

# Supporting Information

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## SI Text S1

**Model Description.** *SI Text S1* provides the mathematical description of the plant model (partially adapted from ref. 1). For parameterization of the model, see *SI Text S2* and Table S1; for the calculated variables by the model, see Table S2; for the measured functional trait values, see Table S3 (model version) and Table S4 (field estimates); for estimations of light and water niches of species, see *SI Text S3*.

The model plant is defined by a shoot system (Fig. S1). The crown has a cylindrical shape, with a given top height  $h_t$  (m), crown bottom height  $h_b$  (m), and crown radius  $r$  (m), and uniform distributions were assumed for the leaf area density  $\lambda_l$  (leaf area per crown volume,  $\text{m}^2\cdot\text{m}^{-3}$ ) and sapwood area density  $\lambda_s$ . The  $\lambda_s$  can be interpreted as the sapwood area connected to leaves (or in the petioles) per crown volume ( $\text{m}^2\cdot\text{m}^{-3}$ ). We defined only the sapwood area density and not heartwood area density, because heartwood has no maintenance costs and does not transport water, and mechanical constraints were not considered. The photosynthetic capacity of the leaves was characterized by an average nitrogen mass per leaf mass,  $N_{\text{mass}}$  ( $\text{kg}\cdot\text{kg}^{-1}$ ), and leaf mass per unit leaf area (LMA) ( $\text{kg}\cdot\text{m}^{-2}$ ).

**Biomass.** The vegetative plant mass  $M$  is the sum of structural leaf mass  $M_w$ , photosynthetic protein leaf mass  $M_n$ , and the sapwood wood mass  $M_s$  (all in kilograms),

$$M = M_w + M_n + M_s \quad [\text{S1}]$$

with

$$M_w = \rho_w A_l \quad [\text{S2}]$$

$$M_n = \rho_n N_{\text{mass}} \text{LMA} \cdot A_l \quad [\text{S3}]$$

$$M_s = \rho_s A_s L_s. \quad [\text{S4}]$$

Here  $\rho_w$  is the structural leaf mass per leaf area ( $\text{kg}\cdot\text{m}^{-2}$ ),  $\rho_n$  is the photosynthetic protein mass per nitrogen mass ( $\text{kg}\cdot\text{kg}^{-1}$ ),  $\rho_s$  is the sapwood wood mass per sapwood volume ( $\text{kg}\cdot\text{m}^{-3}$ ),  $A_l$  is the total leaf area ( $\text{m}^2$ ),  $A_s$  is the stem sapwood area below the crown (no tapering) ( $\text{m}^2$ ), and  $L_s$  is the average sapwood length between the stem base and leaves (m).  $A_l$  and  $A_s$  are calculated from

$$A_l = \lambda_l V_c \quad [\text{S5}]$$

$$A_s = \lambda_s V_c \quad [\text{S6}]$$

with crown volume  $V_c$  ( $\text{m}^3$ ) calculated as

$$V_c = \pi r^2 (h_t - h_b). \quad [\text{S7}]$$

Given the assumed uniform distribution of leaves and sapwood within the crown volume,  $L_s$  (m) is calculated as

$$L_s = 1/2(h_t + h_b) + 2/3r. \quad [\text{S8}]$$

We thus see that  $L_s$  is defined by a vertical length (left term) and a horizontal length (right term). Moreover,  $L_s$  defines a focal point in the crown (Fig. S1), for which the effective crown water potential  $\psi_1$  (MPa) and the effective internal crown  $\text{CO}_2$  pressure across all leaves  $c_i$  (Pa) are calculated.

**Carbon gain rate.** In this study, net photosynthesis or carbon gain rate of plants  $P_n$  ( $\text{kg C}\cdot\text{d}^{-1}$ ) is calculated as

$$P_n = P_g - R_m. \quad [\text{S9}]$$

The maintenance respiration rate  $R_m$  ( $\text{kg C}\cdot\text{d}^{-1}$ ) equals the sum of the respiration rate in the leaves and sapwood,

$$R_m = r_w M_w + r_n M_n + r_s M_s. \quad [\text{S10}]$$

Here  $r_w$  and  $r_s$  are the parameters for mass-based maintenance respiration rates of the structural leaf mass and the sapwood ( $\text{kg C}\cdot\text{kg}^{-1}\cdot\text{d}^{-1}$ ), and  $r_n$  is the respiration rate per photosynthetic leaf protein mass ( $\text{kg C}\cdot\text{kg}^{-1}(\text{proteins})\cdot\text{d}^{-1}$ ).

