In vivo internal tumor illumination by telomerase-dependent adenoviral GFP for precise surgical navigation

Hiroyuki Kishimotoa,b,c, Ming Zhaoa, Katsuhiro Hayashib, Yasuo Uratad, Noriaki Tanakac, Toshiyoshi Fujiwarac,e, Sheldon Penmana,d, and Robert M. Hoffmana,b,1

1Department of Surgery, Okayama University Graduate School of Medicine, Dentistry and Pharmaceutical Sciences, Okayama 700-8558, Japan; 2Oncoly BioPharma, Inc., Tokyo 106-0032, Japan; 3Center for Gene and Cell Therapy, Okayama University Hospital, Okayama 700-8558, Japan; and 4Department of Biology, Massachusetts Institute of Technology, Cambridge, MA 02139-4307

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Cancer surgery requires the complete and precise identification of malignant tissue margins including the smallest disseminated lesions. Internal green fluorescent protein (GFP) fluorescence can intensely illuminate even single cells but requires GFP sequence transcription within the cell. Introducing and selectively activating the GFP gene in malignant tissue in vivo is made possible by the development of OBP-401, a telomerase-dependent, replication-competent adenovirus expressing GFP. This potentially powerful adjunct to surgical navigation was demonstrated in 2 nude mouse models that represent difficult surgical challenges—the resection of widely disseminated cancer. HCT-116, a model of intraperitoneal disseminated human colon cancer, was labeled by virus injection into the peritoneal cavity. A549, a model of pleural dissemination of human lung cancer, was labeled by virus administered into the pleural cavity. Only the malignant tissue fluoresced brightly in both models. In the intraperitoneal model of disseminated cancer, fluorescence-guided surgery enabled resection of all tumor nodules labeled with GFP by OBP-401. The data in this report suggest that adenoviral-GFP labeling tumors in patients can enable fluorescence-guided surgical navigation.

Adenovirus | green fluorescent protein | metastasis

The intent of cancer surgery is to remove malignant tissue together with margins of presumably normal tissue (1–3) to ensure complete removal of abnormal cells. Estimating margin width during surgery is critical and depends on the surgeon’s vision. There have been many developments intended to improve the delineation of tissue margins using morphologic and optical differences between normal and abnormal tissue. This report describes a major enhancement of cancer surgical navigation using the selective fluorescent labeling, in vivo, of malignant tissue. Bright GFP fluorescence clearly illuminates the tumor boundaries and facilitates detection of the smallest disseminated disease lesions.

Highly selective viral replication in malignant cells growing in normal tissue has recently become possible using novel adenoviruses, OBP-301 (4–6) and, more recently, OBP-401 (7, 8). This latter virus, which can enter most cells, contains the replication cassette with the human telomerase reverse transcriptase (hTERT) promoter driving the expression of the viral E1 genes, and the inserted GFP gene. Virus replication and, hence, GFP gene expression occur only in the presence of an active telomerase, i.e., in malignant tissue (7). The OBP-401 virus was first tested by injection directly into Ht-29 human colon tumors orthotopically implanted into the rectum in Balb/c nu/nu mice (7). Subsequent para-aortic lymph node metastasis was observed by laparotomy under fluorescence. The adaption of GFP fluorescence to in vivo labeling of tumor tissue should facilitate precision surgical navigation in live animals and, very possibly, in a clinical surgical setting.

Results

Fluorescence Labeling of Human Cancer Cells with OBP-401 in Vitro. A549 tumor cells, growing in tissue culture, were infected with OBP-401, and the development of GFP fluorescence followed. The fluorescence intensity gradually increased after infection as the virus, with its GFP gene, replicated (Fig. 1A).

The extent of infection was tested by infecting red fluorescent protein (RFP)-expressing cancer cells, growing in cell culture, with OBP-401. These included A549-RFP, PC-3-RFP, HCT-116-RFP, and HT-29-RFP cells. In most cells, the introduction of green fluorescence changes the cell color from red to yellow, showing that most were infected by OBP-401. Any remaining red fluorescence clearly identifies those few cells that remain uninfected by the adenovirus. The color changes increased gradually followed by cell death due to the cytopathic effect of replicating OBP-401 (Fig. 1B and C).

Fluorescence Labeling of Subcutaneous Tumors by Infection with OBP-401. Both nonfluorescent PC-3 and red fluorescent PC-3-RFP human prostate cancer cells were inoculated s.c. (Fig. 2A and B). The resulting s.c. tumors were injected with 1 × 106 plaque-forming units (PFU) of OBP-401 as shown in Fig. 2B. A color change from red to yellow in the s.c. PC-3-RFP tumor and the onset of GFP fluorescence in the nonfluorescent PC-3 tumor were observed by the third day after virus injection (Fig. 2C). An RFP filter selectively showed the tumors’ endogenous RFP fluorescence (Fig. 2D). Similarly, a GFP filter showed GFP fluorescence induced in the tumors by OBP-401 (Fig. 2E). Infecting tumor cells that are endogenously expressing RFP with the GFP-expressing adenoviral vector OBP-401 clearly shows the extent of GFP labeling of the tumor. Cells showing a yellow fluorescence are infected with OBP-401, while the remaining red fluorescent cells clearly indicate the small portion that might remain uninfected.

