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APPLIED BIOLOGICAL SCIENCES, CHEMISTRY
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The authors note that, due to a printer’s error, the affiliation for Moungi G. Bawendi should instead appear as “Department of Chemistry, Massachusetts Institute of Technology.” The corrected author and affiliation lines appear below. The online version has been corrected.

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Quantum dot/antibody conjugates for in vivo cytometric imaging in mice

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Multimodal, phenotypic, intravital cytometric imaging requires novel fluorophore conjugates that have an appropriate size for long circulation and diffusion and show virtually no nonspecific binding to cells/serum while binding to cells of interest with high specificity. In addition, these conjugates must be stable and maintain a high quantum yield in the in vivo environments. Here, we show that this can be achieved using compact (~15 nm in hydrodynamic diameter) and biocompatible quantum dot (QD)-Ab conjugates. We developed these conjugates by coupling whole mAbs to QDs coated with norbornene-displaying polyimidazole ligands using tetrazine–norbornene cycloaddition. Our QD immunoconstructs were used for in vivo single-cell labeling in bone marrow. The intravital imaging studies using a chronically calvarial bone window showed that our QD-Ab conjugates diffuse into the entire bone marrow and efficiently label single cells belonging to rare populations of hematopoietic stem and progenitor cells (Sca1+c-Kit+ cells). This in vivo cytometric technique may be useful in a wide range of structural and functional imaging to study the interactions between cells and between a cell and its environment in intact and diseased tissues.

Significance

One of the key questions in biology is understanding how cells move, interact, and evolve in living organisms. Tremendous efforts have been made to answer these questions in vitro, which have yielded a molecular-level understanding of cellular events. However, an increasing number of studies indicate that cellular activities need to be understood in the context of their natural environments. Single-cell labeling methods in use currently involve immunohistochemistry, genetic manipulation, or irradiation of mice, none of which reflect the native microenvironments. Here, we report quantum dot immunoconstructs that can be used for intravital imaging of single cells in unmanipulated mice and multiplexed in vivo cytometric analysis of rare cell populations.


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incorporating imidazole and PEG) effectively passivate the QD surface to achieve a high QY in aqueous solution (80–90%). PIL-coated QDs show greatly enhanced stability in buffers and blood sera, because the multidentate imidazole motifs of PILs are chemically stable and exhibit a high affinity toward the Cd- and Zn-rich QD surfaces (17). Norbornene, an uncharged functional group, allows incorporation of a high number of derivable functional groups while maintaining an overall neutral charge.

In contrast, conventional QDs displaying amine or carboxyl groups suffer from nonspecific binding to cells and biomolecules because of their surface charge. Implementing the advantages described above, we synthesized QD conjugates [QD-norbornene-tetrazine-Ab conjugates (QD<sub>NB</sub>–TzAb)] by coupling NB-PIL-coated QDs with tetrazine-modified Abs. In detail, QD<sup>NB</sup>–TzAbs were synthesized by exchanging the native ligand of QDs with NB-PILs and coupling whole antibodies to QDs using tetrazine–norbornene cycloaddition as shown in Fig. 1A. Tetrazine-modified antibodies were prepared by reacting a tetrazine derivative with the lysine groups of antibodies followed by immediate conjugation to the QD<sup>NB</sup> by mixing the two components at room temperature. In this study, we synthesized QD<sup>NB</sup>–TzAbs with diverse combinations of QDs [CdSe/CdZnS (QD<sub>570</sub>), CdSe/CdS (QD<sub>612</sub>), and InAs/CdZnS (QD<sub>680</sub>)] and mAbs (anti-mouse CD31, CD45, c-Kit, and Sca-1 antibodies or isotype-matched IgG).

Purification of free antibodies from QD<sup>NB</sup>–TzAb was essential to achieve single-cell labeling in vivo. Without purification, single-cell labeling results were inconsistent, and the labeling signal intensities were extremely low. Inconsistencies and low signals were caused by free antibodies left in the conjugate solution that bind to targeting antigens on the cell surface and block the binding of QD probes. The presence of free antibodies can drastically interfere with the binding of QD probes in vivo due to the following reasons. First, systemically injected QDs are delivered to the target cells by extravasation from vessels and diffusion throughout the interstitial space, and therefore, the effective concentration of the probes around the target cells is low. Second, the hydrodynamic diameter (HD) of free antibodies (~12 nm) is slightly smaller than that of QD conjugates (15–17 nm), yielding faster extravasation and diffusion. A previous method to conjugate EGF, a 6,045-Da protein with an HD of 2.7 nm, to QDs (16) utilizes dialysis filters to purify unconjugated proteins. However, an Ab (a 150-kDa protein with an HD of 12 nm) has a similar size and charge as a QD, and therefore, it is difficult to separate the two using commonly used size- or charge-based purification methods. Instead, free antibodies were successfully purified using gradient centrifugation. Separation using this method is based on density, with denser QDs and QD-Ab conjugate moving faster than free antibodies (SI Appendix, Fig. S1).

