Correction

MEDICAL SCIENCES


The authors note that the following information should be added to the conflict of interest statement: “R.P.F. is a practicing attorney. He has never represented a claimant in a case alleging diagnosed CTE. R.P.F. is a director of the Brain Injury Research Institute (BIRI), which is a 501(c)(3) corporation. He has never billed nor been paid for any work he has done for BIRI.”

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In vivo characterization of chronic traumatic encephalopathy using [F-18]FDDNP PET brain imaging

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Chronic traumatic encephalopathy (CTE) is an acquired primary tauopathy with a variety of cognitive, behavioral, and motor symptoms linked to cumulative brain damage sustained from single, episodic, or repetitive traumatic brain injury (TBI). No definitive clinical diagnosis for this condition exists. In this work, we used [F-18]FDDNP PET to detect brain patterns of neuropathology distribution in retired professional American football players with suspected CTE (n = 14) and compared results with those of cognitively intact controls (n = 28) and patients with Alzheimer’s dementia (AD) (n = 24), a disease that has been cognitively associated with CTE. [F-18]FDDNP PET imaging results in the retired players suggested the presence of neuropathological patterns consistent with models of concussion wherein brainstem white matter tracts undergo early axonal damage and cumulative axonal injuries along subcortical, limbic, and cortical brain circuitries supporting mood, emotions, and behavior. This deposition pattern is distinctively different from the progressive pattern of neuropathology [paired helical filament (PHF)-tau and amyloid-β] in AD, which typically begins in the medial temporal lobe progressing along the cortical default mode network, with no or minimal involvement of subcortical structures. This particular [F-18]FDDNP PET imaging pattern in cases of suspected CTE also is primarily consistent with PHF-tau distribution observed at autopsy in subjects with a history of mild TBI and autopsy-confirmed diagnosis of CTE.

Significance

Mild traumatic brain injuries are frequent events in the general population and are associated with a severe neurodegenerative disease, chronic traumatic encephalopathy (CTE). This disease is characterized by abnormal accumulation of protein aggregates, primarily tau proteins, which accumulate in brain areas responsible for mood, fear, stress, and cognition. There is no definitive clinical diagnosis of CTE at the present time, and this new work shows how a tau-sensitive brain imaging agent, [F-18]FDDNP, may be able to detect the disease in living people with varying degrees of symptoms. Early detection would facilitate the most effective management strategies and provide a baseline to measure the effectiveness of treatments.

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stages I and II) to advanced (neuropathology stages III and IV) (7) (Tables S1 and S2). In addition, more than 80% of analyzed pathologically confirmed CTE cases also show transactive response (TAR) DNA-binding protein of ∼43 kDa (TDP-43) either as inclusions in sparse neurites in cortex, medial temporal lobe structures, and brainstem in CTE neuropathology stages I–III, as widespread neuronal and glial inclusions in severe CTE cases (neuropathology stage IV), or in CTE cases with motor neuron disease (7, 15) (Tables S1 and S2). CTE cases also can exhibit the presence of other fibrillar protein aggregates. McKee et al. (7) and Omulu et al. (8) reported that in autopsies determinations, less than half of all CTE cases and less than one third of “pure” CTE cases show amyloid-β (Aβ) deposits, predominantly as scattered cortical diffuse plaques in low density (Tables S1 and S2). Of note is that subjects with Aβ deposits were significantly older than those without. Moreover, their neuropathology was more severe than that in cases without Aβ deposits and was often combined with α-synuclein deposits (7). As an example, as reported by McKee et al. (7), of 30 CTE cases with at least some cortical Aβ deposits (of 68 confirmed CTE cases), 29 brains were from subjects who died in their seventh decade of life and one from a subject who died in his sixth decade.

Subsequent to our preliminary report (16), in this work we use [F-18]FDDNP, an imaging agent for fibrillar insoluble protein aggregates (16–20), and PET imaging with the aim of establishing (i) topographic brain localization of [F-18]FDDNP PET signals indicative of fibrillar neuroaggregates in retired professional American football players with suspected CTE (mTBI group) vs. controls (CTRL); (ii) determination of [F-18]FDDNP PET signal patterns in the mTBI group; (iii) presence of [F-18]FDDNP PET signal as a measure of neuropathology in the brain areas involved in mood disorders related neurocircuits; (iv) correlation of [F-18]FDDNP PET results with neuropathology distributions in confirmed CTE cases; (v) differential patterns of [F-18]FDDNP PET signals, and thus deposition of fibrillar neuroaggregates, in the mTBI group with respect to the AD group; and (vi) preliminary demonstration of differences in [F-18]FDDNP PET signal patterns in mTBI cases with different etiology, i.e., contact-sport–related mTBI in retired professional American football players vs. blast-induced mTBI in war veterans. We further intended to demonstrate that tau (vs. Aβ) specificity of high affinity PET molecular imaging probes may not be a necessary requirement when used in CTE subjects with primary proteinopathy in the form of PHF-tau (8): PET imaging probes potentially sensitive to TDP-43 aggregates and Aβ deposits, which are present in higher densities almost exclusively in older CTE cases with more advanced neuropathology (e.g., stage IV), could better define disease progression based on quantification of differences in regional loads of combined neuropathologies because additional neuropathologies appear in predictable topographical and temporal patterns.

Results

Brain [F-18]FDDNP PET Patterns in Suspected CTE (mTBI Group). In all types of neurodegenerative proteinopathies, including CTE, pathology-based diagnosis using protein specific immunohistochemistry (IHC) does not result only from the presence of protein pathology in the brain, but rather from detection of disease specific topographic patterns of pathology distribution combined with quantitation of regional and global pathology loads. Molecular imaging agents targeting such fibrillar protein pathologies must therefore accurately and reliably detect these disease-specific patterns of distribution and their changes both in the pattern complexity and in regional pathology loads that may evolve due to the progressive nature of these neurodegenerative diseases.