The gross photosynthesis of the plant  $P_g$  ( $\text{kg C}\cdot\text{d}^{-1}$ ) is calculated from

$$P_g = 43,200 \cdot 12 \cdot 10^{-9} \cdot P_j. \quad [\text{S11}]$$

Here  $P_g$  is the gross photosynthetic rate of the plant ( $\mu\text{mol C}\cdot\text{s}^{-1}$ ), and the number provides the conversion from ( $\mu\text{mol C}\cdot\text{s}^{-1}$ ) to ( $\text{kg C}\cdot\text{d}^{-1}$ ) when accounting for 12 sunlight hours per day.  $P_g$  is calculated from a biochemical photosynthesis model (2) and formulated as a big leaf model. The gross photosynthesis is described as the minimum of two dependent processes, i.e., carboxylation or rubisco limited photosynthesis rate and electron transport limited photosynthesis rate. Here, the carboxylation limited photosynthetic rate  $P_c$  and electron transport limited photosynthetic rate  $P_j$  ( $\mu\text{mol C}\cdot\text{s}^{-1}$ ) can be written as

$$P_c = V_{\text{cmax}} \left\{ \frac{c_i - \Gamma}{c_i + K_{\text{cmm}} \left( 1 + o/K_{\text{omm}} \right)} \right\} \quad [\text{S12}]$$

$$P_j = J_{\text{max}} \varphi_j(\xi) \left\{ \frac{c_i - \Gamma}{c_i + 2\Gamma} \right\}, \quad [\text{S13}]$$

where  $V_{\text{cmax}}$  ( $\mu\text{mol C}\cdot\text{s}^{-1}$ ) is the maximum rate of carboxylation of the plant,  $J_{\text{max}}$  ( $\mu\text{mol}^{\text{els}}\cdot\text{s}^{-1}$ ) is the maximum electron transport rate limited photosynthesis,  $c_i$  is the effective crown  $\text{CO}_2$  pressure across leaves at the focal point (Pa),  $\Gamma$  is a parameter for the  $\text{CO}_2$  compensation point (Pa),  $K_{\text{cmm}}$  (Pa) and  $K_{\text{omm}}$  (Pa) are the Michaelis–Menten constants for carboxylation and oxygenation, respectively,  $o$  is the crown oxygen pressure (Pa) set to the atmospheric  $\text{O}_2$  pressure, and  $\varphi(\xi)$  is a term for the light limitation effects.  $V_{\text{cmax}}$  ( $\mu\text{mol C}\cdot\text{s}^{-1}$ ) and  $J_{\text{max}}$  can be written as

$$V_{\text{cmax}} = k_c (1 - v_{\text{chl}}) (1 - v_j) N_{\text{mass}} \text{LMA} \cdot \text{LAI} \cdot A_c \quad [\text{S14}]$$

$$J_{\text{max}} = \frac{k_j}{4} \left( 1 - v_{\text{chl}} \right) v_j N_{\text{mass}} \text{LMA} \cdot \text{LAI} \cdot A_c. \quad [\text{S15}]$$

Here the parameter  $k_c$  is the carboxylation capacity per nitrogen mass ( $\mu\text{mol C}\cdot\text{kg}^{-1} \text{N}\cdot\text{s}^{-1}$ ),  $k_j$  is the electron transport capacity per nitrogen mass ( $\mu\text{mol C}\cdot\text{kg}^{-1} \text{N}\cdot\text{s}^{-1}$ ),  $v_{\text{chl}}$  is a fixed nitrogen fraction distributed to chloroplasts, and  $v_j$  is the nitrogen fraction partitioned to electron transport. It is assumed that  $v_j$  and the nitrogen fraction partitioned to carboxylation  $v_c$  sum to unity ( $v_j + v_c = 1$ ). We thus assumed that all nitrogen is deployed for photosynthesis.

The  $\varphi(\xi)$  term in Eq. S13 is a dimensionless function that specifies the light limitation effects on the electron transport limited photosynthesis. Such a function has been formulated for leaves, but under the assumption that  $\psi_1$  and  $c_i$  are defined at the focal point (Fig. S1), the function can be up-scaled to the crown. The function  $\varphi(\xi)$  varies between 0 and 1, increases linearly from 0 at low values of  $\xi$ , and approaches 1 asymptotically at high  $\xi$ -values,

$$\varphi(\xi) = \frac{(1 + \xi) - \sqrt{(1 + \xi)^2 - 4\theta_j\xi}}{2\theta_j}, \quad [\text{S16}]$$

with  $\theta_j$  the curvature factor for the nonrectangular hyperbola and  $\xi$  is the ratio of the light absorption rate over the capacity for the electron transport (dimensionless), defined as

$$\xi = \frac{qI_a}{k_j(1 - v_{\text{chl}})v_j \cdot N_{\text{mass}} \cdot \text{LMA} \cdot \text{LAI}}, \quad [\text{S17}]$$

where  $q$  is the quantum yield ( $\mu\text{mol electrons} \cdot \mu\text{mol photons}^{-1}$ ) and  $I_a$  is the absorbed light per unit ground area ( $\mu\text{mol photons} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ ) calculated as

$$I_a = I(1 - \text{Exp}[-K_1\text{LAI}]). \quad [\text{S18}]$$

Here  $I$  is the vertical light intensity on top of the crown ( $\mu\text{mol photons} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ ), and  $K_1$  is the light extinction coefficient (dimensionless) for leaves.

