

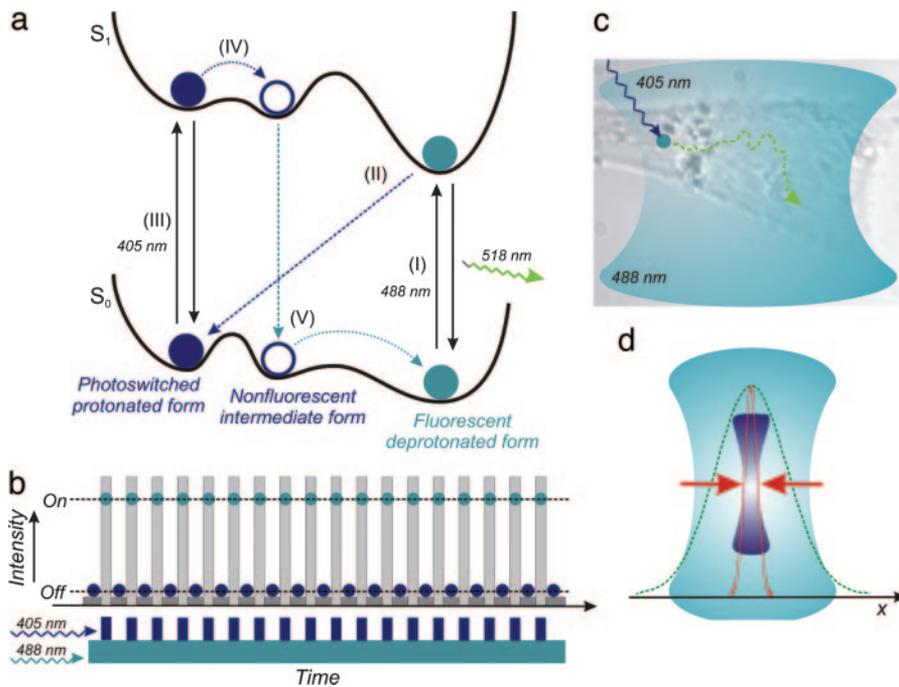
# Reversible molecular photoswitches: A key technology for nanoscience and fluorescence imaging

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In recent years, molecular switches have attracted considerable interest because they hold great promises as molecular electronic and photonic devices. In contrast to commonplace switches that turn electric appliances on and off, molecular switches enable the storage of information on a molecular level, and their application in nanotechnology, biomedicine, and computer chip design opens up whole new horizons. Usually, molecular switches are addressed by an electrical field, a scanning tunneling microscope tip, or a chemical or electrochemical reaction to specifically switch the physical properties between two states (1–3). Alternatively, molecules might be switched optically between two stable forms, and quite an effort has been put into the synthesis of reversibly photoswitchable fulgides and diarylethenes (4). A photoswitch exhibits two stable and selectively addressable states, a fluorescent and a nonfluorescent, which can be conveyed into another in a reversible fashion upon irradiation with different wavelengths of light. Although highly reproducible optical switching of individual chromophores could be achieved in liquid helium temperature experiments (5), the synthetic approach based on a photoswitchable diarylethene derivative was crowned with success only recently (6, 7). Very recently (8, 9), it was demonstrated that even conventional unmodified carbocyanine derivatives such as Cy5 can function as efficient reversible single-molecule photoswitches.

In this issue of PNAS, Habuchi *et al.* (10) report about photoswitching of a mutant of a GFP-like fluorescent protein that was cloned from the coral *Pectiniidae*. They demonstrate reversible photoswitching between dim and bright states of individual molecules embedded in poly(vinyl alcohol) by using 488 and 405 nm laser light with a response time in the millisecond range and a repeatability of >100 times (Fig. 1). So far, only a few successful reversible photoswitching events have been reported for a GFP mutant with a response time of several minutes (11). The intriguing switching performance of the new GFP-like fluorescent protein led to the naming “Dronpa,” after “dron,” a ninja term for vanishing, and “pa,” which stands for photoactivation (12). Because GFP and its fluorescent homologous provide direct genetic encod-



**Fig. 1.** Molecular photoswitches might be advantageously used for the study of protein trafficking in living cells and ultrahigh-density optical data storage at the single-molecule level. (a) Schematic of the operating mode of the reversible and highly reliable GFP-like photoswitch Dronpa. Upon excitation of the deprotonated form at 488 nm, it converts into the ground state of the photoswitched protonated form with a switching yield  $\Phi_{SW}$  of  $3.2 \times 10^{-4}$  (II). Excitation of the protonated form at 405 nm (III) results in excited state proton transfer to an intermediate state (IV), which converts into the deprotonated ground state with a switching yield  $\Phi_{SW}$  of 0.37 (V). (b) Using simultaneous illumination at 488 nm and applying a 30-ms pulse of 405-nm laser light every second, Habuchi *et al.* (10) achieved up to 170 photoswitching events at the single-molecule level with high reliability. (c) Fluorescence of tagged proteins can be erased by applying 488-nm light and switched on at specific locations by using 405-nm irradiation to directly visualize protein movement routes. (d) For nanoscale writing, a 405-nm laser beam could be focused to a diffraction-limited focal spot to induce photoswitching to the bright deprotonated form. A second 488-nm laser beam could switch the molecules at the outer parts of the focus back to the dim protonated form by using a phase mask.

ing of strong visible fluorescence, they have become invaluable tools for *in vivo* imaging of cells and tissue (13, 14). In combination with photobleaching techniques, fluorescent proteins can also provide information on the movement of intracellular proteins. However, photobleaching techniques do not allow direct visualization of protein movement routes within a living cell. Therefore, photoswitchable fluorescent proteins such as Dronpa are strongly desired for improved protein tracking in living cells (14, 15).

The fact that most fluorescent proteins known nowadays exhibit slow response time to light and irreversible switching behavior highlights the importance of the discovery of Dronpa, a genetically encoded monomeric fluorescent protein that can be addressed reversibly for repeated

measurements of protein behavior. Similar to GFP derivatives and other GFP-like proteins, Dronpa shows two absorption peaks, a major peak at 503 nm and a minor peak at 388 nm, which can be attributed to the deprotonated and the protonated form of the chromophore, respectively. Although the deprotonated form shows bright fluorescence at 518 nm with a quantum yield  $\Phi_f$  of 0.85 and a monoexponential fluorescence decay time of 3.6 ns, independent of the detection wavelength deactivation of the excited state of the protonated form is dominated by nonradiative relaxation. Therefore, the

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