Demographic data for all mTBI group subjects are provided in Table 1. We identified four distinctive topographical patterns of brain [F-18]FDDNP PET signal in the mTBI group, which are presented in increasing complexity from T1 to T4 in Fig. 1 (Upper) in three views (transaxial, top row; sagittal, middle row; and coronal, bottom row). All four patterns distinguish the mTBI subjects from cognitively intact control subjects (CTRL group) (Fig. 1, Upper, and Table 2). The observed [F-18]FDDNP PET signal patterns are defined as follows:

i) Pattern T1 is predominantly subcortical in brainstem (midbrain) with localized involvement of the limbic medial temporal lobe structures (limited to amygdala).

ii) Pattern T2 shows [F-18]FDDNP PET signal in all subcortical areas analyzed in this study, in all limbic medial temporal lobe areas [amygdala and medial temporal lobe (MTL); hippocampus, entorhinal cortex, parahippocampal gyrus], and in parts of the frontal cortex including anterior cingulate gyrus (ACG).

iii) Pattern T3 shows further increases in signal intensity and pattern complexity: all affected areas in the T2 pattern plus

Table 1. Demographic information for the mTBI group

<table>
<thead>
<tr>
<th>Subject</th>
<th>Age (y)</th>
<th>Education (y)</th>
<th>Race</th>
<th>Active career (y)</th>
<th>Position</th>
<th>Retirement (y)</th>
<th>Diagnosis</th>
<th>MMSE (score)</th>
<th>HAM-D score</th>
<th>HAM-A score</th>
</tr>
</thead>
<tbody>
<tr>
<td>TBI01</td>
<td>59</td>
<td>18</td>
<td>AA</td>
<td>16</td>
<td>Linebacker</td>
<td>28</td>
<td>MCI-MD</td>
<td>25</td>
<td>11</td>
<td>13</td>
</tr>
<tr>
<td>TBI02</td>
<td>64</td>
<td>17</td>
<td>C</td>
<td>10</td>
<td>Quarterback</td>
<td>38</td>
<td>Normal</td>
<td>30</td>
<td>5</td>
<td>6</td>
</tr>
<tr>
<td>TBI03</td>
<td>73</td>
<td>18</td>
<td>C</td>
<td>16</td>
<td>Offensive guard</td>
<td>44</td>
<td>Dementia</td>
<td>17</td>
<td>8</td>
<td>5</td>
</tr>
<tr>
<td>TBI04</td>
<td>50</td>
<td>15</td>
<td>C</td>
<td>14</td>
<td>Defensive tackle</td>
<td>20</td>
<td>MCI-MD</td>
<td>28</td>
<td>17</td>
<td>12</td>
</tr>
<tr>
<td>TBI05</td>
<td>45</td>
<td>18</td>
<td>C</td>
<td>12</td>
<td>Center</td>
<td>16</td>
<td>MCI-A</td>
<td>29</td>
<td>6</td>
<td>0</td>
</tr>
<tr>
<td>TBI06</td>
<td>86</td>
<td>16</td>
<td>C</td>
<td>11</td>
<td>Running back</td>
<td>58</td>
<td>MCI-A</td>
<td>27</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>TBI07</td>
<td>62</td>
<td>16</td>
<td>C</td>
<td>17</td>
<td>Offensive guard</td>
<td>28</td>
<td>MCI-NA</td>
<td>27</td>
<td>16</td>
<td>12</td>
</tr>
<tr>
<td>TBI08</td>
<td>51</td>
<td>18</td>
<td>AA</td>
<td>14</td>
<td>Defensive lineman</td>
<td>22</td>
<td>MCI-MD</td>
<td>30</td>
<td>17</td>
<td>31</td>
</tr>
<tr>
<td>TBI09</td>
<td>59</td>
<td>15</td>
<td>AA</td>
<td>16</td>
<td>Running back</td>
<td>26</td>
<td>MCI-MD</td>
<td>27</td>
<td>14</td>
<td>12</td>
</tr>
<tr>
<td>TBI10</td>
<td>54</td>
<td>14</td>
<td>AA</td>
<td>12</td>
<td>Wide receiver</td>
<td>22</td>
<td>MCI-MD</td>
<td>25</td>
<td>15</td>
<td>16</td>
</tr>
<tr>
<td>TBI11</td>
<td>40</td>
<td>16</td>
<td>C</td>
<td>17</td>
<td>Center</td>
<td>7</td>
<td>MCI-A</td>
<td>27</td>
<td>17</td>
<td>21</td>
</tr>
<tr>
<td>TBI12</td>
<td>54</td>
<td>16</td>
<td>C</td>
<td>16</td>
<td>Offensive lineman</td>
<td>20</td>
<td>MCI-A</td>
<td>26</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>TBI13</td>
<td>62</td>
<td>16</td>
<td>AA</td>
<td>13</td>
<td>Offensive lineman</td>
<td>32</td>
<td>MCI-A</td>
<td>28</td>
<td>33</td>
<td>30</td>
</tr>
<tr>
<td>TBI14</td>
<td>54</td>
<td>16</td>
<td>C</td>
<td>9</td>
<td>Nose tackles</td>
<td>25</td>
<td>MCI-MD</td>
<td>26</td>
<td>21</td>
<td>22</td>
</tr>
</tbody>
</table>

A, amnestic; AA, African American; C, Caucasian; HAM-A, Hamilton Anxiety Score (mild to moderate symptoms: 18–24; moderate to severe: >24); HAM-D, Hamilton Depression Score (mild symptoms: 8–16; moderate to severe: >16); MCI, mild cognitive impairment; MD, multiple domains; MMSE, mini-mental state examination (dementia range <24); NA, nonamnestic.

