

# Degrees and kinds of selection in spontaneous neoplastic transformation: An operational analysis

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**Spontaneous neoplastic transformation develops within days in the NIH 3T3 line of cells through differential inhibition of their proliferation under contact inhibition. A small fraction of the population continues to multiply after saturation density is reached and is selected to progressively increase saturation density in successive rounds of confluence. The degree of selection at confluence depends on the extent of proliferation of some cells in a heterogeneous population. The development of transformed foci is an extension of the same selective process that increases saturation density. The expression of the foci is enhanced with increases in the saturation density of the surrounding cells. Transformation is also induced by moderately reducing the concentration of calf serum in the medium during low-density passages, which allows selection of cells that require less growth factor. Further stepwise reductions in serum increase the degree of transformation. Contact inhibition and reduction in serum concentration select for the same phenotype of cell that increases saturation density and generates transformed foci. There is mounting evidence that selection is a major factor in the development of common epithelial tumors of humans, but it extends over decades rather than days, and the *in vivo* microenvironment selects from more stable populations of cells than those in culture. The many progressive levels of increased saturation density and transformed focus formation suggest that a very large number of genes participate in neoplastic development. The operational model of variation and selection presented here may aid in understanding chemical carcinogenesis and cancer recurrence after chemotherapy.**

carcinogenesis | epigenetics | microenvironment | progression | promotion

The continuous proliferation and repeated passage of rodent cells in monolayer culture eventually lead to their neoplastic transformation and capacity to produce tumors when injected into rodents of the same genetic background (1). This alteration was called spontaneous transformation, because it occurred in the absence of any known carcinogenic treatment and was assumed to reflect the spontaneous occurrence of mutations. The probability of appearance in mouse fibroblasts of early aspects of transformation, such as increased saturation density, rose with the concentration of cells used in their repeated passage (2). Passages of newly explanted mouse fibroblasts at densities that minimized cell–cell contact did not increase their saturation density over 200 generations in 11 months of subculturing, nor did they become tumorigenic (3). High-density passages with extensive contact among cells, however, increased their saturation density, and they became tumorigenic within 30 generations in 3 months of passage. The latent period of tumor appearance after injection of cells into mice decreased with increases in their saturation density, indicating a close relationship between the two properties. Inhibiting the proliferation of the BALB/c 3T3 line of mouse fibroblasts by suspension in agar resulted in their transformation on several occasions, whereas subculturing the cells at low population density in exponential proliferation while attached to a solid surface produced no transformation in the same period (4). The maintenance of a diploid line of rat hepatocytes under the selective condition of confluence was a more efficient inducer of tumorigenic capacity

than proliferation at lower densities, even when a strong mutagenic carcinogen was added to the latter (5). These findings indicated that selection plays a major role in the spontaneous neoplastic transformation of cells in culture.

The NIH 3T3 established line of mouse fibroblasts expressed very small lightly stained focal areas of overgrowth if maintained at confluence for  $\approx 10$  days in 10% calf serum (CS); much larger fully transformed foci appeared in a second round of confluence (6). Cells from such foci were highly tumorigenic in nude mice (7). It was possible to maintain uniformity in a confluent sheet of cells in the first and even in a second round of confluence when CS concentration was reduced to 2%. With very rare exceptions, the cells could be maintained without undergoing visible transformation, as measured by focus formation in 2% CS if they were kept in frequent low-density passages (LDP) in 10% CS. These methodological developments facilitated systematic and quantitative study of the progressive development of spontaneous transformation. The results with NIH 3T3 cells were scattered through the literature over a 10-year period and embedded in data on other related issues, which led to assorted conclusions about the nature of spontaneous transformation. The cardinal results for understanding spontaneous transformation are presented here and unified under an operational mechanism of progressive selection of cells with increasing capacity to proliferate under growth-regulatory conditions.