The hydrodynamic size of nanomaterials influences their biological accessibility, local distribution, and clearance in a live animal, and it can affect the motion of the targeted species (12, 13, 18, 19). To achieve sufficient transvascular permeability and interstitial diffusion, QD<sup>NB</sup>–TzAb was made as compact as possible by optimizing the incubation duration of QD<sup>NB</sup> with 12Ab and quenching unreacted tetrazine on the antibodies using 5-norbornene-2,2-dimethanol to prevent additional aggregation. Polyimidazole ligands also contribute to the small HD of our QD conjugates, because they directly bind to the QD surface (coordinating ligands). In contrast, typical commercial water-soluble QDs are prepared by encapsulating hydrophobic QDs with amphiphilic polymers and therefore, display higher HD. Dynamic light scattering measurements show that both QD<sub>612</sub>–TzAb and QD<sub>612</sub>–TzAb were more compact than commercial Qdot 625 and Qdot 625-Ab (Life Technologies) (Fig. 1B and Table 1).

Charge has a major effect on the transport behavior of nanoparticles in vivo and in vitro (20). A high surface charge, either positive or negative, tends to trigger nonspecific uptake of the particles by macrophages and results in more accumulation of the particles in the liver than in the targeting region after systemic administration. ζ-Potential measurements indicate that the surfaces of our QD<sup>NB</sup> and QD<sup>NB</sup>–TzAb conjugates were moderately negative (Fig. 1C and SI Appendix, Table S2).

Minimizing nonspecific protein adsorption on nanoparticles is critical for single-cell targeting in vivo. Nonspecific protein adsorption on particles not only hinders the application of the particles to specific targeting but also, can trigger unwanted biological responses in vivo. The level of nonspecific interaction of our QD probes with serum proteins was evaluated by incubating QD<sup>NB</sup> emitting at 612 nm (QD<sub>612</sub>–NB) in FBS at 37 °C for 4 h and monitoring the size change. The sizes of the HDs before and after incubation were measured using fluorescence correlation spectroscopy (SI Appendix, Fig. S2 and Table S5) and dynamic light scattering. As shown in Table 1, the HD of QD<sub>612</sub>–NB did not change after incubation in serum, indicating that QD<sub>612</sub>–NB does not absorb serum proteins nonspecifically. In contrast, commercial Qdot 625 (Life Technologies) showed a significant size increase after serum incubation (Table 1).

**In Vitro Specificity Study**

The targeting specificity of QD<sup>NB</sup>–TzAb was first tested in vitro using flow cytometry (Fig. 2). First, nonspecific binding of the QD<sup>NB</sup> and the QD<sup>NB</sup>–TzAb on cells was tested by incubating QD<sub>612</sub>–TzAb, QD<sub>612</sub>–TzAb on peripheral blood mononuclear cells (PBMCs), which were obtained from mice after lysis of RBCs. Ab(IgG) stands for nonspecific control IgG, which should show no binding ability. We did not detect any nonspecific adsorption of the QDs and the QD<sup>NB</sup>–TzAb(IgG) on the PBMCs.

![Fig. 1. QD<sup>NB</sup>–TzAb nanoconjugate synthesis and characterization. (A) Conjugation of QDs to Abs. Ab (IgG<sub>Ab</sub>) is used to represent the Ab, and Lys residues of Ab(IgG<sub>Ab</sub>) are marked in red. This diagram is not to scale. (B) Dynamic light scattering data of the QDs and the QD<sup>NB</sup>–TzAb illustrate that QD<sup>NB</sup>–TzAb conjugation does not yield noticeable aggregates and that the size of QD<sup>NB</sup>–TzAb is compact. (C) ζ-Potential measurements of the QDs and the QD<sup>NB</sup>–TzAb show that the surface charge of QD<sup>NB</sup> and QD<sup>NB</sup>–TzAb is moderately negative. (D) Gel filtration chromatography (GFC) traces of QD<sub>612</sub>–NB before and after serum incubation show that there is virtually no nonspecific binding of QD<sup>NB</sup> to serum proteins.](image-url)
In Vivo Specificity Study and Optimization of the Ratio Between QDs and Abs