We can formulate  $c_i$  as a function of the effective crown water potential  $\psi_1$  in the focal point. We assumed steady state for the  $\text{CO}_2$  influx and the  $\text{CO}_2$  consumption by the crown,

$$G_s \frac{(c_a - c_i)}{p_a} = P_n. \quad [\text{S19}]$$

Here  $G_s$  is the stomatal conductance of the plant ( $\mu\text{mol C} \cdot \text{s}^{-1}$ ),  $P_n$  is the net crown photosynthesis rate of the plant ( $\mu\text{mol C} \cdot \text{s}^{-1}$ ),  $c_a$  is atmospheric  $\text{CO}_2$  pressure (Pa), and  $p_a$  is the atmospheric pressure set at  $1 \times 10^5$  Pa.  $G_s$  is modeled by scaling of a function for leaf stomatal conductance (3) to the whole crown,

$$G_s = G_{s0} + a \frac{P_n}{c_i - \Gamma} g_{\psi}. \quad [\text{S20}]$$

Here  $G_{s0}$  is the residual stomatal conductance of the plant ( $\mu\text{mol C} \cdot \text{s}^{-1}$ ) and the parameter  $a$  is a scaling parameter set to the value  $2p_a$ . A logistic equation describes a dimensionless stomatal sensitivity  $g_{\psi}$  to  $\psi_1$  (3),

$$g_{\psi}(\psi_1) = \frac{1 + e^{a_{\psi} \times \psi_{\text{ref}}}}{1 + e^{a_{\psi} \times (\psi_{\text{ref}} - \psi_1)}}, \quad [\text{S21}]$$

which for the given parameter values  $a_{\psi}$  and  $\psi_{\text{ref}}$  varies from 0 ( $\psi_1 \ll \psi_{\text{ref}}$ ) to 1 (at  $\psi_1 = 0$  MPa). Because we assumed that  $G_{s0} = 0$ ,  $P_n$  cancels from the  $\text{CO}_2$  balance equation (Eq. S20), and the calculation of  $c_i$  is simplified to an equation of  $\psi_1$  and a number of parameters,

$$c_i(\psi_1) = \frac{(a/p_a)g_{\psi}(\psi_1)c_a + \Gamma}{(a/p_a)g_{\psi}(\psi_1) + 1}. \quad [\text{S22}]$$

**Water flow.** Steady state is assumed for the plant transpiration  $E$  and the plant water transport through the sapwood  $F$ . Although not realistic at a timescale of minutes to hours (4, 5), such a steady state is a reasonable assumption for tree life and size patterns in the daily time-step model used here. Using Eq. S20 to define the conductance for water, we can write this steady-state

assumption for the transpiration and stem water flow, i.e.,  $E - F = 0$ ,

$$\gamma_{\text{wc}} \alpha \frac{P_n(\psi_1)}{c_i(\psi_1) - \Gamma} g_{\psi}(\psi_1) D - (\psi_b - \psi_g - \psi_1) \frac{K_s A_s}{L_s} = 0. \quad [\text{S23}]$$

For the left term,  $\gamma_{\text{wc}}$  is the ratio of water diffusivity over carbon dioxide diffusivity ( $\sim 1.6$ ) and  $D$  is the vapor pressure difference between leaf and air. The right term is the product of the pressure difference between stem base and focal point in the crown and the average sapwood conductance between the stem base and that focal point (Fig. S1). The pressure difference is based on the stem base pressure  $\psi_b$ , crown potential pressure  $\psi_1$ , and the pressure loss due to gravity  $\psi_g$ . Sapwood conductance is written as the product of the specific conductivity of sapwood  $K_s$ , the sapwood area  $A_s$  divided by the sapwood length  $L_s$  between stem base and focal point (Fig. S1).

Both steady-state assumptions [ $P_c = P_j$ , Eqs. S12 and S13 (see also Eqs. S14 and S15), and  $E = F$ , Eq. S23] will be parameterized for all traits, except for the crown water potential  $\psi_1$  and the optimal nitrogen partitioning, which is expressed by the nitrogen fraction allocated to electron transport  $v_j$ . The two steady-state equations are solved for  $\psi_1$  and  $v_j$ , and the solution for  $\psi_1$  also provides the crown  $\text{CO}_2$  pressure  $c_i$  (Eq. S22).

## SI Text S2

**Model Parameterization.** The sources and values used for the parameterization are provided in Table S1. Below, we shortly explain this parameterization.

**Environmental conditions.** We considered averaged diurnal environmental conditions, because this allows us to evaluate carbon gain rate patterns in a most simplified way. Atmospheric conditions were fixed; i.e., air temperature  $T = 25$  °C, air  $\text{CO}_2$  pressure  $c_a = 30$  Pa, atmospheric air pressure  $p_a = 100,000$  Pa, and air vapor pressure deficit  $D = 950$  Pa, which corresponds to a relative humidity of 70% at 25 °C. In the simulations, we evaluated the consequences of light availability by varying the external vertical light intensity  $I$  from 0 to  $1,500 \mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$  and of soil water availability by varying the soil water potential at the stem base  $\psi_b$  from  $-8$  to  $0$  Pa.