## Materials, Methods, and Results

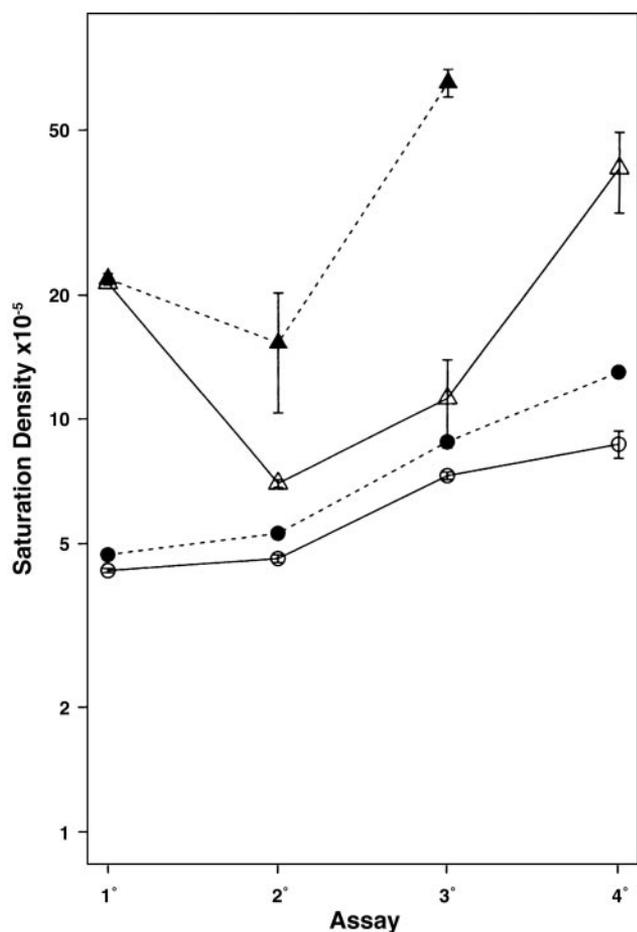
**Significance of Conditions for Assaying and Promoting Spontaneous Transformation.** The cells used in these experiments were from a frozen stock of the NIH 3T3 line provided in 1987 by S. A. Aaronson of the National Cancer Institute, Bethesda (8). The specific stock of cells used in the present experiment was described previously (9). The cells, which had been maintained in weekly LDP, were seeded in five cultures per experimental condition in primary ( $1^\circ$ ) assays for saturation density and transformed foci at  $10^5$  per 60-mm dish in 2% or 10% CS for 2 or 3 wk and were designated as follows: [2-2], 2% CS for 2 wk; [2-3], 2% CS for 3 wk; [10-2], 10% CS for 2 wk; and [10-3], 10% CS for 3 wk. For each group, one of the five cultures was fixed in Bouin's fluid and stained with Giemsa at the end of the  $1^\circ$  assay, and the other four were trypsinized for cell counting. Each counted culture was then reseeded in duplicate for secondary ( $2^\circ$ ) assays, all of which were carried out in 2% CS for 2 wk regardless of the conditions of the  $1^\circ$  assay. That resulted in four lineages per group. Half of the cultures were fixed and stained at 2 wk. The other four were used for tertiary ( $3^\circ$ ) and quaternary ( $4^\circ$ ) assays in succession, all under the same conditions as the  $2^\circ$  assay.

The aim of the experiment was to determine the extent to which the different conditions of the  $1^\circ$  assay influenced the saturation densities and transformed focus formation of the

Abbreviations: CS, calf serum; LDP, low-density passages;  $1^\circ$ , primary assay;  $2^\circ$ , secondary assay;  $3^\circ$ , tertiary assay;  $4^\circ$ , quaternary assay.

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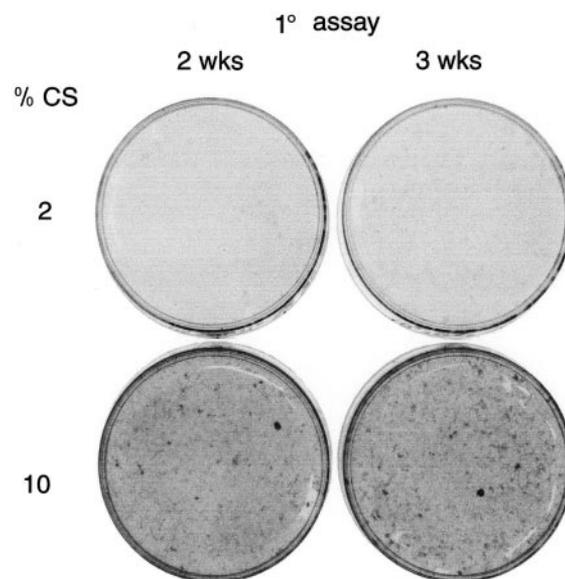