Before proceeding to single-cell labeling, we tested the in vivo specificity of our QD-Ab conjugates by labeling blood vessels, which are mesoscale objects. For the in vivo specificity test, QDs or QD-Ab conjugates were administered through systemic injection in the retro-orbital sinus to transgenic mice, which selectively express GFPs in endothelial cells, and the blood vessels were imaged through a mammary fat pad window using a multiphoton microscope. Nonspecific binding of QD-612 and QD-612,750 to cells and tissue was first tested by injecting QD-612,750 and QD-612,750(IgG) and imaging vessels over 24 h. Fig. 3 A and B and SI Appendix, Fig. S11 show that both of QD-612 and QD-612,750(IgG) were cleared from the bloodstream within 20 h with no evidence of nonspecific accumulation. In contrast, we observed nonspecific accumulation of commercial Qdot 625 and Qdot 625(IgG) in random spots after the QDs were cleared from systemic circulation (Fig. 3 C and D and SI Appendix, Fig. S12). In vivo specificity of QD-612,750 was then studied by injecting QD-612,750,Ab(CD31) systemically and monitoring the labeling signals of endothelial cells over time. Colocalization of the QD signal with the GFP signal confirmed the specificity of the QD-612,750,Ab(CD31) toward endothelial cells (Fig. 3 E and F).

The performance of QD-612,750 samples with different ratios between QDs and Abs was evaluated to achieve bright and stable labeling signal in the vessel-labeling study. QD-Ab conjugates containing a higher number of QDs are expected to yield a brighter signal; however, a high number of QDs per Ab may increase the chance of detachment of QD-612,750,Ab from the labeled objects by perturbing the binding kinetics (lower affinity) or triggering unexpected effects in vivo. To find the best ratio, we synthesized QD-612,750,Ab(CD31) with QD to Ab(CD31) ratios of 3:1, 1:1, and 1:3 and analyzed the intensity and pattern of the labeling signals. As shown in Fig. 3 E and F and SI Appendix, Fig. S13, QD-612,750,Ab(CD31) with all three ratios specifically labeled the GFP+ vascular endothelium from 3 to 4 h up to 24 h after systemic injection. After 24 h, the QD signal from the vessels was still visible, but the observed pattern became more punctate. The punctate staining pattern may be caused by either endocytosis of the QD-612,750,Ab(CD31) into endothelial cells or detachment of the probe because of low affinity. The analyses of the labeling intensities of vessels illustrate that the signal was greater for QD-612,750,Ab(CD31) having a higher ratio of QDs but that the punctate pattern was also more dominant (SI Appendix, Figs. S14 and S15). To achieve a balance between signal intensity and signal stability, we chose the QD-612,750,Ab with the QD:Ab ratio of 1:1 for in vivo cytometry experiments.

Multiplexed Single-Cell Imaging in Vivo

We used our newly developed QD-612,750,Ab to show the imaging of single cells from an endogenous rare population in unmanipulated mice. As an example of rare cells, Sca1+c-Kit+ cells in the intact calvarial bone were imaged through a newly established chronic calvarial window placed in C57BL6 mice. In mice, Sca1+c-Kit+ cells consist of multipotent progenitors (short- and long-term hematopoietic stem cells) (21), which generate all blood cells throughout the lifetime of the organism. Although the function of hematopoietic stem and progenitor cells has been well-characterized, locating the cell niche within the entire bone marrow and studying cellular dynamics in their native environment is still difficult. By multiplexing the detection of QD-612,750,Ab(CD31) with QD-570,Ab(c-Kit) and QD-612,750,Ab(CD45), we can label Sca1+c-Kit+ cells in the bone marrow in vivo (Fig. 2 A and B). As shown in Fig. 2 C, the Sca1+c-Kit+ cells were labeled with QD-612,750,Ab(CD31) (red), QD-570,Ab(c-Kit) (green), and QD-612,750,Ab(CD45) (blue). Close examination revealed that 75% of PBMNCs showed high intensity of the QD-612,750,Ab(CD45), confirming efficient and specific immunostaining. The performance of QD-612,750,Ab(CD45) was also tested specifically for single-cell imaging in vivo, was also tested similarly with viable BMCs that were extracted from tibia and femur bones of FVB mice. As expected, a good correlation between the signal from QD-612,750,Ab and commercial antibodies was observed (Fig. 2 B). Of note, the expression level of antigens varies significantly depending on both the type of antigen and the cell type. Therefore, in some cases, antigen-positive cells are clearly distinguishable from antigen-negative cells (Fig. 2 A), whereas in the other cases, the antigen expression level is rather continuous (Fig. 2 B).
environments have been challenging because of the lack of appropriate methodologies (22). Recently, a method combining laser-scanning cytometry on tissue sections and confocal/multiphoton microscopy on thick femoral bone marrow slices was reported to study distribution patterns and the oxygenation status of stem cell niches in the entire femoral bone marrow (23). However, even this method used bone sectioning followed by ex vivo imaging. Our single cell-labeling method enables single Sca1<sup>+</sup> c-Kit<sup>+</sup> cell imaging across the entire calvarial window in unmanipulated live animals.

