

# Natural selection in action during speciation

Sara Via<sup>1</sup>

Departments of Biology and Entomology, University of Maryland, College Park, MD 20742

The role of natural selection in speciation, first described by Darwin, has finally been widely accepted. Yet, the nature and time course of the genetic changes that result in speciation remain mysterious. To date, genetic analyses of speciation have focused almost exclusively on retrospective analyses of reproductive isolation between species or subspecies and on hybrid sterility or inviability rather than on ecologically based barriers to gene flow. However, if we are to fully understand the origin of species, we must analyze the process from additional vantage points. By studying the genetic causes of partial reproductive isolation between specialized ecological races, early barriers to gene flow can be identified before they become confounded with other species differences. This population-level approach can reveal patterns that become invisible over time, such as the mosaic nature of the genome early in speciation. Under divergent selection in sympatry, the genomes of incipient species become temporary genetic mosaics in which ecologically important genomic regions resist gene exchange, even as gene flow continues over most of the genome. Analysis of such mosaic genomes suggests that surprisingly large genomic regions around divergently selected quantitative trait loci can be protected from interracial recombination by “divergence hitchhiking.” Here, I describe the formation of the genetic mosaic during early ecological speciation, consider the establishment, effects, and transitory nature of divergence hitchhiking around key ecologically important genes, and describe a 2-stage model for genetic divergence during ecological speciation with gene flow.

divergence hitchhiking | ecological speciation | reproductive isolation

The origin of species is only slightly less mysterious now than it was 150 years ago when Darwin published his famous book (1). Although Darwin’s idea that natural selection drives speciation has finally been widely accepted (2), we still have much to learn about the nature and time course of the genetic changes that cause speciation under natural selection (3–5).

## The Spyglass

Pivotal ideas developed during the modern synthesis of the 1930s–1940s have largely determined the course of modern speciation research. Ernst Mayr (6) developed the biological species concept, putting reproductive isolation at the center of speciation and making analysis of the evolution of reproductive isolation a clear target for speciation research. Mayr also stressed that the evolution of reproductive isolation is a fragile process that can only proceed if geographical separation renders gene flow impossible, firmly establishing allopatric speciation as the norm. Theodosius Dobzhansky (7) identified a wide array of traits that could cause reproductive isolation, but focused much of his own research into speciation on postzygotic genetic incompatibilities (8), as did H. J. Muller (9). At the time, hybrid sterility was a particularly problematic aspect of speciation because it had been unclear since Darwin (1) how such a disadvantageous trait could evolve under natural selection. By providing a clear mechanism by which hybrid sterility could evolve (ref. 2, pp. 269 and 270), Dobzhansky-Muller genetic incompatibilities (DMIs) took center stage in the genetic analysis of speciation, where they have remained ever since. Collectively, the architects of the synthesis outlined a retrospective approach to speciation that I call “the spyglass,” because it starts late in the process (or after it is complete) and looks back in time to infer the causes of speciation (Fig. 1A).

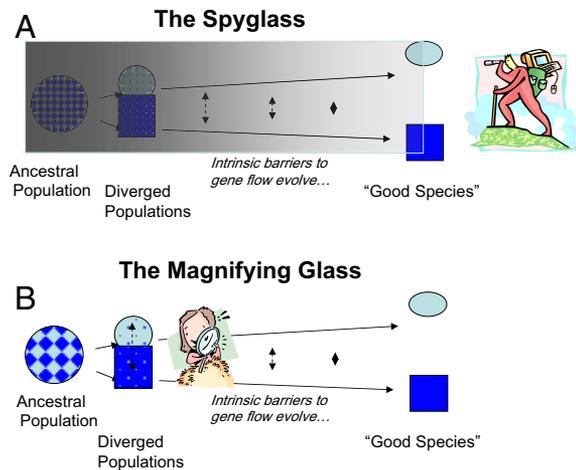


Fig. 1. Two ways to study the process of speciation, which is visualized here as a continuum of divergence from a variable population to a divergent pair of populations, and on through the evolution of intrinsic barriers to gene flow to the recognition of good species. (A) Using the spyglass, the process is studied by attempting to look back to see the details of speciation from the vantage point of the present. (B) Using the magnifying glass, the mechanisms of reproductive isolation are studied in partially isolated divergent ecotypes that are used as models of an early stage of speciation.