**Fig. 1.** Increases in saturation densities in serial 2°, 3°, and 4° assays at confluence in 2% CS for 2 wk resulting from variations in CS concentration and elapsed time in a preliminary 1° assay. Those 1° assay variations were: [2-2] 2% CS for 2 wk, O-O; [2-3] 2% CS for 3 wk, ●-●; [10-2] 10% CS for 2 wk, Δ-Δ; [10-3] 10% CS for 3 wk, ▲-▲. Where standard deviation bars are not seen, they are smaller than the symbols.

succeeding assays under a single common condition. Four independent lineages were maintained in each group to evaluate the variation within the group in the two properties. The experiment was originally intended to discriminate between epigenetic and genetic origins of changes in saturation density and in transformed focus formation. It was assumed that the uniformity of change in these properties among cultures of the same group would favor an epigenetic origin, whereas variation would favor a genetic origin.

The saturation densities are presented in Fig. 1, and the morphological appearance of the cultures can be seen in Figs. 2 and 3. The saturation densities of [2-3] cultures in the 1° assay were only slightly higher than those in [2-2], indicating very little net proliferation of the cells in the extra week of incubation in 2% CS. The saturation densities in [10-2] and [10-3] for the 1° assay were not significantly different from each other, indicating almost no net proliferation between 2 and 3 wk. The saturation densities of the 1° assays in 10% CS were 5-fold higher than those in 2% CS, signifying a direct relationship with the growth factors in serum.

The confluent cell sheets in both 1° assays in 2% CS were light and uniform in appearance, with no grossly visible sign of focus formation (Fig. 2). The cultures in 10% CS were much denser throughout and had many very small focal overgrowths, plus one or two slightly larger and still denser focal overgrowths sugges-

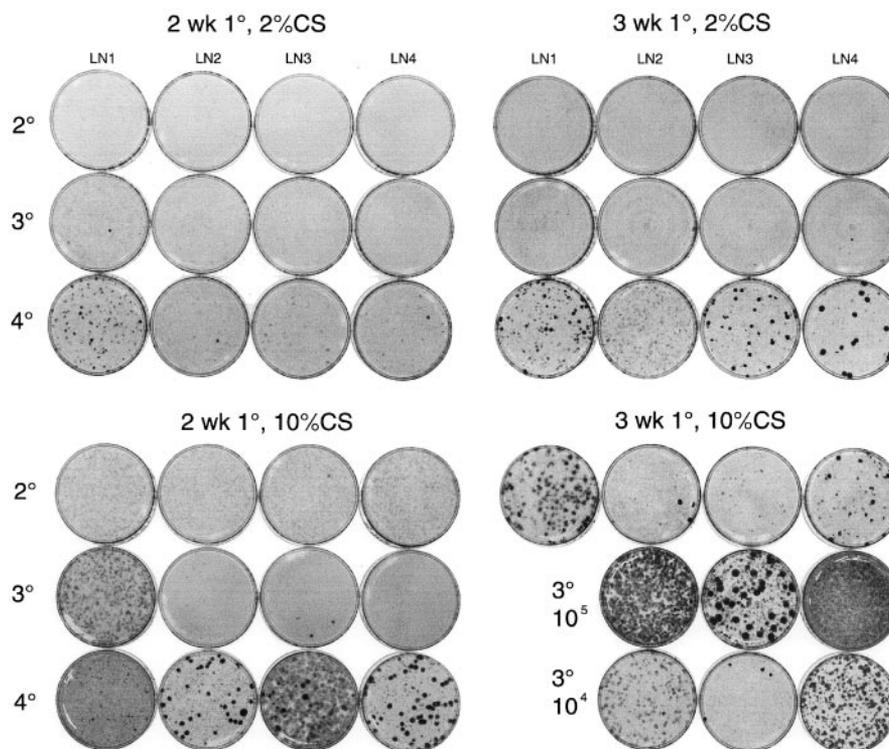


**Fig. 2.** Assays (1°) in 2% and 10% CS, fixed and stained at 2 and 3 wk. Note the light featureless monolayers in 2% CS at 2 and 3 wk in contrast to the darker cultures in 10% CS with small areas of spontaneous overgrowth, which increase in size and number between 2 and 3 wk, plus one or two larger denser focal areas that resemble early transformed foci.