Intactness of a newly developed calvarial window was tested by performing single-photon imaging of the bone marrow of Ac<sup>C</sup>GFP/C57BL/6 mice, which express GFP ubiquitously in all cells. As shown in SI Appendix, Fig. S18, the bone marrow can be imaged from day 0 to day 8 without any signs of inflammation or hemorrhage.

Unbiased statistical analyses from single-cell imaging requires fluorescent probes that diffuse into the entire area of interest and label all of the cells of interest. To verify whether QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab diffuses throughout the whole bone marrow, we injected QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab (CD45) into FVB mice through the tail vein, and we analyzed the percentage of CD45<sup>+</sup> BMCs that are labeled with our QD immunoconstructs. The diffusion of our newly developed QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab in vivo was then compared with that of commercially available antibodies. The diffusion experiment was carried out by (i) administering QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(CD45) or commercial Ab (CD45)-APC through the tail vein into mice, (ii) waiting 24 h to allow the Ab-based probes to clear from vessels and interstitial space, (iii) extracting BMCs from tibia and femur bones, (iv) staining BMCs with Ab(CD45)-PE/Cy7 ex vivo, and (v) analyzing the proportion of CD45<sup>+</sup> BMCs that are labeled with QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab (CD45) using flow cytometry (Fig. 4A). To target cells in vivo, the Ab probes used in step i need to extravasate from vessels and diffuse throughout the interstitial space. In contrast, Ab(CD45)-PE/Cy7 used in step iv is directly incubated with BMCs and therefore, labels all CD45<sup>+</sup> BMCs. Fig. 4B and C shows the flow cytometry results on entire BMCs that have gone through the procedures described above. The horizontal axis in Fig. 4B and C is the fluorescent intensity from QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(CD45), and the vertical axis in Fig. 4B and C is the fluorescent intensity from Ab(CD45)-PE/Cy7. CD45<sup>+</sup> BMCs are colored green [based on the Ab(CD45)-PE/Cy7 signal] and CD45<sup>−</sup> BMCs are colored red in Fig. 4B and C. As shown in Fig. 4B and C, there were almost no nonspecifically labeled cells, which would be Ab(CD45)-PE/Cy7–negative and QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(CD45)– or Ab(CD45)-allophycocyanin (APC)–positive. In Fig. 4B, Right and C, Right, both CD45<sup>+</sup> BMCs (green) and CD45<sup>−</sup> BMCs (red) are replotted against the signal of the probes injected in vivo to clearly compare the diffusion of QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(CD45) (Fig. 4B) and Ab(CD45)-APC (Fig. 4C). In Fig. 4B, most CD45<sup>+</sup> BMCs (green) displayed high fluorescent intensity in the QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(CD45) channel, which indicates that QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(CD45) successfully labeled most of CD45<sup>−</sup> BMCs. In comparison, a bimodal distribution (stained and nonstained) of the Ab(CD45)-APC signal intensities was observed for CD45<sup>−</sup> BMCs in Fig. 4C. This bimodal distribution indicates that Ab(CD45)-APC did not label >30% of CD45<sup>+</sup> BMCs (Fig. 4C). Considering that the same amount of Ab was injected in both experiments (100 μg), we can conclude that QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(CD45) is more efficient in labeling CD45<sup>+</sup> BMCs than Ab(CD45)-APC. The higher population of observed labeled cells using QD immunoconstructs than using commercial antibodies may be explained by the higher optical absorption coefficient of QDs. Because of their high absorption coefficient, we expect that fewer QD conjugates are required for visualizing single cells in vivo than commercial antibodies. Based on our diffusion study, we claim that it is easier to detect cells that are distant from vessels using our QD conjugates than commercial antibodies. Differences in the deep tissue-labeling performance between the QD conjugates and the dye-labeled antibodies are even more extreme for multiphoton microscopy, because the multiphoton action cross-section is at least two orders of magnitude lower for organic dyes than for QDs (24).