Barrio et al.
additional cortical areas [posterior cingulate gyrus (PCG), lateral temporal lobe (LTL), and parietal lobe]; this pattern is not associated with severe ventricular enlargement and prominent cortical atrophy commonly observed in aged retired boxers with dementia pugilistica.

iv) Pattern T4 shows high [F-18]FDDNP PET signal throughout the cortical, subcortical, and limbic medial temporal lobe structures, as well as in the white matter areas; this pattern was associated with significant brain atrophy (MRI or CT); possible comorbidity of CTE with other neurodegenerative diseases may be suspected, e.g., AD or end stage CTE progressing to, and simulating, AD.

All mTBI subjects \((n = 14)\) were treated as a single group for further group comparison with the CTRL group \((n = 28)\). Analysis of [F-18]FDDNP PET signal patterns T1–T4 identified the dorsal midbrain and amygdala as two core regions with consistently increased [F-18]FDDNP PET signals, which separated the mTBI group from the CTRL group as shown in the 2D correlation graph in Fig. 1A (Lower). Statistical analysis (ANOVA with post hoc Tukey–Kramer test; Table 2) shows a high degree of statistical significance \((P < 0.0001)\) for group separation in both areas and for a positive correlation between these two areas within the mTBI group (Spearman rank correlation coefficient: \(r_S = 0.745, P < 0.01; \) Table S3). All subcortical and limbic medial temporal lobe areas have also met strict criterion for good separation of mTBI and CTRL groups (Tukey–Kramer test at \(P < 0.0001)\) (Table 2). Other subcortical areas also show positive Spearman rank correlations with both core regions (Fig. S1 and Table S3) reflective of similar trends of increasing regional [F-18]FDDNP DVR values from T1 to T4. However, some subcortical structures like thalamus and caudate/putamen in mTBI subjects show increased DVR values only in more severe cases.

In contrast to subcortical regions, cortical areas showed different trajectories of involvement in the [F-18]FDDNP PET signal patterns T1–T4 with a clear anterior–posterior gradient. This difference in cortical trajectories is reflected in separation of the mTBI and CTRL groups only in the ACG (Fig. 1B, Lower, and Fig. S1A) and frontal lobe (Figs. 1C, Lower, and 2A, z axis, and Table 2). Spearman rank correlations within the mTBI group were determined for combinations of all analyzed regions that have shown mTBI-CTRL group separations (and involvement in the [F-18]FDDNP PET signal patterns T1–T4) and correlations of the core regions (amygdala and dorsal midbrain) with a large number of subcortical regions [brainstem (pons), diencephalon (thalamus), and basal ganglia (striatum)]

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Fig. 1. (Upper) [F-18]FDDNP distribution volume ratios (DVR) parametric images showing patterns T1 to T4 of increased [F-18]FDDNP signal observed in the mTBI group compared with cognitive control subjects (Left). The T1 pattern shows involvement of two core areas which have consistently increased [F-18]FDDNP signal in all four patterns: amygdala (limbic) and dorsal midbrain (subcortical). Patterns T2 to T4 are marked by increase of [F-18]FDDNP signal in these two core regions and progressively larger number of subcortical, limbic, and cortical areas. Although more complex patterns (e.g., T4) overlap with AD in the cortex, midbrain and amygdala signals are elevated above the levels in AD (Table 2). An AD case is shown in the right column for comparison. (Lower) A is a 2D scatter plot showing [F-18]FDDNP DVR values in two core areas consistently involved in CTE (subcortical structures (dorsal midbrain) and limbic structures (amygdala)), clearly demonstrating separation of mTBI and CTRL groups. B and C demonstrate similar separation effect when dorsal midbrain is compared with cortical areas typically associated with CTE and its mood disorders, namely anterior cingulate gyrus (ACG) (B) and frontal lobe (C). mTBI subjects are represented by green circles, and CTRL subjects are represented by blue circles. See SI Materials and Methods for additional correlations of [F-18]FDDNP PET DVR values in dorsal midbrain and amygdala with several cortical and subcortical areas (Fig. S1).
and cortical regions (frontal, ACG, LTL, parietal) were all found to be significant ($P < 0.05$; Table S3). It is noteworthy that these structures—brainstem (midbrain, pons), limbic medial temporal lobe (amygdala), and frontal lobe (frontal cortex, ACG)—are interconnected by brain circuitry supporting normal mood and behavioral functioning (21), which is perturbed early in subjects with a history of multiple concussions (2, 22).

Results of ANOVA statistical tests for separation of DVR values in all three groups are given for all areas analyzed, and areas that meet the Tukey–Kramer test criterion of $P < 0.0001$ are identified for comparisons of the mTBI group with the CTRL and the AD groups separately (Table 2). Tables S4 (mTBI group), S5 (CTRL group), and S6 (AD group) show all regional DVR values for all subjects in all groups. Tables S7

### Table 2. [F18]FDDNP DVR group mean values for mTBI group (14 subjects), control group (28 subjects), and Alzheimer’s disease group (24 subjects)