**Physiological parameters: photosynthesis.** For  $T = 25$  °C, most photosynthetic parameters are relatively stable across C3 plants (6). We estimated the carboxylation capacity per unit nitrogen  $k_c = 83,000 \mu\text{mol} \cdot (\text{kg N})^{-1} \cdot \text{s}^{-1}$  and the electron transport capacity per unit nitrogen  $k_j = 1,050,000 \mu\text{mol} \cdot (\text{kg N})^{-1} \cdot \text{s}^{-1}$  (7), the Michaelis–Menten constant for carboxylation  $K_{\text{cmm}} = 40.4$  Pa and for oxygenation  $K_{\text{omm}} = 24,800$  Pa, the oxygen concentration in the leaf  $o = 21,000$  Pa, the quantum yield  $q = 0.25$  ( $\mu\text{mol electrons} \cdot (\mu\text{mol photon})^{-1}$ ), the ratio for  $\text{H}_2\text{O}/\text{CO}_2$  diffusivity  $\gamma_{\text{wc}} = 1.6$ , and the curvature factor for the electron transport rate process  $\theta_j = 0.5$ . Other rather stable parameter values included the fraction of incident light absorbed by a leaf,  $K_1 = 0.86$ ; the protein mass per unit photosynthetic nitrogen mass  $\rho_n = 5.88$  was estimated.

**Physiological parameters: respiration.** Mass-based respiration rate parameters were estimated. The respiration rate of the plant is calculated from mass-based respiration rates for leaves and sapwood. The reported estimates for the sapwood mass-based respiration rate vary a lot in the literature (e.g., ref. 8 for saplings) and are prone to error (9). For comparative purposes, we gave the species the same sapwood mass-based respiration rate  $r_s = 0.4 \mu\text{mol C} \cdot \text{kg}^{-1} \cdot \text{sapwood} \cdot \text{s}^{-1}$ . For leaves, the structural leaf maintenance respiration rate  $r_w$  was set at  $4 \mu\text{mol C} \cdot \text{kg}^{-1} \cdot \text{leaf} \cdot \text{s}^{-1}$  for all species. For photosynthetic respiration rate, a maintenance respiration rate per unit protein  $r_n = 1,000 \mu\text{mol C} \cdot \text{kg}^{-1} \cdot (\text{protein}) \cdot \text{s}^{-1}$  was assumed.

**Functional trait variation across species.** We used species-specific measurements to estimate various functional trait values (Tables

S3 and S4). We set the sapling height  $h_t$ , crown bottom height  $h_b$ , crown radius  $r$ , leaf area index (LAI) (total leaf area per crown area), and stem sapwood area  $A_s$  for each species, on the basis of averaged values across 10 individuals per species. We also set LMA, nitrogen mass per leaf mass  $N_{\text{mass}}$ , and specific hydraulic conductivity  $K_s$  on the basis of measured values of the same individuals (10). For stomatal sensitivity, we used minimum leaf water potentials  $\psi_{\text{min}}$  as entrances for  $\psi_{\text{ref}}$ , i.e., the leaf water potential at which plants lost half of their stomatal conductance capacity. The sapwood mass density  $\rho_s$  was also measured for each species (as sapwood density) (10).

### SI Text S3

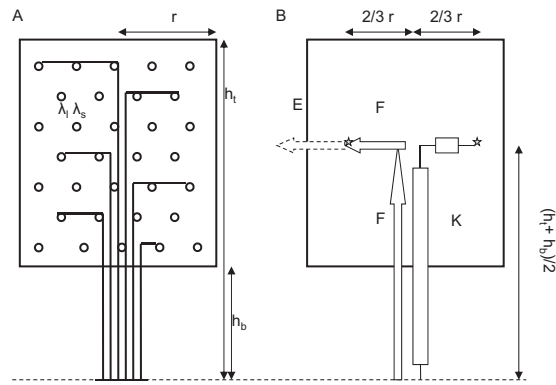
#### Species Estimates for Actual Water and Light Niches in the Forest.

Simulated carbon gain patterns along potential light and water gradients were compared with the average light and water conditions encountered by 2-m tall saplings of the studied species. Our light index quantifies the light conditions from an independent, objective, and continuous measure of crown exposure. This exposure measure was estimated for each species in relation to the height of  $\sim 1,253$  (range: 48–9,064) trees per studied species (11), on the basis of an inventory of these and other species in 80 ha of forest. This forest is the same area where we sampled our saplings for functional traits. Crown exposure was scored as 1 if the tree does not receive any direct light, 2 if it receives lateral light, 3 if it receives overhead light on 10–90% of the crown, 4 if it receives full overhead light on >90% of the crown, and 5 if it has an emergent crown (12). Crown exposure was measured repeatedly (mean difference between two independent observers is  $0.1 \pm 0.01$  SE), and the relation between the crown exposure and both canopy openness and incident ra-