Labeling of single cells from rare populations of hematopoietic stem and progenitor cells was successfully achieved using our QD immunoconstructs and multiphoton microscopy. Multiphoton microscopy was used to increase the imaging depth and minimize cell damage and background signals. As imaging probes, QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(C-Kit) and QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(Sca-1) (population markers for hematopoietic stem and progenitor cells), QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(IgG) (nonspecific control), and Hoechst 33342 (a DNA-binding dye that labels most of the cells in the bone marrow) were injected retro-orbitally in mice. Contrast-enhanced angiography was used to detect the bone marrow vessels, and second harmonic generation microscopy was used to image the bone (3) (Fig. 5). Intravital multiphoton microscopy imaging was performed 24 h after the injection of the QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab when unbound QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab was completely cleared from the blood circulation and the bone marrow interstitial space (25). Twenty-four hours after the injection, we observed that >0.3% of cells within the bone marrow were labeled with QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(Sca-1) and QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(C-Kit), but none were labeled with QD<sub>NB</sub><sup>612</sup>,<sup>Tz</sup>Ab(IgG) (Fig. 5 and SI Appendix, Figs. S19–S21). To confirm that we labeled all Sca1<sup>−</sup>c-Kit<sup>−</sup> cells from BMCs, we extracted BMCs from tibia and femur bones, stained them with Ab(Sca-1)-PE/Cy7 and Ab(c-Kit)-APC/Cy7 ex vivo, and analyzed
the percentage of Sca1+ c-Kit+, and Sca1+c-Kit+ cells using flow cytometry. As shown in SI Appendix, Fig. S22, populations of c-Kit+, Sca1+, and Sca1+c-Kit+ cells were 2.779%, 2.132%, and 0.217%, respectively. The numbers closely match with the percentages that are labeled with QD612 Tz Ab(c-Kit), QD562 Tz Ab(Sca-1), and QD562 Tz Ab(Sca-1) QD612 Tz Ab(c-Kit) in vivo, which were ~3%, ~1.7%, and ~0.3%, respectively. The consistency between in vitro and in vivo studies shows that our QD-Ab conjugates specifically and accurately label cells in vivo. Note that these population percentages are calculated from entire BMCs and not from Lineage− BMCs. A detailed method for processing raw images and identifying Sca1+c-Kit+ cells is described in SI Appendix, Fig. 19. In addition, we were able to track single cells in calvarial bone marrow for an extended period (>3 h). Movie S1 shows a stably bound intravascular Sca1+c-Kit+ cell interacting with a rolling WBC.

Lastly, we proved that the color or the composition of the QDs does not affect the quality of the imaging by labeling Sca1+c-Kit+ cells using QDNb,Tz Ab with different combinations of QDs and antibodies [QD900 Nb,Tz Ab(c-Kit) and QD562 Nb,Tz Ab(Sca-1)]. In this experiment, bone marrow vessels were visualized using endogenous GFP expression in endothelial cells in transgenic Tie2:GFP/FVB mice. SI Appendix, Fig. S23 shows successful labeling of Tie2+ Sca1+c-Kit+ and Tie2 Sca1+c-Kit+ cells, which shows that diverse combinations of QDNb,Tz Abs that emit from the visible to the near-IR (NIR) can be used for multiplexed phenotypic imaging in vivo.