Unquestionably, this approach has been a rich source of information about the kinds of barriers to gene flow that can isolate species (e.g., refs. 10 and 11), but alternative ideas about speciation and how to study it have met with considerable resistance during the past 70 years. Even today, allopatric speciation remains the null model against which all other mechanisms for speciation must be tested (ref. 2, p. 158), and DMIs are widely regarded as the appropriate focus of research in speciation genetics (12–16).

## The Magnifying Glass

A different view of speciation genetics is now gaining in popularity: the population-level analysis of how ecology and genetics interact in various situations to cause the evolution of barriers to gene flow (4, 17, 18). By analyzing partially reproductively isolated ecotypes or races, the genetic changes contributing to reproductive isolation can be studied before they become confounded by additional genetic differences between species that accumulate after speciation is complete. Indeed, studying barriers to gene flow in populations that are not yet completely reproductively isolated may reveal important aspects of the process that have never been seen clearly before. This approach is particularly suitable for the analysis of speciation under divergent selection, now called “ecological speciation” (4). To contrast this approach with the more classic retrospective analyses, I call the population-level analysis of the

This paper results from the Arthur M. Sackler Colloquium of the National Academy of Sciences, “In the Light of Evolution III: Two Centuries of Darwin,” held January 16–17, 2009, at the Arnold and Mabel Beckman Center of the National Academies of Sciences and Engineering in Irvine, CA. The complete program and audio files of most presentations are available on the NAS web site at [www.nasonline.org/Sackler.Darwin](http://www.nasonline.org/Sackler.Darwin).

Author contributions: S.V. designed research, analyzed data, and wrote the paper.

The author declares no conflict of interest.

This article is a PNAS Direct Submission.

<sup>1</sup>E-mail: [svia@umd.edu](mailto:svia@umd.edu).

ecological and genetic causes of reproductive isolation the “magnifying glass” (Fig. 1B).

The validity of population-level analyses of the ecology and genetics of the partial reproductive isolation has occasionally been questioned. Because there is no guarantee that ecotypes or races will ever attain species status, some argue that barriers to gene flow between them caused by divergent selection are irrelevant to the study of speciation (2, 19). Others disregard ecologically based reproductive isolation between ecotypes because it lacks permanence and could be reversed if the pattern of divergent selection changes (2). However, many valid species concepts do not require or even consider permanence, and doing so simply underscores the observation that different species concepts apply at different points along a continuum of divergence from populations to well-established and permanent species (20).

To supporters of the population-level approach, divergent populations are an early stage on that continuum. They argue that barriers to gene flow in partially isolated ecological races or ecotypes must be similar to those that would have been seen long ago between a pair of present-day sister species in a similar ecological situation. Although it is certainly true that not all divergent races will go on to become full species, many contemporary species must have passed through the stage of population divergence typified by ecotypes. Because the particular ecological conditions associated with divergence under selection cannot be seen clearly through the spyglass, that approach is unlikely to reveal much about the initial causes of reproductive isolation during speciation under divergent selection.

Speciation research is plagued by this awkward gulf between population-level and species-level analyses. To fully understand the mechanisms of speciation, we must cross that space, integrating magnifying glass analyses of the early parts of the process with the view of the end products through the spyglass.

### How Does Natural Selection Cause Speciation?

With the possible exception of reinforcement, natural selection does not directly favor phenotypic traits or genetic incompatibilities simply because they block gene flow. Instead, it is generally thought that reproductive isolation occurs indirectly, as a “by-product” of the genetic changes that increase adaptation (ref. 2, p. 385). However, pooling ecologically based reproductive isolation with that caused by the accumulation of genetic incompatibilities under the term by-product does not do much to advance our mechanistic understanding of speciation. To fully connect the views through the magnifying glass and the spyglass, we must understand how different ecological situations lead to particular types of barriers to gene flow and on what timetable these different forms of reproductive isolation evolve. It is particularly important to ask under what circumstances and how often ecologically based reproductive isolation produce virtually complete reproductive isolation before appreciable genetic incompatibility has evolved (ref. 2, pp. 57–59, and ref. 21).

### The Effect of Geography on Speciation

Ecological speciation can occur in either geographically isolated populations (allopatry) or in settings with no physical barriers to gene flow (sympatry or parapatry). When gene exchange is physically impossible, the conditions under which reproductive isolation can evolve are nonrestrictive: allopatric speciation can be driven by strong or weak divergent selection, sexual selection, uniform selection, or even stabilizing selection. It may occur quickly under divergent selection or extremely slowly under uniform or balancing selection.