<table>
<thead>
<tr>
<th></th>
<th>Limbic</th>
<th>Subcortical</th>
<th>Cortical</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Amygd</td>
<td>MTL</td>
<td>Midb-V</td>
</tr>
<tr>
<td>mTBI</td>
<td>1.397</td>
<td>1.183</td>
<td>1.330</td>
</tr>
<tr>
<td></td>
<td>(0.095)</td>
<td>(0.042)</td>
<td>(0.090)</td>
</tr>
<tr>
<td>CTRL</td>
<td>1.162</td>
<td>1.112</td>
<td>1.135</td>
</tr>
<tr>
<td></td>
<td>(0.036)</td>
<td>(0.023)</td>
<td>(0.057)</td>
</tr>
<tr>
<td>AD</td>
<td>1.242</td>
<td>1.189</td>
<td>1.137</td>
</tr>
<tr>
<td></td>
<td>(0.060)</td>
<td>(0.020)</td>
<td>(0.062)</td>
</tr>
</tbody>
</table>

|                | ANOVA  | F value | 59.82* | 73.26* | 29.26* | 26.56* | 24.36* | 15.01* | 13.97* | 21.82* | 45.83* | 13.72* | 72.00* | 44.93* | 82.04* | 5.43 |
|                | Student t test | mTBI-CTRL | * | * | * | * | * | * | * | * | **** | NS | N | S | N | S | NS |
|                |        | mTBI-AD | * | NS | * | * | * | * | * | * | NS | NS | NS | NS | NS | NS | NS |
|                |        | AD-CTRL | * | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS |
|                |        | Tukey–Kramer test | mTBI-CTRL | * | * | * | * | * | * | * | * | NS | NS | NS | NS | NS | NS | NS |
|                |        | mTBI-AD | * | NS | * | * | * | NS | NS | * | NS | NS | NS | NS | NS | NS | NS |
|                |        | AD-CTRL | * | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS | NS |

Mean group DVR values (SD values given in parentheses). ACG, anterior cingulate gyrus; Amygd, amygdala; F, frontal; Hypo-Th, hypothalamus; LTL, lateral temporal lobe; Midb-D, dorsal midbrain; Midb-V, ventral midbrain; MTL, medial temporal lobe; NS, not significant ($P \geq 0.0001$); Occ, occipital; P, parietal; PCG, posterior cingulate gyrus; Str, striatum (caudate nucleus and putamen); Th, thalamus (medial thalamus). *$P < 0.0001$; NS, not significant ($P \geq 0.0001$).

Results of ANOVA statistical tests for separation of DVR values in all three groups are given for all areas analyzed, and areas that meet the Tukey–Kramer test criterion of $P < 0.0001$ are identified for comparisons of the mTBI group with the CTRL and the AD groups separately (Table 2). Tables S4 (mTBI group), S5 (CTRL group), and S6 (AD group) show all regional DVR values for all subjects in all groups. Tables S7
(mTBI group), S8 (CTRL group), and S9 (AD group) show all regional Z-score values for all subjects in all groups.

**[F-18]FDDNP Results: Correlation with Autopsy of Confirmed CTE Cases.**

[F-18]FDDNP PET signal patterns T1–T4 are consistent with pathology reports showing involvement of subcortical structures (e.g., locus coeruleus, substantia nigra, dorsal raphe, thalamus, basal ganglia), limbic medial temporal lobe structures (amygdala and MTL), and frontal cortex (6–8, 10, 11, 23). The F-18 FDDNP PET signal in the subcortical structures is in accordance with the examples of tau IHC labeling of pons, midbrain, thalamus, and basal ganglia in CTE shown on large brain tissue samples in refs. 7, 10, 11, and 23 (Fig. S2, Left). Presence of tau IHC in subcortical regions is further supported by our evidence from confirmed CTE cases that were not scanned with [F-18]FDDNP PET (Fig. 3II and Fig. S2, Right). Midbrain and amygdala are unique brain regions with elevated [F-18]FDDNP signals found in all 14 mTBI subjects. DVR values shown in Table S4 suggest that the PET signal is not uniformly distributed throughout all midbrain structures. A dorsal-ventral gradient observed with PET imaging agrees well with tau IHC results (Fig. 3II, Left), which shows high IHC density in the periaqueductal gray in dorsal midbrain and lesser IHC levels in substantia nigra, red nucleus, and other nuclei situated in the ventral midbrain (Fig. S2, bottom example in the left panel) (11). Similarly, high [F-18]FDDNP signals in the amygdala and other limbic medial temporal lobe structures coincide with common CTE autopsy observations of high involvement of amygdala and MTL shown in deceased retired professional American football players (Fig. 3III) (7, 23).

**[F-18]FDDNP PET in mTBI vs. AD.** From a perspective of clinical diagnosis, CTE can be mistaken for AD, particularly at later stages (24). However, the pattern of subcortical and limbic medial temporal lobe (amygdala vs. MTL) [F-18]FDDNP signal distribution in AD is quite different from that of concussion-based mTBI (suspected CTE) (Figs. 1 and 2 and Fig. S1) (25). To

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**Fig. 3.** Involvement of amygdala and midbrain areas in concussion-based mTBI is supported by both mechanistic concept of injury (I) and by the results of neuropathological examinations in deceased retired American football players with premortem complaints of functional impairments (II and III). (I) Rotation of the brain in the sagittal plane during a concussion, associated with significant accelerations and deceleration, will have significant negative effect on the brain tissue in the midbrain and thalamus (green shaded area) and on the affected cortical areas (red area). Stretching, compression, and shearing of axons during such sudden brain movements is hypothesized to be the cause of axonal injury (reprinted from ref. 33; reproduced with permission from Massachusetts Medical Society.) Online version of ref. 33 also contains an animated version of this figure (www.nejm.org/doi/full/10.1056/NEJMcp064645). Similarly, rotation in the coronal plane has been shown to lead to consistent damage to midbrain region tracts (27). (II) A–D show results of tau immunohistochemistry and demonstrate that in the mTBI group areas of increased [F-18]FDDNP signal in amygdala and dorsal midbrain coincide with presence of dense tau deposits in periaqueductal gray (PAG) in dorsal midbrain (A and B) and in amygdala (C and D; reprinted from ref. 21; reproduced with permission from Wolters Kluwer Health). (III) Amygdala and MTL areas are affected in the brains of retired professional American football players who died due to suicide (Left; 45-y-old retired player; reprinted from ref. 21; reproduced with permission from Wolters Kluwer Health) or due to natural causes (Right; 80-y-old retired NFL player; © Oxford University Press [brain.oxfordjournals.org/content/136/1/43] (reprinted from ref. 11). Amygdala and MTL areas are the first areas with high density of tau deposits in the neocortex and remain one of the most affected cortical regions in the majority of retired professional American football player cases.
facilitate visual comparison of AD and mTBI groups, we converted all DVR values in these two groups to Z-scores defined as 
\[ Z = \frac{\text{DVR}_{\text{AD}} - \text{DVR}_{\text{CTRL}}}{\text{SD}_{\text{CTRL}}} \]

where SD is the standard deviation. In both groups, our evidence shows that we can completely separate both groups using a combination of both core regions (amygdala and dorsal midbrain; x axis and y axis in the 3D graph in Fig. 2A).