Conclusions

For single endogenous cell labeling for microscopy imaging in animals, fluorescent probes are administered systemically and delivered to the target site through extravasation and diffusion. Because the delivery determines the effective concentration of the probes available for cell targeting, fluorophores for single-cell tracking should meet highly strict criteria, such as compact size, neutral charge, high brightness, high purity, and extremely low

Fig. 5. In vivo imaging of single endogenous Sca-1+c-Kit+ cells in the intact bone marrow using QDNb,Tz Ab. (A, Upper) A representative multiphoton microscopy image of a bone marrow cavity shown after removing the signal from the QD/Ab channels outside of the cells, which were segmented from Hoechst 33342 signal. Yellow cells (arrows) represent single Sca-1+c-Kit+ cells. Red and green cells represent Sca-1+ and c-Kit+ cells, respectively. An area containing an Sca-1+c-Kit+ cell is magnified in Upper Right. Translucent red and gray areas represent vessels and the bone, respectively. Lower Left shows the unmodified maximum intensity projection (~10 μm in the z direction). Lower Right shows the maximum intensity projections of each channel individually. (B) Additional images of bone marrow cavities containing Sca-1+c-Kit+ cells. Signals outside of the Hoechst 33342+ cells were discarded in these images, e.g., in A, Upper. (Scale bars: 100 μm.)
non-specific interactions with biomolecules. These design rules are much more stringent than those for commonly used in vitro probes, where the probes are delivered to target cells by direct incubation. For in vitro labeling, the lack of non-specific binding to serum/cell and the high purity of the probes are less critical owing to intense signals from the high concentration of labeling probes. The size of the probes is also not as critical as it is for in vivo labeling, where diffusion in the dense in vivo environment must be efficient to reach target cells.

Because of the strict requirements for single cell-labeling probes in vivo, the development of such fluorophores requires innovation and rigorous characterization. Here, we present QD-Ab conjugates that (i) are easily and efficiently derivable with targeting molecules or secondary reporters, (ii) display high stability and maintain a high QY in complex in vivo environments, (iii) possess minimal non-specific cell/serum binding, (iv) exhibit long circulation half-life for enhanced delivery to target sites, (v) are purified thoroughly from unbound targeting molecules, because free-targeting molecules can block target sites and yield a significantly decreased QD signal, (vi) have a compact size for enhanced diffusion throughout dense in vivo environments, and (vii) have narrow emission for multiplexed imaging. QD\textsuperscript{11,12}Ab conjugates bind specifically to cellular antigens both in vitro and in vivo and perform better than the commercial antibodies that we tested to label BMCs after systemic injection. Our in vivo microscopy imaging results show the applicability of QD-Ab conjugates for multiplexed in vivo cytomtric imaging of endogenous rare cells in unmanipulated mice. Note that the QD probes have several advantages over fluorescently labeled commercial Abs. Commercial fluorescent Abs use organic dyes (e.g., Phycobilins and Cy7) as chromophores, and there are many disadvantages associated with organic dyes, such as small multiphoton action cross-section, low photostability under intense illumination, and asymmetric emission with red tails.

This technique offers new possibilities to study single-cell interactions and responses and their microenvironments in live animals. First, intense and stable fluorescent signal of QDs allows long-term tracking of single cells in intact or diseased tissues in vivo. Second, our technique can be applied to un-manipulated animals as opposed to genetically modified, immunodeficient, or preirradiated mice. This method may be useful for the functional analysis of single endogenous cells in their native environments, such as measuring the oxygen level, glucose concentration, or local mechanical stress near the cell of interest. Oxygen measurements in the bone marrow and the Ska-1\textsuperscript{c-Kit\textsuperscript{+}} cell niche will be presented elsewhere. Third, the high photostability of QDs allows for FRET studies in vivo for extended periods, which can be used to study the microenvironments of target cells and cellular interactions. High photostability is essential for FRET studies to maintain the QY of donors and acceptors constant. Fourth, the technique described here can be extended to single-cell imaging in dense tissues or highly scattering environments by the use of NIR/IR QD-Ab conjugates. QDs are especially useful for NIR/IR imaging, because NIR/IR dyes display limited QY and orders of magnitude lower photostability (26, 27). The challenges addressed and overcome in this paper are not limited to QDs. Our work provides insights for the rational design of any nanoconstructs for in vivo applications.

**Materials and Methods**

Detailed methods on the following subjects are available in SI Appendix, SI Materials and Methods: vendor information on the antibodies, mice models, synthesis of QDs, QD ligands, tetrazine-modified Abs and QD\textsuperscript{11,12}Ab conjugates, fluorescence correlation spectroscopy, purification of QD\textsuperscript{11,12}Ab using gradient centrifugation, bicinchoninic acid (BCA) assay for the measurement of the ratios of QD to Ab in the QD\textsuperscript{11,12}Ab conjugates, flow cytometry (including gating and compensation strategies), preparation of chronic bone marrow window, intravital multiphoton imaging, physiological monitoring of mice during intravital microscopy, and image analysis.

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