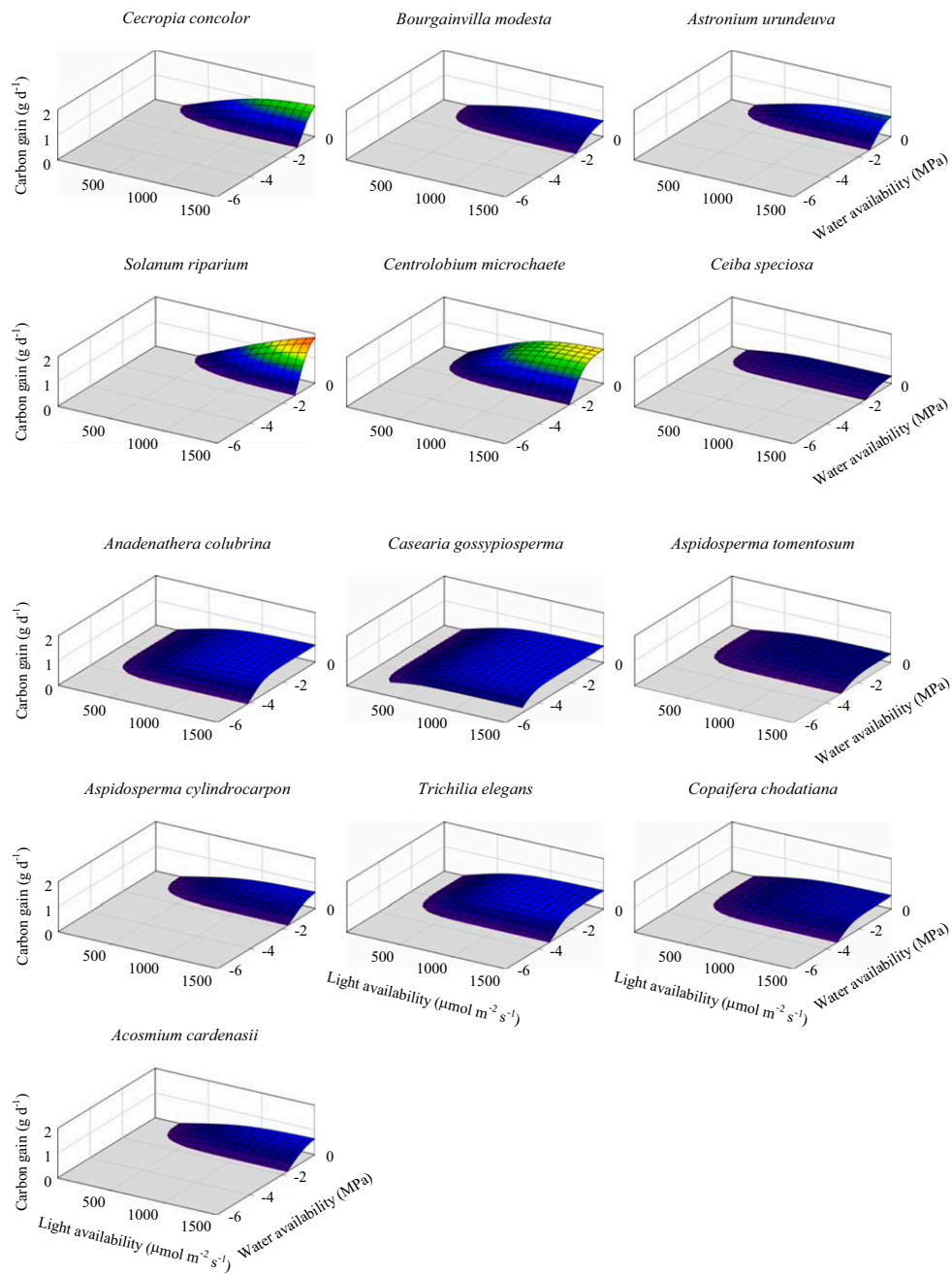
diation was good (13). Crown exposure was related to tree height using a multinomial regression analysis (14). The population average of crown exposure at a tree height of 2 m was used as the light index in our study.

Our water index quantified the water availability for each species, using an inventory of these species in the forest (12). In this inventory, a total of sixty  $10 \times 10$ -m plots were established along topographical gradients in 80 ha of forest area (the same forest area as used for the crown exposure measures, i.e., the light index). On the basis of the inventory, 20 plots were established in bottom, slope, and crest conditions, respectively. These topographic positions are good indicators of soil water availability in the studied forest, as well as in other tropical forests. Potential plot locations of each topographic location were randomly selected in the landscape with the help of elevation maps, and the suitability was verified in the field. Individual plots were located at least 50 m from each other. All trees and shrubs >30 cm tall were measured and identified to species. On average, a sample of 78 individuals (range 12–401) were scored per study species (10). We used the abundance (counts) of each of the 13 study species in the valley, slope, and crest plots to calculate the water index. We first calculated the relative frequencies (values between 0 and 1) per species for each topographic category, which sum to 1. We multiplied these relative frequencies by 1 for crest, 2 for slope, and 3 for valley to weight them for water availability and, in turn, took the means of these adjusted frequencies. We thus obtained a water index running from 0.33 to 1.0, which acts as a proxy of increasing water availability from crest, to slope, to valley. The validity of this index is supported because these topographic categories create a major source of variation in the soil water potential of the studied forest (15), as well as in other tropical forests (e.g., ref. 16).