Labeling Peritoneal Carcinomatosis with OBP-401. Peritoneal carcinomatosis was induced in the abdominal cavity of nude mice by inoculating 3 × 106 red fluorescent HCT-116-RFP human colorectal cancer cells. Various sized peritoneal disseminated nodules developed within 12 days. These were clearly visible by fluorescence imaging using a long-pass filter and/or a specific RFP filter (Fig. 3A and B). Even very small disseminated nodules were illuminated by RFP fluorescence (Fig. 3B). Although there was some autofluorescence from adjacent organs visible, the tumor nodules were not visible through a GFP filter (Fig. 3A and B).

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1To whom correspondence may be addressed. E-mail: penman@mit.edu or all@anticancer.com.
Once the malignant nodules were established at 12 days after intraperitoneal (i.p.) implantation of HCT-116-RFP cells, 1 × 10⁸ PFU OBP-401 were injected into the mouse abdominal cavity. Selective color filters showed that the HCT-116-RFP disseminated nodules expressed GFP fluorescence as well as RFP when examined 5 days later (Fig. 3C). RFP fluorescence...
was essentially coincident with that of GFP (Fig. 3C). These results indicate that i.p. injection of OBP-401 efficiently infected and labeled disseminated cancer.

**Labeling of Pleurally Disseminated Cancer with OBP-401.** These experiments assessed the effectiveness of OBP-401 labeling of pleural carcinomatosis in a mouse model of unlabeled A549 human lung cancer cells. The thoracic space of nude mice was inoculated with 2 × 10^6 cancer cells. Various sized disseminated plural nodules appeared within 10 days after implantation. At this time, 1 × 10^8 PFU of OBP-401 were injected into the thoracic cavity. Five days after injection of OBP-401, the cavity was examined using GFP fluorescence imaging. A representative mouse is shown in Fig. 4. Disseminated plural nodules were visualized by GFP expression (Fig. 4A and B). Even very small lesions, which are normally undetectable, were clearly illuminated by GFP fluorescence (Fig. 4C). Histological examination confirmed that these GFP-expressing tissues were adenocarcinomas. A representative histological section is shown in Fig. 4D. These results suggest that intrapleural injection of at least 1 × 10^8 PFU of OBP-401 can efficiently label disseminated pleural cancer. Lower doses of OBP-401 resulted in less efficient labeling.

**OBP-401 Fluorescence-Guided Resection of Disseminated Peritoneal Tumors.** In order to test the effectiveness of OBP-401-guided cytoreduction surgery, we used the peritoneal carcinomatosis model with nonfluorescent HCT-116 human colon cancer cells. Mice with peritoneal carcinomatosis were injected i.p. with OBP-401 at a dose of 1 × 10^8 PFU. Five days after viral administration, laparotomy was performed to remove intra-abdominal disease using fluorescence-guided navigation under anesthesia (Fig. 5A and B). A representative mouse after cytoreduction surgery with OBP-401-navigation is shown in Fig. 5C. Disseminated cancer nodules, which would otherwise be undetectable, were clearly visible by bright GFP fluorescence. The resected nodules were visualized as frozen sections under both fluorescence (Fig. 5D) and after hematoxylin and eosin (H&E) staining (Fig. 5E and F). These results suggest that OBP-401-labeling has significant potential for guiding cytoreduction surgery of disseminated cancer.
Discussion

The peritoneal surface is involved in more than 20% of patients with gastric, colon, and pancreatic cancers (1). Cytoreduction surgery requires resection of all visible tumors and stripping of all peritoneal surfaces that contain metastatic nodules (1, 2, 9). Therefore, visceral peritoneal involvement often requires concomitant resection of intra-abdominal organs such as the small intestine and colorectum.

The detection for fluorescence-guided surgery similar to that described for use in mice (11). In the present study, during surgery, even very small peritoneal lesions could be identified with GFP fluorescence (11).

Materials and Methods

Recombinant Adenovirus. OBP-401, containing the GFP gene under the control of the CMV promoter with the hTERT promoter driving the Kishimoto et al. PNAS Recombinant Adenovirus. Materials and Methods fluorescence (11). As described for use in mice (11). In the present study, during surgery, high sensitivity in a pleural carcinomatosis model (Fig. 4). OBP-401 GFP labeling could detect dissemination nodules with tumors with GFP. The change from red to yellow fluorescence highly selective induction of bright GFP fluorescence.