The following general observations can be made:

i) The AD group showed higher DVR values compared with the CTRL group in subcortical regions (Table 2 and Fig. S1 C–F). Both the CTRL and AD groups had increased variability in [F-18]FDDNP DVR values in subcortical regions compared with cortical and limbic areas (Table 2). In contrast to subcortical regions, the AD group showed significantly higher signals than did the CTRL in cortical areas, with the exception of occipital lobe and the ACG (Table 2 and Fig. 2B).

The results were consistent with our previous reports on [F-18]FDDNP PET imaging in AD (18).

ii) The mTBI group showed higher DVR values compared with the AD group in all subcortical areas (Table 2 and Fig. S1 C–F), which is most clearly observed in more complex [F-18]FDDNP PET signal patterns T2–T4. Similarly, amygdala (a limbic medial temporal lobe area) also demonstrated significantly higher involvement in the mTBI group than in the AD group, which is consistent with pathology observations in neuropathology stages III and IV (7). By contrast, other limbic medial temporal lobe structures (MTLs) showed DVR values in the mTBI group that were comparable to those in the AD group. Fig. 2A presents a 3D scatter plot of Z-scores with limbic medial temporal lobe structures (amygdala), subcortical (dorsal midbrain), and cortical structures (frontal) demonstrating a good separation of the mTBI group from the AD and CTRL groups. Fig. S1 illustrates similar correlative measures under the Tukey–Kramer test criterion of \( P < 0.0001 \) involving subcortical, limbic, and selected cortical structures, which provided distinctive separation of the mTBI group from AD and CTRL groups.

iii) As previously reported (18), the AD group showed significantly higher [F-18]FDDNP signals in all cortical areas, and significant overlap with the mTBI group was observed in all cortical regions including MTL (Table 2, Fig. 2, and Fig. S1A). As predicted from the clinical symptomatology (mood vs. cognitive), the mTBI group showed predominant involvement in the frontal and ACG ROIs compared with the AD group. Fig. 2B further demonstrates that cortical [F-18]FDDNP PET signals do not discriminate between the mTBI and AD groups, whereas subcortical or limbic signals do. This lack of cortical discrimination becomes more obvious in the more severely affected and older (7) suspected CTE subjects (pattern T4).

These [F-18]FDDNP PET observations are consistent with the hypothesis that in individuals with suspected CTE, initial selected vulnerability is present in those circuits involving subcortical and limbic structures, including limbic structures in the medial temporal lobe. Through the medial temporal lobe and nodes in the prefrontal lobe (23), these circuits are connected with cortical areas involved in cognition via the default mode network (DMN), which includes the medial temporal lobe structures, the medial prefrontal cortex, posterior cingulate gyrus, and precuneus and parietal cortex, which may provide a predictable pathway for tau progression in CTE through cortical areas. In AD, tau and Aβ deposition are observed relatively early in the medial temporal lobe structures and the cascade of cortical pathology deposition typically follows the DMN with relatively lesser involvement of mood circuits in subcortical and limbic areas (18, 25, 26), which provides the basis for differentiation of the mTBI and AD groups.

**Discussion**

**Biomechanical Models of Concussions, Subject Population, and Brain Pathology.** TBI has a profound medical and social impact, and it may occur under a variety of circumstances, from domestic accidents to war-related events. A concussive brain injury results from rapid rotational and translational accelerations and decelerations as well as impact decelerations, which exert significant forces on brain tissue and cause membrane and axonal injuries without overt gross bleeding or focal parenchymal damage (27). Following axonal and membrane cellular injury, isomorphic astroglisis and microglial activation occur, which further interferes physically and chemically with regeneration and repair of disconnected axons (28), and leads to PHF-tau and TDP-43 deposition by a yet unknown mechanism. Because inflammation and white matter degeneration can persist for years (29), the brain is increasingly vulnerable to re-injury via traumatic axonal injury (30). Focal types of brain injury such as contusions or lacerations that commonly occur in moderate and severe traumatic injuries in humans or primate models of traumatic brain injury (27, 31, 32) are only rarely reported in concussions (33).

As such, American football players present a relatively homogeneous group based on the type of concussions and subconcussions frequently present in this subject population (2). The initial, pioneering observation by Omalu et al. of extensive tauopathy deposition in the brains of American football players (8, 34), involving subcortical, limbic areas, and brain cortices, led to the unequivocal demonstration of CTE as a unique clinical entity (35). Most recent detailed studies by McKee et al. (7, 11, 21) provided confirmatory results of the unique pathological characteristics of CTE. In a study describing 85 brains including American football players, boxers, and veterans, among others, McKee et al. demonstrated the extensive tau aggregate distribution in subcortical and limbic structures, as well as cerebral cortices, in 68 brains with pathologically confirmed CTE in various stages of the disease (7).