1. Sterck F, Schieving F (2011) Modelling functional trait acclimation for trees of different height in a forest light gradient: Emergent patterns driven by carbon gain maximization. *Tree Physiol* 31:1024–1037.
2. Farquhar GD, von Caemmerer S, Berry JA (1980) A biochemical model of photosynthetic  $\text{CO}_2$  assimilation in leaves of  $\text{C}_3$  species. *Planta* 149(1):78–90.
3. Tuzet A, Perrier A, Leuning R (2003) A coupled model of stomatal conductance, photosynthesis and transpiration. *Plant Cell Environ* 26:1097–1116.
4. Zweifel R, Steppe K, Sterck FJ (2007) Stomatal regulation by microclimate and tree water relations: Interpreting ecophysiological field data with a hydraulic plant model. *J Exp Bot* 58:2113–2131.
5. Meinzer FC, Johnson DM, Lachenbruch B, McCulloh MA, Woodruff DR (2009) Xylem hydraulic safety margins in woody plants: Coordination of stomatal control of xylem tension with hydraulic capacitance. *Funct Ecol* 23:922–930.
6. Lambers H, Chapin FS, III, Pons TL (1998) *Plant Physiological Ecology* (Springer, New York).
7. Wullschlegel SD (1993) Biochemical limitations to carbon assimilation in  $\text{C}_3$  plants – a retrospective analysis of the  $A/C_i$  curves from 109 species. *J Exp Bot* 44:907–920.
8. Veneklaas EJ, Poorter L (1998) Growth and carbon partitioning of tropical tree seedlings growing in contrasting light environments. *Inherent Variation in Plant Growth. Physiological Mechanisms and Ecological Consequences*, eds Lambers H, Poorter L, van Vuuren M (Backhuys, Leiden, The Netherlands), pp 337–361.
9. Teskey RO, Saveyn A, Steppe K, McGuire MA (2008) Origin, fate and significance of  $\text{CO}_2$  in tree stems. *New Phytol* 177:17–32.
10. Markesteijn L, Poorter L, Paz H, Sack L, Bongers F (2011) Ecological differentiation in xylem cavitation resistance is associated with stem and leaf structural traits. *Plant Cell Environ* 34:137–148.
11. Poorter L, Kitajima K (2007) Carbohydrate storage and light requirements of tropical moist and dry forest tree species. *Ecology* 88:1000–1011.
12. Dawkins HC, Field DRB (1978) *A Long Term-Surveillance System for British Woodland Vegetation* (Department of Forestry, Oxford University, Oxford).
13. Clark DB, Clark DA, Rich PM (1993) Comparative analysis of microhabitat utilization by saplings of nine tree species in Neotropical rain forest. *Biotropica* 25:397–407.
14. Poorter L, Bongers F, Sterck FJ, Wöll H (2005) Beyond the regeneration phase: Differentiation of height-light trajectories among tropical tree species. *J Ecol* 93:256–267.
15. Markesteijn L, Iriapi J, Bongers F, Poorter L (2010) Seasonal variation in soil and plant water potentials in a Bolivian tropical moist and dry forest. *J Trop Ecol* 26:1–12.
16. Harms KE, Condit SP, Hubbel SP, Foster RB (2001) Habitat associations of trees and shrubs in a 50-ha Neotropical forest plot. *J Ecol* 89:947–959.



**Fig. S1.** Illustrations of the plant structure and water flow system. (A) The vegetative plant status elements visualized: plant height  $h_t$  (m), crown bottom height  $h_b$  (m), crown radius  $r$  (m), leaf area density or leaf area per crown volume  $\lambda_l$  ( $\text{m}^2 \cdot \text{m}^{-3}$ ), and sapwood area density or sapwood cross-section area (connected to leaves, i.e., in petioles) per crown volume  $\lambda_s$  ( $\text{m}^2 \cdot \text{m}^{-3}$ ). (B) The water fluxes in a plant, driven by a water potential gradient and resistances along the flow path.  $F$  stands for the water flux in the sapwood and  $E$  for the transpiration by the whole crown. The fluxes are calculated from the stem base to the focal point in the crown (star) (*SI Text S1*).  $K$  stands for the conductance of the sapwood between stem base and focal point (adapted from ref. 1).



**Fig. S2.** The predicted carbon gain  $P_n$  ( $\text{g}\cdot\text{d}^{-1}$ ) along gradients of light and water availability. Light availability is given by the vertical light intensity ( $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ), and water availability by the soil water potential (MPa). To make the zero carbon gain isoclines better visible, we do not show negative carbon gain where species are supposed to die due to carbon starvation (but those carbon gain rates are shown in Figs. 1A and 2A).

**Table S1. List of parameters (“constants”) in the model: symbols, units, explanation, input values, and literature sources**

