytes negatively correlated with the social interaction ratio (Fig. 2B). To determine the specific type of leukocyte that predicts susceptibility, we collected whole blood before RSDS and separated leukocytes via flow cytometry in a separate cohort of mice. We found that before stress exposure, monocytes were elevated in animals that developed a susceptible phenotype after stress exposure (Fig. S5A) and negatively correlated with the social interaction ratio (Fig. S5B and C).

Because of the physical nature of RSDS, we performed physical examinations of animals. There were no differences in the number of wounds and no correlation between number of wounds and social-interaction (SI) ratio (Fig. S4C and D), suggesting that individual differences in susceptibility to RSDS were not due to physical injury. Because IL-6 is involved with the fever response (21), we examined core body temperature after 10 d of RSDS as general sickness behavior could alter social interaction (22). There were no significant effects of RSDS on core body temperature (Fig. S4E). Furthermore, RSDS increased corticosterone levels to the same extent in both susceptible and resilient animals (Fig. S4F and G). These data suggest that both groups received similar levels of trauma-related stress and further dissociate general hypothalamic pituitary adrenal stress responses from stress-induced levels of IL-6.

To determine whether immune differences in stress-naïve mice can predict development of susceptible versus resilient responses to a subsequent RSDS, we conducted an ex vivo leukocyte immune challenge (Fig. S2B). Whole blood was collected by submandibular bleed 4 d before the first physical interaction with an aggressor. Leukocytes were isolated and counted. Mice that later exhibited a susceptible phenotype following RSDS had more circulating leukocytes than those that exhibited a resilient phenotype (Fig. 2A and B). To measure IL-6 release to an ex vivo immune challenge, leukocytes were collected before RSDS, plated at equal confluence, and stimulated with the toll-like receptor 4 agonist, lipopolysaccharide (LPS), for 24 h. Leukocytes from animals that later exhibited a susceptible behavioral phenotype released more IL-6 in response to LPS before stress than those that exhibited a resilient phenotype (Fig. 2C). Stimulated IL-6 release also negatively correlated with the social interaction ratio (Fig. 2D). These studies confirm the existing individual differences in leukocyte numbers as well as leukocyte response to an immune challenge before RSDS, suggesting that dysregulated immune physiology and concomitant increased IL-6 release are a risk factor for stress susceptibility.

Peripheral Immune System Controls Behavioral Susceptibility. To examine the functional contribution of increased immune activation to stress, animals were reconstituted with bone marrow (BM) hematopoietic progenitors isolated from susceptible or resilient mice.
Inhibition of Peripheral IL-6 Promotes Behavioral Resilience. To determine whether alterations in peripheral IL-6 contribute to individual differences in susceptibility to RSDS, we generated IL-6−/− BM chimeras as described (Fig. S2D). Once again we confirmed ~70% of viable donor leukocyte chimerism and found no differences in proportions of donor-derived leukocytes between IL-6−/− and WT BM chimeras (Fig. 4 and Fig. S7D). This ruled out nonspecific effects of IL-6 deletion on stem cell engraftment and differentiation into mature leukocytes. The WT microglia population was left largely intact (Fig. 3C and Fig. S7C). This is in line with previous studies using lead shielding to protect the CNS from radiation (23).

To determine the behavioral consequences of peripheral immune transplantation, stress-susceptible and control BM chimeric mice were subjected to a physical subthreshold defeat that does not induce social avoidance in controls but can reveal a susceptible phenotype if an animal’s stress threshold is shifted by the experimental manipulation (24). Stress-susceptible BM chimeras displayed increased social avoidance following subthreshold defeat compared with controls (Fig. 3D). There were no differences in measures of anxiety-associated exploratory behavior between the two groups as indicated by the elevated plus maze (Table S2). To determine whether the peripheral immune system impacts the behavioral response to a purely emotional stressor (with no physical component), we generated a separate group of susceptible BM chimeric mice and allowed them to witness another mouse undergoing RSDS using a modified subthreshold protocol. Susceptible BM chimeras showed a greater social avoidance behavior compared with control BM chimeras following this subthreshold witness stress (Fig. 3E).