tive of early-stage unequivocally transformed foci. The saturation densities of the serial 2°, 3°, and 4° assays that originated from the 1° in 10% CS were much higher than those that originated from 1° assays in 2% CS despite the presence of the same 2% CS for 2 wk in all of the later assays (Fig. 1). The saturation densities, in addition, were much higher in the [10-3] group than in the [10-2] group, and that difference increased sharply in successive assays. These differences indicated that considerable increase in capacity for growth at high density had occurred during the extra week in 10% CS despite the lack of significant increase in cell number in the 1° assay. The 2°, 3°, and 4° assays of the [2-3] group were modestly higher than those of the [2-2] group, but the difference between the two gradually increased with successive assays, indicating a moderate increase in capacity for growth at high density during the extra week of the 1° assay despite only a marginal increase in cell number.

The lack of variation in saturation densities within groups of most of the later assays derived from the 1° assays in 2% CS, and the 2° assays derived from the [10-2] 1° assay were originally suggestive of an epigenetic origin, particularly because an increase in saturation density in the absence of dense focus formation would necessarily be derived from a large fraction of the population. However, the presence of many small areas of overgrowth in the 1° assay of the [10-2] group and their increase in size and number in the [10-3] group (Fig. 2) introduce the likelihood of selection of cells with an advantage for overgrowth at high density in 10% CS. Although there were no grossly visible focal cell densities in 1° assays of the [2-2] and [2-3] groups, microscopic examination revealed such a trend. Therefore, opportunity for selective growth in 2% CS was present, but its extent was limited because of the low overall saturation density. Selective growth despite the meager net increases in cell number between 2 and 3 wk in both 2% and 10% CS can be explained by an offsetting detachment of cells with low capacity for survival in extended periods at high population density.

There was great variability in saturation density among the 2° assays of the [10-3] group, which could be credited to the appearance of many large, dense, transformed foci in lineage 1 and lesser but variable numbers in the other lineages within the



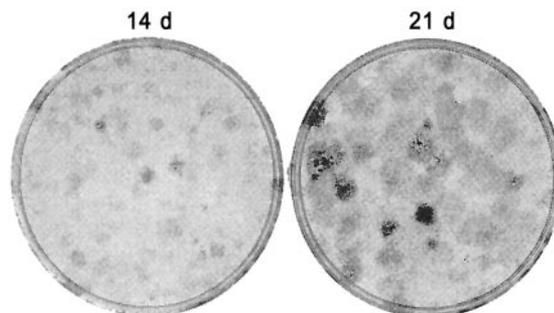
**Fig. 3.** Assays (2°, 3°, and 4°) for 2 wk in 2% CS of four lineages each for sets [2-2], [2-3], [10-2], and [10-3] of the 1° assay. Lineage 1 of set [10-3] was not carried beyond the 2° assay, because it already had so many foci in the 2° assay that it would certainly have produced confluent transformation in the 3° assay. Lineages 2, 3, and 4 of the same set were not carried beyond the 3° assay for the same reason, but they were also diluted 10-fold to  $10^4$  cells in the 3° assay and mixed with  $10^5$  cells from the standard LDP stock to form a confluent background for display of individual foci. All other assays were seeded with  $10^5$  cells. Note the reduction in size and density of the foci of lineages 2 and 3 when the cells were surrounded by LDP cells that had not previously been at confluence. LN, lineage.

group (Fig. 3). Similarly, large and variable numbers of foci appeared in the 3° and 4° assays of the [10-2] group, which accounted for the variability of saturation densities in those assays. The extra week of the [10-3] group in the 1° assay gave it a considerable head start over the [10-2] group in transformed focus formation as well as saturation density. Significant numbers of transformed foci did not appear until the 4° assay of the groups originating from 1° assays in 2% CS, and they were much smaller than those originating from 1° assays in 10% CS. The foci were larger and more common among the [2-3] than the [2-2] lineages, again indicating that the extra week in the 1° assay allowed significant progress in transformation despite only marginal net increase in cell number.