In contrast, the conditions under which sympatric or parapatric speciation with gene flow can occur are not so forgiving: genetically based phenotypic divergence requires much stronger selection to occur and be maintained when gene flow is possible than when geography makes it an impossibility (e.g., refs. 17 and 22). In the

presence of migration, the establishment of genomic regions that resist gene flow sufficiently to maintain phenotypic differentiation is only likely if divergent (or possibly sexual) selection is strong, and so the initial barriers to gene flow in sympatry are likely to evolve quickly (17, 22, 23). Speciation with gene flow (22) is thus unlikely to occur under weak divergent selection, and it is certainly not expected under uniform or balancing selection (except perhaps by polyploidy). One fortuitous effect of the strong selection required for speciation with gene flow is that the genomic regions that cause reproductive isolation become particularly distinctive relative to the rest of the genome. This facilitates their discovery in empirical analyses.

Until recently, genetic models of speciation with gene flow have been extremely simplified, and they have suggested quite a restrictive set of conditions for sympatric speciation. In particular, critics cite the difficulty of evolving assortative mating in the face of free recombination (ref. 2, pp. 127–141). Although Felsenstein (24) showed that this constraint is significantly reduced under linkage between genes affecting performance and mating, that result has been largely ignored (17).

A variety of conditions that facilitate ecological speciation with gene flow are now well described (17, 22). They include strong divergent selection on multiple traits associated with resource or habitat use and ecologically based selection against migrants and/or hybrids. Recent work suggests that assortative mating can evolve rather easily if habitat choice determines the choice of mates (17, 22), if mate choice is a correlate of the traits under divergent selection (4), or if recombination is reduced by physical linkage or pleiotropy (24, 25). Some of the best-studied divergent races in the wild, including the ecologically specialized host races of the pea aphid [*Acyrtosiphon pisum* (Harris)] satisfy these conditions (17), providing strong empirical support for the argument that reproductive isolation can evolve, or at least persist, in the face of gene flow.

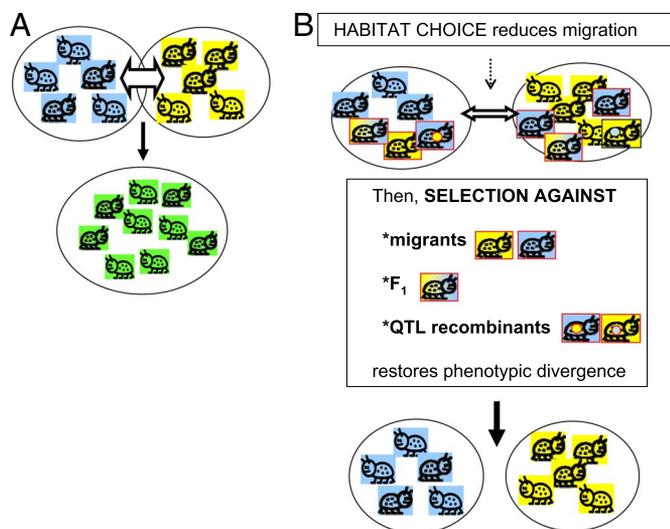
The first step in using the magnifying glass to study speciation involves estimating the magnitude of gene flow between a pair of partially reproductively isolated taxa. That process is not as straightforward as it may seem.

### Potential vs. Realized Gene Flow During Ecological Speciation Under Divergent Selection

Empirical estimates of gene flow assume neutrality of markers and a balance between migration and random drift (e.g., ref. 26). This emphasis on gene flow under neutrality implies that there is just 1 “true” estimate of gene exchange between 2 taxa. Moreover, the minimal gene flow required to counter drift under Wright’s island model (ref. 26, pp. 194 and 195) conjures up a picture of gene flow as a force that will easily homogenize adjacent populations (Fig. 2A). Yet, one can easily find phenotypically divergent and ecologically specialized populations living in close adjacency with no physical barrier to gene flow. Is this a contradiction? Not necessarily, because the degree to which a genomic region affecting a given trait is homogenized by migration depends not on the estimated gene flow under neutrality, but on the realized gene flow at that region after selection.

It is a maxim of population genetics that migration and genetic drift affect the entire genome, whereas the effects of natural selection are limited to genomic regions harboring loci that affect the selected phenotypic traits. In ecologically specialized populations, divergent selection on traits associated with the use of resources or habitats is strong enough to maintain divergence in the parts of the genome that affect those traits, while gene flow continues in other genomic regions (Figs. 2B and 3). Between such populations, the estimated gene flow under neutrality may grossly overestimate the realized gene exchange experienced by divergently selected genomic regions.