Fig. 1. Individual differences in the peripheral immune system predict behavioral responses to RSDS. (A) A within-subjects time course of plasma levels of IL-6 indicated a significant interaction between time point of blood draw and behavioral phenotype (F_{1, 19} = 10.83, P < 0.001, control = 9, susceptible = 7, resilient = 6). Post-hoc analysis indicated no significant differences in IL-6 levels at baseline (P > 0.05). Twenty minutes after the first defeat, animals that developed a susceptible phenotype had higher circulating levels of IL-6 than control or resilient mice (P < 0.001). Forty-eight hours after the last defeat, susceptible mice had higher levels of IL-6 than resilient or control mice (P < 0.001). (B) Patients with a diagnosis of treatment-resistant MDD (n = 19) scored higher on the QIDS than healthy controls (n = 18) (t_{25} = 15.68, P < 0.0001, two tailed). (C) Patients with treatment-resistant MDD had higher circulating levels of IL-6 than healthy controls (t_{25} = 3.17, P < 0.01, two tailed). (D) Thirty-five days after the last defeat, susceptible mice injected with saline (n = 7) had basal elevations in serum levels of IL-6 compared with controls (saline, n = 13, Imipramine, n = 13) and resilient (n = 6) mice (F_{2, 45} = 3.66, P < 0.05) but not susceptible mice injected with Imipramine (n = 11, P < 0.05). (E) Twenty-four hours after the last day of CVS, mice showed significant elevations of IL-6 (t_{24} = 2.48, P < 0.05, two tailed). (F) Mice exposed to the nonphysical social stress of witness defeat (witness = 10, control = 11) demonstrated elevations in IL-6 30 d after the last stressor, when serum levels were measured immediately after the social interaction test (t_{23} = 3.1, P < 0.01, two tailed). Bar graphs display mean ± SEM. # denotes a significant interaction. * denotes a significant difference in phenotype. For t tests, * denotes a significant difference between means.

control mice (Fig. S2C). To exclusively target peripheral immune cells and spare central nervous system (CNS) microglia during whole body irradiation of CD45.2+ C57BL/6 mice, we used lead shields to cover the animals’ heads (Fig. S6D). For the susceptible donors, CD45.1+ C57BL/6 mice were selected based on high release properties of IL-6 in response to an ex vivo leukocyte challenge with LPS and the development of social avoidance following RSDS (Fig. S7A). Control donors did not undergo RSDS and showed low IL-6 release properties to LPS ex vivo. BM hematopoietic progenitor cells from susceptible or control mice were transplanted into the irradiated CD45.2+ C57BL/6 recipients, and animals were given 5–6 wk to allow for full donor hematopoietic cell reconstitution before exposure to social stress. Blood was collected at the end of each experiment to determine donor chimerism. Approximately 70% of the viable leukocytes derived from donor progenitor cells, although proportions of donor-derived leukocytes did not differ between susceptible and control donors (Fig. 3.4 and Fig. S7B). After transplant, stress-susceptible BM chimeras overall had more circulating leukocytes compared with wild-type control chimeras (Fig. 3B). Examination of brain tissue indicated that proliferation of cells in neurogenic brain regions known to regulate antidepressant responses was not affected by irradiation (Fig. S6 B and C). Further, >98% of the microglia remained of host origin, suggesting that their resident