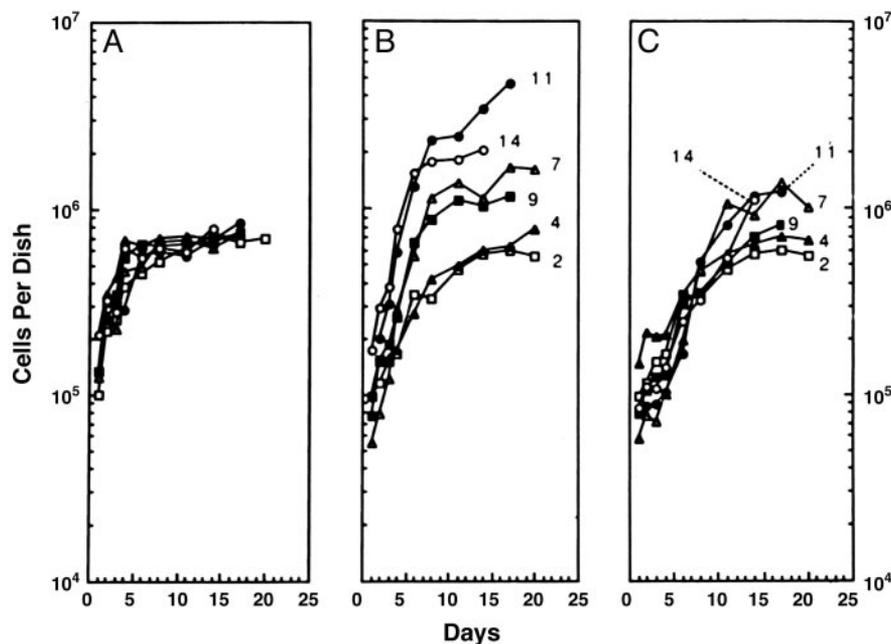
Because lineage 1 of the 2° assay of the [10-3] group exhibited many dense foci (Fig. 3), it was terminated at that point. Based on the presence of some dense foci in the 2° assay of the other lineages of the [10-3] group, it was anticipated that a straight 3° assay of  $10^5$  cells might give too many overlapping foci to discern their individual morphology. Therefore,  $10^4$  cells of the 2° assay were mixed with  $10^5$  LDP stock cells to make a uniform monolayered background against which discrete foci could be displayed. The foci of lineages 2 and 3 from the  $10^5$  cell 3° assay were larger and denser than those from the  $10^4$  cell assay, whereas light foci from the  $10^5$  cell assay of lineage 2 disappeared entirely in the  $10^4$  cell assay. This difference in focus formation indicated that the nonfocus-forming background of the straight assay of  $10^5$  cells, which consisted of cells that had undergone the same two rounds of confluence as their sibling focus formers, was more permissive to the development of foci than were the LDP stock cells that formed the background for the  $10^4$  cell assay.

The promoting effect of increased saturation density on the

development of transformed foci suggested that the development of dense foci might preferentially occur within broad light foci if the latter contained enough cells to constitute a significant fraction of the entire population. In Fig. 4, such foci occupy a large fraction of the surface of a culture at 14 days, and some of them display the punctate beginnings of dense foci (10). Given another 7 days to develop, these foci within foci have become more prominent and, in several cases, they are coalescing to form large dense foci, whereas no such changes appear in the flat background that surrounds the light foci. These observations visually confirm that continued proliferation at confluence



**Fig. 4.** Origin of dense foci within light foci. The original 1987 frozen stock of NIH 3T3 cells was thawed and cultured for 3 days in 10% CS before subculture of  $10^5$  cells for the 1° assay in 2% CS. The culture was fixed and stained at 14 days (Left) and at 21 days (Right). Broad very light foci are visible in the 14-day culture, a few of them containing tiny groups of densely stained cells. The light foci have become slightly darker in the 21-day culture and contain multiple darkly stained punctate foci that in some cases are coalescing to form large dense foci.