In CTE cases resulting from playing American football, tau deposits consist of PHF-tau IHC positive neuronal neurofibrillary tangles (NFTs) and neuropil threads, as well as astrocytic tangles. Distribution of PHF-tau IHC positive structures follows a pattern that increases in complexity both with age and symptom severity (6, 7, 11, 21). In cases of CTE with advanced pathology (stages III and IV) (7), dense deposits of tau pathology were consistently found predominantly in subcortical structures and limbic areas of frontal and temporal lobes with extension to cortical areas. Aβ aggregates are not frequently a distinctive pathological feature of CTE, with less than 50% of the cases presenting predominantly scattered sparse deposition of cortical Aβ in diffuse form, most frequently in later stages of pure CTE (stage IV) (7, 11). Often other protein aggregates, such as TDP-43 in more than 80% of all pathologically confirmed CTE cases, and more rarely α-synuclein, can be present as secondary proteinopathies, also most prominently in later stages of the disease. TDP-43 is present in high density in midbrain of all CTE patients with motor neuron disease, but it is more variable in CTE patients stages I–III without motor neuron disease (7, 15, 21) (for topographic details of different pathologies in various disease stages see Fig. S2 and Tables S1 and S2). Thus, it cannot be discounted that some of the [F-18]FDDNP signal in the brainstem, thalamus, and basal ganglia of the mTBI group, particularly in disease stages with more advanced pathology and
increased deposit densities (stage IV) (7), originates from the presence of TDP-43. TDP-43 contains amino acid sequences that make it aggregation prone in vitro and also in vivo because a portion of neuronal and glial TDP-43 inclusion bodies in motor neuron disease and CTE appear as filamentous structures (36) but are generally not considered amyloid due to missing amyloid specific thioflavin-S fluorescent stain. Recent evidence shows that at least some of these inclusion bodies do show the typical thioflavin-S staining, which provides evidence for presence of amyloid fibrillar structures within them (36).

**Brain-Specific Neuropathology Accumulation Measured with [F-18]FDDNP PET.** This work offers compelling evidence of the ability of [F-18]FDDNP PET to detect neuropathology in the living brain of American football players in a manner consistent with the pattern of deposition found at autopsy (6, 7, 34). [F-18]FDDNP has a high affinity for tau fibrils ($K_\text{D} = 36.7 \pm 11.6$ nM vs. $K_\text{D} = 5.52 \pm 1.97$ nM for Aβ42 fibrils) (37), which provides a high binding potential (BP; defined as $R/K_\text{D}$; where $R$ = number of tau aggregate binding sites per volume of brain tissue) for tau imaging visualization in vivo (38). The very low regional density of other proteinaceous β-sheet-containing aggregates (e.g., TDP-43, diffuse Aβ) would result in a low [F-18]FDDNP binding potential for non-tau aggregates, below the limit of sensitivity of PET, in the early CTE stages (stages I and II) as previously shown with the low abundance of α-synuclein in a Lewy body dementia patient (17). However, the ability of [F-18]FDDNP PET to detect Aβ becomes important in helping to elucidate the burden of AD comorbidly when it is present, particularly in advanced neuropathology CTE stages (stage IV) when proteinaceous neuropathologies are widespread, and quantitation of total neuropathology load can reflect additional pathology burden due to comorbid conditions. This investigation further confirms the unique sensitivity of [F-18]FDDNP to tau aggregates in vivo, as demonstrated by earlier work in patients with a neurodegenerative tauopathy, progressive supranuclear palsy (PSP) (19). All these studies in humans are supported by the demonstration of the ability of the 6-dialkylamino naphthalenyl-2-cyanoacrylate scaffold (present in FDDNP) to bind tightly to tau aggregates as previously shown by X-ray analysis of crystals obtained by cocrystallization of DDNP with tau segments (39), in vitro binding affinities with tau fibrils, tau rodent models, and postmortem autopsy results and comparisons with premortem scans (16, 17).

These findings of brain tau accumulation and distribution in this subject population are also consistent with earlier observations that subcortical structures such as brainstem and thalamus are considered as the fulcrum or center point of force vectors that bear the maximal rotational forces in American football players receiving concussions (Fig. 3) (2, 33). Axonal degeneration due to thousands of cranial impacts resulting in concussive and subconcussive injuries leads to other neurodegenerative changes including PHF-tau deposition throughout the areas of selective vulnerability in heavily interconnected cortico-striato-pallido-thalamic loops that support emotions, mood, and behavior (for a review of this topic, see ref. 22). Initial widespread involvement of the limbic areas of the medial temporal lobe (followed by the frontal lobe) is also observed in the subjects studied in this investigation and defines core regions that are consistently affected as neuropathology distribution increases in complexity during CTE evolution (Fig. 1, Upper).

These [F-18]FDDNP PET imaging patterns are observed in all cases studied thus far, suggesting a “fingerprint” PET scan of neuropathology accumulation that is characteristic of CTE. This [F-18]FDDNP PET imaging topographic distribution, moreover, is quite different from that found in AD (Fig. 2) and is consistent with profiles of neuropathology distribution described by Omalu et al. (6) and McKee et al. (7) in autopsy specimens with confirmed CTE. Omalu et al. defined four distinct pathology phenotypes (6), of which phenotype III (brainstem predominant) closely resembles the T1 pattern, whereas phenotypes I and II fit the description of patterns T2 and T3 described in this work. Comparison with the pathology stages as defined by McKee et al. (7) clearly identifies that [F-18]FDDNP PET signal patterns T1–T4 may primarily result from more affected examples of neuropathology stage II and from stages III and IV (Table S2) (7, 11), as they all show prominent neuropathology in medial temporal lobe structures. [F-18]FDDNP PET signal pattern T4 parallels neuropathology of severe stage IV in widespread distribution of signal and in signs of severe atrophy of ventricles. Patterns T2 and T3 would fit into stage III and milder examples of stage IV (Table S2) (7).