Fig. 2. Leukocyte profiles before stress. (A) Before RSDS, mice that displayed a susceptible phenotype at the end of RSDS had more circulating leukocytes than mice that displayed a resilient phenotype (F_{2, 36} = 3.03, P < 0.01, two tailed; susceptible, n = 25, resilient, n = 13). (B) Prestress circulating leukocytes before social defeat stress negatively correlated with the social interaction ratio following RSDS (r = −0.44, P < 0.01). (C) Before RSDS, leukocytes from susceptible animals released more IL-6 than from resilient animals when stimulated with LPS (F_{1, 6} = 15, P < 0.001; susceptible, n = 25, resilient, n = 13). (D) Prestress levels of IL-6 released by leukocytes in response to LPS negatively correlated with the social interaction ratio following RSDS (r = −0.48, P < 0.01). Bar graphs display mean ± SEM. # denotes a significant interaction. * denotes a significant difference in phenotype. Circles denote individual animals. For t tests, * denotes a significant difference between means.
control (CD45.2+ C57BL/6) was chosen based on high IL-6 release ex vivo following leukocyte stimulation with LPS, whereas IL-6 levels were undetectable in IL-6−/− mice. IL-6−/− BM chimeras and IL-6−/− mice along with WT controls for each were exposed to 10 d of RSDS. IL-6−/− BM chimeras and IL-6−/− mice showed similar levels of resilience measured by the increased social interaction ratio (Fig. 4B), suggesting that leukocyte-derived IL-6 is critical in the development of social avoidance. Neither constitutive IL-6 deletion nor hematopoietic progenitor cell-specific IL-6 deletion affected anxiety-associated exploratory behaviors in the elevated plus maze (Table S2). To determine whether IL-6 from leukocytes impacted the behavioral response to a purely emotional stressor (with no physical component), we exposed IL-6−/− and WT BM chimeric mice to the witness model described previously (15). BM chimeras from high-IL-6 releasing controls displayed robust social avoidance behavior, whereas IL-6−/− BM chimeras demonstrated resilient behavior (Fig. 4C). Lastly, we injected neutralizing IL-6 monoclonal antibodies (mAbs), IgG isotype control antibody (IgG mAb), or saline systemically, 5 min before RSDS (Fig. S2E). IL-6 mAb is too large to enter the brain, but rather acts by sequestering and neutralizing IL-6 in the periphery (Fig. 4D). Consistent with the genetic deletion models, pharmacological blockade with IL-6 mAb, but not IgG mAb or saline alone, prevented the development of social avoidance (Fig. 4E). Treatment with IL-6 mAb did not reduce anxiety-associated exploratory behavior in the elevated plus maze compared with IgG mAb (Table S2).

Discussion

Here, we demonstrate the validity of a translational social stress animal model that recapitulates aspects of immune dysregulation observed in clinically depressed patients. Individual differences in the sensitivity of the peripheral immune system to social stress are preexisting and confer a greater risk of developing a stress-related disorder. Mice prone to developing a stress-susceptible phenotype had higher prestress levels of circulating leukocytes, largely due to monocyte populations, and these cells produced more IL-6 in response to acute stress and when stimulated ex vivo with LPS. Both in vivo and ex vivo IL-6 levels in response to stimulation were the strongest predictor of the behavioral response to a subsequent social stress. Although previous studies have identified elevated serum IL-6 and circulating leukocytes in stress disorders, this generally followed either depression diagnosis in humans (4, 25) or controlled stress exposure in rodents (26, 27). To our knowledge, this is the first study to show that the IL-6 response before social stress exposure can predict individual differences in vulnerability to a subsequent social stressor.

It is of particular interest that these individual differences in the sensitivity of the peripheral immune system occur within an inbred, genetically similar strain. As genetic differences are likely not driving these alterations, the possibility exists that they are due to epigenetic/environmental factors. Recent work has indicated that there is paternal transmission of stress sensitivity (28). Offspring of fathers that underwent RSDS display increased depression- and anxiety-related behavioral responses to stress (28). Additionally, stability of social hierarchy within the home cage has been demonstrated to induce anxiety-associated behaviors and alter monocyte trafficking to the brain (29). Differences in the social hierarchy within each cage may well affect susceptibility versus resilience to social stress. It is also important to note that our findings dissociate sickness behavior or general malaise from depression-like behavior. Despite large increases