Symbol	Units	Explanation	Input	Ref.
<b>Roman</b>				
$a$	Pa	Parameter for stomatal sensitivity function $g_{\psi}$	2	(1)
$a_{\psi}$	MPa <sup>-1</sup>	Slope parameter in stomatal sensitivity function $g_{\psi}$	3.2	(1)
$c_a$	Pa	Atmospheric CO <sub>2</sub> pressure	30	(2)
$D$	Pa	Vapor pressure difference between leaves and atmosphere	950	(2)
$G_{s0}$	$\mu\text{mol C}\cdot\text{s}^{-1}$	Residual stomatal conductance of the plant	0	
$I$	$\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$	Light intensity in horizontal plane above the plant or canopy	0–1,500	
$k_c$	$\mu\text{mol C}\cdot\text{kg N}^{-1}\cdot\text{s}^{-1}$	Carboxylation capacity per unit nitrogen	83,000	(3)
$K_{\text{cmm}}$	Pa	Michaelis–Menten constant for carboxylation	40.4	(4)
$k_j$	$\mu\text{mol C}\cdot\text{kg N}^{-1}\cdot\text{s}^{-1}$	Electron transport capacity per unit nitrogen	1,050,000	(3)
$K_{\text{omm}}$	Pa	Michaelis–Menten constant for oxygenation	24,800	(4)
$K_l$	—	Light extinction coefficient of crown	0.86	(4)
$o$	Pa	O <sub>2</sub> concentration in leaf, same as in atmosphere	21,000	(4)
$p_a$	Pa	Atmospheric pressure	100,000	(2)
$q$	$\mu\text{mol}\cdot\mu\text{mol}^{-1}$	Quantum yield ( $\mu\text{mol}$ electrons per photon)	0.25	(4)
$r_n$	$\mu\text{mol C}\cdot\text{kg protein}^{-1}\cdot\text{s}^{-1}$	Mass-based respiration rates for photosynthetic leaf mass	1,000	
$r_s$	$\mu\text{mol C}\cdot\text{kg}^{-1}\cdot\text{s}^{-1}$	Mass-based respiration rates for sapwood	0.4	
$r_w$	$\mu\text{mol C}\cdot\text{kg}^{-1}\cdot\text{s}^{-1}$	Mass-based respiration rates for structural leaf mass	4.0	
<b>Greek</b>				
$\gamma_{\text{wc}}$	—	Ratio H <sub>2</sub> O/CO <sub>2</sub> diffusivity	1.6	(4)
$\theta_j$	—	Curvature factor for the electron transport rate process	0.5	(4)
$\rho_w$	$\text{kg}\cdot\text{m}^{-2}$	Structural leaf mass per unit leaf area	0.05	
$\rho_n$	$\text{kg protein}\cdot\text{kg}^{-1}\text{ N}$	Protein mass per nitrogen mass	5.88	
$\Gamma$	Pa	CO <sub>2</sub> compensation point	3.7	(1)
$\psi_b$	MPa	Water potential at the stem basis	–8–0	

1. Tuzet A, Perrier A, Leuning R (2003) A coupled model of stomatal conductance, photosynthesis and transpiration. *Plant Cell Environ* 26:1097–1116.
2. Campbell GS, Norman JM (1998) *Introduction to Environmental Physics* (Springer, New York).
3. Wullschlegel SD (1993) Biochemical limitations to carbon assimilation in C<sub>3</sub> plants – a retrospective analysis of the A/C<sub>i</sub> curves from 109 species. *J Exp Bot* 44:907–920.
4. Lambers H, Chapin FS, Ill, Pons TL (1998) *Plant Physiological Ecology* (Springer, New York).

**Table S2. The variables that are calculated by the model and used for explanation of the model in SI Text S1**

Symbol	Units	Explanation
<b>Roman</b>		
$A_c$	m <sup>2</sup>	Crown area
$c_i$	Pa	Internal leaf CO <sub>2</sub> pressure
$E$	$\mu\text{mol}\cdot\text{s}^{-1}$	Transpiration rate of plant
$F$	$\mu\text{mol}\cdot\text{s}^{-1}$	Water flux through plant
$G_s$	$\mu\text{mol C}\cdot\text{s}^{-1}$	Stomatal conductance of the plant
$I_a$	$\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$	Photon absorption rate by plant per ground area
$L_s$	m	Average distance between stem base and crown leaves
$M_w$	kg	Structural leaf mass
$M_n$	kg	Photosynthetic mass in leaves (proteins)
$M_s$	kg	Living sapwood mass in plant
$M$	kg	Total vegetative plant mass
$P_c$	$\mu\text{mol C}\cdot\text{s}^{-1}$	Carboxylation limited photosynthesis rate of plant
$P_g$	$\text{kg C}\cdot\text{d}^{-1}$ (or $\mu\text{mol C}\cdot\text{s}^{-1}$ )	Gross photosynthesis rate of the plant per day
$P_j$	$\mu\text{mol C}\cdot\text{s}^{-1}$	Electron transport limited photosynthesis rate of the plant
$P_n$	$\text{kg C}\cdot\text{d}^{-1}$ (or $\mu\text{mol C}\cdot\text{s}^{-1}$ )	Net photosynthesis rate of the plant per day: $P_g - R_m$
$R_m$	$\text{kg C}\cdot\text{d}^{-1}$	Maintenance respiration rate
$V_c$	m <sup>3</sup>	Crown volume
<b>Greek</b>		
$v_j$	—	Fractions of nitrogen allocated to electron transport
$\psi_l$	MPa	Water potential at the focal point in crown
$\psi_g$	MPa	Water potential loss due to gravity in crown cylinder