**[F-18]FDDNP Signal Is Consistent with Mood and Cognitive Disorders in Suspected CTE.** Dysfunction within brain circuits, caused by axonal damage leading eventually to PHF-tau aggregate accumulation in CTE, is believed to be the trigger for the mood disorders observed in these subjects who have experienced multiple concussions and subconcussions. Presence of axonal injury in brainstem and cortical white matter has been demonstrated postmortem in teenagers who had a history of playing high school American football (11). Such neurodegenerative processes can produce typical manifestations, initially involving behavioral and mood symptoms and leading to subsequent impairments of cognition and, in some cases, motor impairments in later stages (22). The [F-18]FDDNP PET results in this work show direct early involvement of brain areas that participate in processing of emotions, mood, and behavior. We observed neuropathology deposition using [F-18]FDDNP PET even in the minimally affected subjects, in the amygdala, several areas of the frontal cortex including the ACG, medial thalamus, hypothalamus, and dorsal midbrain (which contains the periaqueductal gray; Fig. 3). This midbrain pathology distribution pattern has also been reported recently in a retired Australian rugby player with CTE (stage IV) (11). Our [F-18]FDDNP PET imaging results, together with neuropathology observations at autopsy, highlight the specific vulnerability of these midbrain structures, some of which are involved in maintaining consciousness, modulating pain, and controlling defensive behavior (27).

Resting state fMRI studies also have shown that connectivity in these circuits is altered in generalized anxiety disorder (40), panic disorder (41), and depression (42). Some aspects of these neuronal circuits have also been closely connected with other neurodegenerative tauopathies, e.g., progressive supranuclear palsy, as demonstrated by Gardner et al. (43) and further confirmed by direct measurement of PHF-tau accumulation with [F-18]FDDNP PET (19). It has been speculated that in this case, the connectivity of these circuits offers an opportunity for transmissibility of tau aggregation to different brain regions in a predictable manner (44). It is possible that similar relationships between neuropathology accumulation and the pattern of involved neuronal circuits also exists in CTE, and our current observations seem to confirm this possibility.

Regardless of the predictable mechanism of neuropathology transmissibility through the connectivity of these circuits, the present study supports a correlation between the mechanical effects of concussions and subconcussions on specific brain areas and accumulation of neuroaggregates with the subsequent neuronal dysfunction and their corresponding neuropsychiatric consequences. However, effective confirmation awaits a larger cohort of subjects, also including subjects suffering brain concussions that may be biomechanically different from those found in American football players (e.g., war veterans, boxers). Exposure to blasts from explosive devices has caused changes consistent with TBI in war veterans, with clinical signs and changes in their DTI MRI scans consistent with traumatic diffuse axonal injury. Our initial [F-18]FDDNP PET studies in war veterans and other severe TBIs ($n = 8$) seem to indicate that blast-induced
Conclusions present with a different neuropathology deposition profile than that found in blunt-force TBI or contact-sport–related concussions (Fig. S3).

Clinical diagnosis of CTE remains elusive, and alternative approaches using blood-based biomarkers of cellular degeneration (45) are being explored; however, such methods may provide information about the presence of neurodegenerative processes but not the brain regions affected and thus may be inferior to brain imaging biomarkers. DTI fMRI (46) can provide useful information about axonal injury in the white matter in CTE but is not specific to this condition as other neurodegenerative diseases (e.g., AD) can have similar patterns of white matter degeneration caused by different mechanisms. Resting state fMRI has also been used to assess changes in large-scale cognitive networks in concussed athletes and in subjects with a history of TBI (47).

The present work suggests that, based on the definition of disease-specific tau pathology deposition stages in suspected CTE, we can use neuropathology deposition as a brain tissue target for PET molecular imaging probes with in vivo sensitivity for these neuroaggregates. However, considering that tau deposits are not specific for CTE but are present in other tauopathies and also in AD, a simple positive or negative reading with PHF-tau PET imaging probes is in itself clinically irrelevant. Only when combined with the regional sensitivity of PET will these probes provide significant in vivo information about regional tau deposition throughout the entire brain, at the earliest possible stage when medical management is most likely to succeed with CTE. This potential can be realized if the molecular imaging probes have sufficient sensitivity for in vivo detection of relevant neuropathology aggregates and reliably reflects the presence of these aggregates in all brain regions affected in a concentration–dependent manner, in agreement with autopsy determinations and with the prevalent mood and cognitive symptoms observed in this patient population.

Conclusion

This work with [F-18]FDDNP PET offers a sensitive method to visualize and quantify the regional presence of neuroaggregates in the living brain of human subjects with mTBI and suspected CTE and generates useful information about the mechanisms underlying disease staging and the mood disorders observed in these patients. It also offers the potential for early diagnosis of CTE in living subjects, when the probability of successful experimental therapeutic interventions would be greatest. Based on neuropathology indicators (7) (Tables S1 and S2), these [F-18]FDDNP distribution patterns in suspected CTE are largely due to PHF-tau in milder CTE stages (stages I–III), with possible TDP-43 and Aβ contributions in advanced CTE stages (stage IV) and in comorbidities. These [F-18]FDDNP topographic distribution patterns are unique among neurodegenerative diseases and significantly different from those found in AD (Figs. 1 and 2), progressive supranuclear palsy (19), and frontotemporal lobar degeneration (20). Thus, [F-18]FDDNP PET offers critical help in their differential diagnosis within a context of clinical history, physical examination, neuropsychiatric evaluation, and conventional radiological scans. These promising results provide the basis for a larger trial to determine the scope of this imaging procedure for CTE and for the use of this PET imaging technique to monitor disease progression in follow-up studies.

Materials and Methods

Written, informed consent for this [F-18]FDDNP PET imaging study was obtained from subjects in accordance with procedures of the Office of the Human Research Protection Program (OHRRP) of the University of California, Los Angeles, under strict, standard ethics guidelines.