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**Fig. 3.** BM hematopoietic progenitor cell transplant from a susceptible donor induces depression-like behavior in naive host animals. (A) A greater percentage of cells came from the donor versus host in BM chimeras across cell types (χ²(14, n = 58) = 24.11, P < 0.05, two tailed). The phenotype of the donor did not alter the distribution of leukocytes in either condition (viable cells, χ²(2, n = 58) = 0.5, P > 0.05, two tailed; T cells, χ²(2, n = 58) = 1.08, P > 0.05, two tailed; B cells, χ²(2, n = 29) = 0.04, P > 0.05, two tailed; monocytes, χ²(2, n = 58) = 1.28, P > 0.05, two tailed). (B) In a subset of animals tested, naïve host animals that received BM transplants from susceptible donors (n = 14) had higher levels of circulating leukocytes 28 d after transplant and before stress than host mice that received BM grafts from control donors (n = 15) (25, P < 0.05, two tailed). (C) Donor phenotype did not alter the percentage of microglia from the host or donor measured in a subset of the animals (χ²(1, n = 15) = 0.0, P > 0.05, two tailed). More than 98% of the microglia were of host origin. (D) Stress-susceptible BM chimeras that underwent a subthreshold defeat (n = 7) displayed social avoidance behavior, as indicated by a significant interaction and post hoc analysis, compared with control (no defeat, n = 8; subthreshold defeat, n = 7) and stress-susceptible BM chimeras that did not undergo subthreshold defeat (n = 7), as indicated by a significant interaction (F1,25 = 6.94, P < 0.05). (E) Stress-susceptible BM chimeras that witnessed RSDS (subthreshold witness, n = 7) but not animals with transplants from the same donors that did not witness RSDS (n = 8) or control BM chimeras (no witness, n = 6; subthreshold witness, n = 8) displayed greater social avoidance behavior, as indicated by a significant interaction and post hoc analysis (F1,25 = 4.99, P < 0.05). Bar graphs display mean ± SEM. * denotes a significant main effect of phenotype. # denotes a significant interaction between donor phenotype and exposure to subthreshold defeat/witness. For t tests, * denotes a significant difference between means.
Peripheral IL-6 controls susceptibility versus resilience to social defeat.

(A) Percent of donor versus host leukocyte concentration in blood of BM chimeras ($\chi^2(14, n = 32) = 70.76, P < 0.001$, two-tailed). IL-6$^{-/-}$ BM chimeras had similar distribution of leukocytes in blood compared with WT BM chimeras (viable cells, $\chi^2(2, n = 32) = 0.63, P > 0.05$, two-tailed; T cells, $\chi^2(2, n = 32) = 1.02, P > 0.05$, two-tailed; B cells, $\chi^2(2, n = 32) = 1.47, P > 0.05$; monocytes $\chi^2(2, n = 32) = 1.18, P > 0.05$). (B) Constitutive IL-6$^{-/-}$ (IL-6$^{-/-}$, n = 14; WT, n = 15) or IL-6$^{-/-}$ BM chimeras (IL-6$^{-/-}$, n = 9; WT, n = 8) are resilient following 10 d of RSDS, as indicated by a significant main effect of IL-6 deletion ($F_{1,24} = 9.18, P < 0.01$). (C) IL-6$^{-/-}$ BM chimeras (n = 8) are more resilient to witness defeat ($t_{12} = 2.21, P < 0.05$, two-tailed) compared with control BM chimera mice (n = 6). (D) An i.p. injection of IL-6 (n = 5), but not IgG (n = 4) mAb, blocked an increase in circulating IL-6 within 20 min of the first defeat ($t_{12} = 4.85, P < 0.01$, two-tailed). (E) Daily systemic injection of IL-6 (n = 22) but not IgG (n = 23) mAb or saline (n = 28) blocked the development of social avoidance behavior ($F_{2,37} = 7.14, P < 0.01$). Bar graphs display mean ± SEM. * denotes a significant main effect of IL-6 deletion. For $t$ test, * denotes a significant difference between means.