**Table S3. List of functional traits**

Symbol	Units	Explanation
<b>Roman</b>		
$A_l$	m <sup>2</sup>	Total leaf area
$A_s$	cm <sup>2</sup>	Stem sapwood area
$h_t$	m	Height of the top of the plant
$h_b$	m	Height of the bottom of the crown
$K_s$	kg·m <sup>-1</sup> ·s <sup>-1</sup> ·MPa <sup>-1</sup>	Specific hydraulic conductivity of sapwood
LAI	m <sup>2</sup> ·m <sup>-2</sup>	Leaf area index or total leaf area per unit ground area: LAI = $A_l \cdot A_c^{-1}$
LMA	kg·m <sup>-2</sup>	Leaf mass per unit leaf area
$N_{mass}$	kg N·kg <sup>-1</sup> leaf	Average photosynthetic nitrogen mass per unit leaf mass
$r$	m	Radius of the crown cylinder
<b>Greek</b>		
$\lambda_l$	m <sup>2</sup> ·m <sup>-3</sup>	Leaf area density in the crown cylinder: $\lambda_l = A_l \cdot V_c^{-1}$
$\lambda_s$	m <sup>2</sup> ·m <sup>-3</sup>	Sapwood area density in the crown cylinder: $\lambda_s = A_s \cdot V_c^{-1}$
$\rho_s$	kg·m <sup>-3</sup>	Sapwood mass per unit volume
$\psi_{ref}$	Pa	Water potential for stomatal sensitivity function $g_{\psi}$ is about half the maximum; minimum leaf water potential $\psi_{min}$ is taken as proxy*

Model entries are based on data of saplings of 13 tree species of a Bolivian forest.

\*See Table S4.

**Table S4. Species-specific functional trait values for six pioneer tree species and seven shade-tolerant tree species of the Bolivian Inpa forest**

Trait units	LAI, m <sup>2</sup> ·m <sup>-2</sup>	$A_s$ , cm <sup>2</sup>	LMA, kg·m <sup>-2</sup>	$\psi_{min,r}$ , MPa	$\rho_s$ , kg·m <sup>-3</sup>	$K_s^*$	$N_{mass,r}$ , kg·kg <sup>-1</sup>
<b>Pioneers</b>							
<i>Cecropia concolor</i>	1.640 (0.52)	0.31 (0.054)	0.057 (0.003)	-0.17 (0.020)	211 (23)	5.0 (0.679)	0.022
<i>Bougainvillea modesta</i>	0.741 (0.031)	0.17 (0.023)	0.134 (0.006)	-0.96 (0.087)	419 (17)	0.7 (0.074)	0.019
<i>Astronium urundeuva</i>	0.988 (0.205)	0.09 (0.023)	0.092 (0.006)	-0.5 (0.044)	383 (19)	16.0 (1.340)	0.025
<i>Solanum riparium</i>	2.331 (0.155)	0.23 (0.029)	0.167 (0.012)	-0.39 (0.033)	250 (23)	11.3 (0.974)	0.018
<i>Centropogon microchaete</i>	2.390 (0.586)	0.25 (0.065)	0.060 (0.004)	-1.35 (0.378)	301 (27)	9.1 (0.604)	0.027
<i>Ceiba speciosa</i>	0.473 (0.046)	0.06 (0.006)	0.086 (0.003)	-0.84 (0.06)	241 (23)	7.3 (0.419)	0.020
<b>Shade tolerants</b>							
<i>Anadenanthera colubrina</i>	0.968 (0.057)	0.15 (0.008)	0.063 (0.002)	-3.42 (0.156)	499 (17)	2.6 (0.159)	0.027
<i>Casearia gossypiosperma</i>	1.062 (0.116)	0.15 (0.010)	0.049 (0.001)	-2.8 (0.178)	524 (14)	2.2 (0.222)	0.034
<i>Aspidosperma tomentosum</i>	0.459 (0.085)	0.10 (0.013)	0.090 (0.002)	-2.34 (0.12)	624 (21)	2.2 (0.081)	0.024
<i>Aspidosperma cylindrocarpon</i>	0.970 (0.082)	0.13 (0.012)	0.052 (0.010)	-1.75 (0.07)	492 (35)	2.5 (0.103)	0.026
<i>Trichilia elegans</i>	1.123 (0.274)	0.18 (0.016)	0.051 (0.003)	-2.59 (0.172)	571 (38)	1.9 (0.265)	0.034
<i>Copaifera chodatiana</i>	0.620 (0.227)	0.17 (0.063)	0.121 (0.003)	-2.6 (0.032)	629 (16)	2.8 (0.169)	0.019
<i>Acosmium cardenasii</i>	0.840 (0.097)	0.16 (0.013)	0.103 (0.003)	-3.3 (0.134)	514 (37)	1.2 (0.079)	0.025

We present mean values and SEs (in parentheses) for five saplings per species, except for  $N_{mass}$  (for which a pooled sample of five saplings was analyzed) (1). Functional plant trait abbreviations:  $A_s$ , the stem sapwood cross-section area;  $K_s$ , the specific hydraulic conductivity; LAI, leaf area index; LMA, the leaf mass per area;  $N_{mass,r}$ , the nitrogen mass per leaf mass;  $\psi_{min,r}$ , the minimum leaf water potential after 2 dry months; and  $\rho_s$ , the wood density.

\*Units for  $K_s$ : kg·m<sup>-1</sup>·s<sup>-1</sup>·MPa<sup>-1</sup>.

1. Markesteijn L, Poorter L, Bongers F (2007) Light-dependent leaf trait variation in 43 tropical dry forest tree species. *Am J Bot* 94:515–525.