Patient Population and Neurobehavioral Symptoms. The mTBI group included subjects with an increased risk of developing CTE as a result of having received repetitive concussions and subconcussions and presenting persistent cognitive, behavioral, and psychiatric problems, as defined by McKee et al. (7). It consisted of 14 retired male professional athletes who played American football professionally with histories of mood and cognitive symptoms as described above and in Table 1. Subjects TBI01–TBI05 from a preliminary report (16) have been included in this work. All mTBI subjects were college–educated retired professional American football players (age range, 40–86 y; mean ± SD: 57.2 ± 11.6 y; mean years of education ± SD = 16.2 ± 1.4 y), who had played from 9 to 17 combined professional (high school, college) and professional years (mean ± SD = 13.4 ± 3.6). All players, with the exception of a 64-y-old former quarterback (TBI02; Table 1), showed evidence of cognitive–impairment. A standard neuropsychological test battery indicated that 12 subjects had a diagnosis of mild cognitive impairment (MCI), which is a risk state for dementia. A 73-y-old former offensive guard had a diagnosis of dementia (TBI03). Most subjects also showed symptoms of depression (mean ± SD: HAMD-D score ± SD = 12.8 ± 8.3) and anxiety (mean ± SD: HAMA score ± SD = 2.9 ± 8.7). The mTBI subjects also had more prominent cognitive and mood symptoms than motor symptoms. As part of the neurological examinations, motor and balance performance were measured on nine subjects using a modified Balance Error Scoring System (BESS) assessment (mean score ± SD = 18.9 ± 5.6) (48) and the Chronic Brain Injury Scale (mean score ± SD = 2.9 ± 1.1) (49).

Radiosynthesis. [F-18]FDDNP was prepared as previously described (50) following current US Pharmacopeia 823 requirements for chemistry, manufacture, and control of PET radiopharmaceuticals for human use (51).

PET Scanning. All PET scans of the mTBI group were performed on a Biograph PET/CT camera, except for one subject who was scanned using an ECAT HR+ PET camera (subject TBI01) (both Siemens/CTI), with subjects in the supine position and with the imaging plane parallel to the orbito-meatal line as described previously (18). A bolus of [F-18]FDDNP (320–550 MBq) was injected through an indwelling venous catheter as a bolus i.v. injection following a 10-min attenuation scan. The dynamic data acquisition was started at the time of injection, and the following time frames were collected: 6 × 30, 4 × 180, and 10 × 300 s (total duration: 65 min). All PET scans were decay corrected and reconstructed using filtered back-projection (FBP) (60×60×60 mm3 FOV; 5.5-mm FWHM with scatter correction and motion correction). The resulting images from the PET/CT camera contained 109 contiguous slices with a plane-to-plane separation of 2.0 mm. Performance of PET scanners (e.g., Biograph, ECAT HR or ECAT EXACT HR+ scanner; Siemens CTI) was tested using the Hoffman brain phantom, and no significant differences between images from the two scanners were observed. All subjects from the AD and CTRL groups were scanned as previously reported (18) and summarized in the SI Materials and Methods.
MRI Scanning. MRI scans were performed for the purpose of anatomical reference to aid the analysis of PET data as described previously (18). T1-weighted magnetization-prepared rapid acquisition gradient-echo (MPRAGE) MRI volumetric scans, using a 3-T Siemens Allegra MRI scanner, were obtained for nine mTBI subjects (sagittal plane; repetition time, 2,300 ms; echo time, 2.93 ms; 160 slices; slice thickness, 1.3 × 1.3 mm; field of view, 256 × 256; flip angle, 8°). Four of the other mTBI subjects received CT scans because they could not tolerate an MRI scan (e.g., because of anxiety, claustrophobia, or metal in the body). One subject (TB10) was unable to undergo either MRI or CT scans.

Statistical Analysis. Statistical analyses were performed with the use of SAS software (version 9.2). P < 0.0001 was considered statistically significant unless otherwise stated. One-way ANOVA with Tukey-Kramer post hoc multiple comparisons was used to test for statistically significant differences in regional [F-18] FDDNP binding (DVR values) among the three groups of subjects. Multivariate ANOVA (MANOVA) was used to determine if a selected pair of regional DVR variables showed normal controls and CTEs. Spearman rank correlations were computed between regional [F-18] FDDNP binding within the mTBI group to determine their associations, and scatter plots were generated to visualize how regional DVR variables related to one another (P < 0.05 was considered statistically significant for this test). Discriminant analysis (DA) (54) was used to evaluate the performance of classification of subjects with known group memberships (CTRL, mTBI, or AD) based on the selected combinations of various regional [F-18] FDDNP DVR values as predictor variables. Because of a large number of possible combinations of predictor variables for DA, a backward elimination analysis was performed to determine the set of predictor variables that optimally discriminating the three groups (54). All predictor variables were first included, and the least significant variable (F-test, P > 0.15) was removed in a stepwise manner. This variable elimination procedure was repeated until no variable could be removed. The resulting reduced DA model functions and the canonical variables were used to assess the degree of overlapping between groups by calculating the percent of correct group classification and the overall classification accuracy, both of which were cross-validated by a leave-one-out method (54).

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8. Johnson VE, et al. (2013) Visualizing pathology deposits in known group memberships (CTRL, mTBI, or AD) based on the selected combinations of various regional [F-18] FDDNP DVR values as predictor variables. The resulting reduced DA model functions and the canonical variables were used to assess the degree of overlapping between groups by calculating the percent of correct group classification and the overall classification accuracy, both of which were cross-validated by a leave-one-out method (54).

29. Johnson VE, et al. (2013) Visualizing pathology deposits in known group memberships (CTRL, mTBI, or AD) based on the selected combinations of various regional [F-18] FDDNP DVR values as predictor variables. The resulting reduced DA model functions and the canonical variables were used to assess the degree of overlapping between groups by calculating the percent of correct group classification and the overall classification accuracy, both of which were cross-validated by a leave-one-out method (54).

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