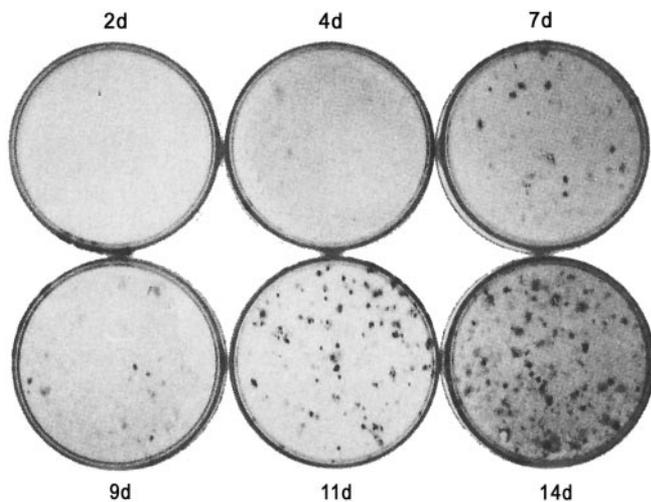


**Fig. 5.** Rates of increase in saturation density during LDP in 2% CS vs. undisturbed maintenance in 2% CS. Groups of cultures were maintained in LDP in 10% CS (A), LDP in 2% CS (B), and undisturbed maintenance in 2% CS (C). At the intervals in days indicated at the curves in B, the cells in A and B were serially subcultured at low density. On the same days, as indicated in C, two cultures were trypsinized. At each interval,  $10^5$  cells from each group were seeded in 2% CS and counted for their saturation density at 2 wk.

engenders progressive neoplastic development. If it is assumed that the light foci contain no more than 1,000 cells, and at least 10 dense focal areas arise within each light focus, then the rate of progression of this step is  $10^{-2}$  or higher per cell division.

**Transformation in LDP with Low CS.** Variation of transformed focus formation among cultures within groups, which included their size, density, and number as well as the time of their appearance (Fig. 3), suggested they might arise by the induction of new

mutations rather than by progressive selection of spontaneous variants that accounted for the increases in saturation density (9). That suggestion was reinforced by the absence of transformed foci in three of the four groups until the 3<sup>o</sup> or 4<sup>o</sup> assay and by the later observation that all of the cells recovered after long-term confluence had heritable reductions in rate of proliferation in LDP (11, 12), indicative of chromosomal damage that might transform a small fraction of them. However, increases in saturation density occurred in LDP ( $2-5 \times 10^4$  cells three times per wk in 60-mm dishes) when the CS concentration was reduced from 10% to 2% and was more efficient than with prolonged confluence in 2% (Fig. 5) (13). In parallel with the increases in saturation density, transformed foci appeared in 1<sup>o</sup> assays made from the LDP in 2% CS (Fig. 6) (14). The cells proliferated exponentially in the LDP, albeit at a 20% lower rate than in 10% CS (15), with no indication of heritable damage (16). Further stepwise reduction of CS in LDP to 1% resulted in an increase in the size and density of the foci, which was more apparent when the assays were done in 1% rather than 2% CS (Fig. 7). A single large step-down from 10% to 1% CS in LDP was much less effective in producing transformed foci than an intermediate multipassage step through 2% CS to 1% CS. Multistep reduction to 0.25% CS followed by many passages in 0.25% CS resulted in exponential proliferation and high saturation density with transformation at that very low CS concentration, which ordinarily supports no detectable proliferation in the absence of selection. These results indicate that the production of foci occurs by the same process as the increases in saturation density but involves further selection of heritable variants that are continuously generated in cultures.



**Fig. 6.** Appearance of transformed foci in 1<sup>o</sup> assay initiated by cells after each LDP in 2% CS. The cells were the same as those used in Fig. 5B, in which LDP of cells were made at 2, 4, 7, 9, 11, and 14 days, and  $10^5$  cells of each passage were seeded in 2% CS for 2 wk. The photograph shows sister cultures to those in Fig. 5B, but they are fixed and stained to display transformed foci rather than trypsinized and counted for saturation densities. [Reproduced with permission from ref. 14 (Copyright 1990, American Association for Cancer Research).]

## Conclusions

1. The degree of neoplastic transformation is measured here by increases in saturation density of NIH 3T3 cell cultures and the appearance of transformed foci in various sizes, densities, and numbers in standardized serial assays at confluence.