Implanting RFP-expressing cancer cell lines gave rise to fluorescent nodules whose color change clearly indicated the efficiency with which OBP-401 labeled disseminated peritoneal tumors with GFP. The change from red to yellow fluorescence indicated successful infection by OBP-401 (Fig. 3). Similarly, OBP-401 GFP labeling could detect dissemination nodules with high sensitivity in a pleural carcinomatosis model (Fig. 4).

Perhaps most importantly, we could remove disseminated disease in a peritoneal carcinomatosis model by using fluorescence-guided resection. These results suggest developing a dedicated excited-state fluorescence imaging system for applications to CT-guided surgery similar to that described for use in mice (11). In the present study, during surgery, even very small peritoneal lesions could be identified with GFP fluorescence (11).

Production of RFP Retroviral Vector. For RFP retrovirus production, the HindIII/Nott fragment from pDsRed2 (Clontech), containing the full-length RFP cDNA, was inserted into the HindIII/Nott site of pLNCX2 (Clontech) containing the neomycin-resistance gene. pTF7, a NIH 3T3-derived packaging cell line (Clontech), expressing the viral envelope, was cultured in DMEM supplemented with 10% FBS. For vector production, PT67 packaging cells, at 70% confluence, were incubated with a precipitated mixture of LipofectAMINE reagent (Life Technologies) and saturating amounts of pLNCX2-DsRed2 plasmid for 18 h. Fresh medium was replenished at this time. The cells were examined by fluorescence microscopy 48 h post-transduction. For selection of a clone producing high amounts of RFP retroviral vector (PT67-DsRed2), the cells were cultured in the presence of 200 to 1,000 μg/mL G418 (Life Technologies) for 7 d. The isolated packaging cell clone was termed PT67-DsRed2 (12–15).

RFP Gene Transduction of Cancer Cells. For RFP gene transduction, cancer cells were incubated with a 1:1 precipitated mixture of retroviral supernatants of PT67-DsRed2 cells and RPMI 1640 containing 10% FBS for 72 h. Fresh medium was replenished at this time. Tumor cells were harvested with trypsin/EDTA (12) post-transduction and subcultured at a ratio of 1:15 into selective medium, which contained 200 μg/mL G418. To select brightly fluorescent cells, the level of G418 was increased up to 800 μg/mL in a stepwise manner. RFP-expressing cancer cells were isolated with cloning cylinders (Bel-Art Products) using trypsin/EDTA. Cells were amplified by conventional culture methods in the absence of selective agent (12–15).

Animal Experiments. Athymic nude mice were kept in a barrier facility under HEPA filtration. Mice were fed with autoclaved laboratory rodent diet (Teklad LM-485, Western Research Products). All animal studies were conducted in accordance with the principles and procedures outlined in the National Institutes of Health Guide for the Care and Use of Laboratory Animals under assurance A3873–01.

Subcutaneous Tumor Model. Subcutaneous tumors were produced by injection of 3 × 106 noncolored PC-3 or PC-3-RFP human prostate cancer cells in 5-week-old nude mice. When tumors reached approximately 6 mm in diameter, the tumors were intratumorally injected with PBS for control or OBP-401 at a dose of 1 × 108 PFU in 100 μL PBS. Mice were examined for fluorescence expression with a long-pass filter (a filter for simultaneous observation of both GFP and RFP) or with specific filters for GFP or RFP.

Peritoneal Carcinomatosis Model of HCT-116 Human Colon Cancer Cells. Five-week-old nude mice were i.p. injected with noncolored HCT-116 or HCT-116-RFP human colon cancer cells (3 × 106 in 200 μL HBSS) using a 27-gauge needle. Twelve days after cancer cell inoculation, mice were injected i.p. with OBP-401 at a dose of 1 × 108 PFU in 200 μL PBS. Five days after virus injection, the abdominal cavity was directly examined by fluorescence imaging under anesthesia.

Pleural Carcinomatosis Model of A549 Human Lung Cancer Cells. Five-week-old nude mice were inoculated with noncolored A549 cells (2 × 106 in 200 μL HBSS) into the thoracic space using a 27-gauge needle. Ten days after cancer cell inoculation, OBP-401 at a dose of 1 × 108 PFU in 200 μL PBS was intrapleurally injected. Five days after virus injection, the pleural cavity was directly imaged for GFP expression. GFP-expressing tissues were removed and examined microscopically.

Fluorescence Optical Imaging and Processing. An Olympus OV100 Small Animal Imaging System containing an MT-20 light source was used. High-resolution images were captured directly on a PC (Fujitsu Siemens). Images were analyzed with the use of Cell® software (Olympus Biosystems) (16).

Histological Examination. For histological studies, GFP-expressing tissues were removed at the time of sacrifice and put into buffered formalin for 24 h at room temperature. All of the tissues were subsequently processed through alcohol dehydration and paraffinization. Tissues were embedded in paraffin and sectioned at 5 μm. All slides were stained by H&E, and examined microscopically.

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