IL-6 may reduce stress-related relapse events for MDD patients in remission. It will be interesting to test more direct anti-IL-6 therapeutics in these patient populations. Similar strategies to block TNF-α with infliximab have shown promise in MDD patients with heightened inflammatory load (45). Although the direct clinical application of our findings remains unknown, prophylactic therapeutic strategies to inhibit IL-6 may reduce stress-related relapse events for MDD patients in remission.

Materials and Methods

Animals. All subjects were male. CD45.1+CD45.2+ C57BL/6 mice were used for RSDS and witness studies. BM transplant hosts (CD45.1+CD45.2+ C57BL/6) were obtained at 3 wk. IL-6$^{-/-}$ mice (B6.129S2-Il6tm1kopf)) on a CD45.2+ C57BL/6 background were bred at the Icahn School of Medicine at Mount Sinai from stock obtained from Jackson Laboratory. CD-1 mice (Charles River Laboratories) used as aggressors were sexually experienced retired breeders at least 4 mo of age. Aggressors were singly housed at all times other than during the social defeats. All other animals were group housed before social defeat and single housed following social defeat.

RSDS/Threshold Defeat. RSDS and threshold defeat were performed as previously described, and animals were tested for social interaction (13, 19).

Witness Defeat. Witness defeat was performed as previously described (15). Subthreshold witness defeat used a 14-d incubation period.

IL-6 mAb. Mice were given daily i.p. injections of mouse anti–IL-6 mAb (R&D Systems, Clone M5P-20F3), rat IgG, isotype control (R&D Systems, Clone 434141), or saline vehicle. Antibodies were given at a dose of 4 μg per mouse per day in 0.2 mL of saline vehicle. Antibodies/saline were injected 5 min prior to RSDS. The last injection was 24 h before SI.

Human Participants (Cohort 1). Following informed consent, male and female healthy volunteers or clinical patients underwent a medical and psychiatric evaluation and were required to be medically healthy to participate in the study. Clinical patients were diagnosed with MDD based on the Structured Clinical Interview for Diagnostic and Statistical Manual of Mental Disorders,
Flow Cytometry and Cell Cycle Analysis. Flow cytometry studies were performed using a Fortessa and LSRII (Becton Dickinson) and subsequently analyzed using FlowJo software (Tree Star).

Statistical Analysis. Differences between two groups were compared using t tests. Comparisons of three or more groups were analyzed by univariate ANOVA when appropriate, with Newman–Keuls used for post hoc analysis. Comparisons of multiple factors or repeated measures were analyzed using univariate ANOVAs with Bonferroni post hoc test. Percentages of leucocyte population using流 were analyzed using χ² tests. All statistical analyses were performed using GraphPad Prism 5.0 software (Graphpad Software Inc.). Statistical significance was set at P < 0.05.

Additional detailed information about procedures used in these studies is described in SI Materials and Methods.

ACKNOWLEDGMENTS. The authors thank Cristina Costantino for her guidance in separating leukocytes and additionally they thank Eric J. Nestler and Ming-Hu Han for helpful discussions about this project. This research was supported by US National Institute of Mental Health Grants RO1 MH090264 (to S.J.R.) and RO1 MH104559 (to S.J.R. and M.M.), the Johnson and Johnson International Mental Health Research Organization Rising Star Award (to S.J.R.), Irina T. Hirsch/Monique Weill-Caulier Trust Research Award (to S.J.R.), a grant from Janssen Pharmaceuticals (to S.J.R.), the Brain and Behavior Research Foundation (G.E.H.), and US National Institutes of Health Grants T32 MH087004 (to M.P. and M.H.) and T32MH096678 (to M.L.P.), as well as a generous gift from Ms. Julia Jackson.