Because of the localized genomic effects of divergent selection on realized gene exchange, divergence early in ecological speciation

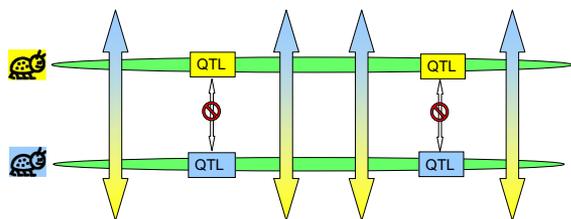


**Fig. 2.** Gene flow between sympatric populations does not necessarily prevent differentiation. (A) The simple vision of gene flow as a homogenizing force in populations. Free migration and random mating between locally adapted populations is simplistically visualized as homogenizing populations and eradicating local adaptation. (B) Ecologically based barriers to gene flow that evolve under divergent selection permit genetically based phenotypic divergence under selection to be maintained in the face of gene flow: habitat choice reduces migration, then selection against migrants,  $F_1$ , and QTL recombinants reduces introgression of locally adapted alleles, maintaining divergence.

with gene flow is expected to be greater in genomic regions that harbor key quantitative trait loci (QTL) than it is in regions that have no effect on the phenotypic divergence of the populations (Fig. 3). If, as usually thought, speciation with gene flow involves only a handful of characters (22), these divergent regions may comprise a relatively small fraction of the genome, leaving the genomes of incipient species largely homogenized by ongoing gene flow and “profoundly genetically similar” (Fig. 3 and ref. 27). We call the resulting pattern of genomic heterogeneity in divergence and gene exchange early in ecological speciation with gene flow “the genetic mosaic of speciation” (3).

### Using $F_{st}$ Outlier Analysis to Identify Selected Genomic Regions in the Genetic Mosaic

Wright’s  $F_{st}$  is a widely used measure of genetic divergence between populations (26). In 1973, Lewontin and Krakauer (28) proposed



**Fig. 3.** A diagram version of genomic heterogeneity of gene exchange and genetic divergence under divergent selection. One chromosome from each specialized parental population is shown, with the genomic regions that contain locally adapted genes (QTL) indicated in boxes. The color of each QTL corresponds to the specialized parent from which it came. Bidirectional arrows indicate gene exchange, with the color of the arrow tips showing that alleles from the other population are introgressing. Gene exchange can occur outside the regions containing the specialized genes, but is blocked from occurring within those regions for the reasons shown in Fig. 2B. This heterogeneous pattern of gene exchange under selection establishes the genetic mosaic of speciation in which genetic divergence is restricted to divergently selected portions of the genomes, whereas other regions share polymorphisms freely through ongoing gene exchange.

that genetic markers with aberrantly high  $F_{st}$  values (“outliers”) could be inferred to be affected by divergent selection. Recently, outlier analyses have enjoyed a renaissance because of the development of new methods based on coalescent simulation (29, 30).  $F_{st}$  variation is still used to eliminate selected markers from population genetic analyses that require neutrality (31), but it is now of primary interest as a signature of divergent selection that permits detection of genomic regions involved in adaptive divergence (32, 33).

Significant heterogeneity in marker  $F_{st}$  has been documented between ecologically divergent races (34–38), with the interpretation that  $F_{st}$  outliers must be linked to loci affecting the phenotypic traits known to distinguish the divergent ecotypes, races, or subspecies. However, outlier analysis alone cannot reveal the cause of deviant  $F_{st}$  values, and the conclusion that  $F_{st}$  outliers must be associated with genes causing the most obvious differences between ecotypes is premature.

Using a linkage map that shows locations of the QTL affecting phenotypic traits known to be under divergent selection, the hypothesis that outliers are linked to key phenotypic traits under divergent selection can be tested: Are  $F_{st}$  outliers scattered randomly on the map, or are they clustered near divergently selected QTL? This approach requires a system in which the phenotypic traits involved in divergence and reproductive isolation are well known, the strength of selection on these traits has been measured, and QTL affecting the key traits have been localized on a linkage map, so that mapped markers can be used in an  $F_{st}$  analysis of field-collected samples. The pea aphid host races on alfalfa and red clover [*A. pisum pisum* (Harris)] are such a system.

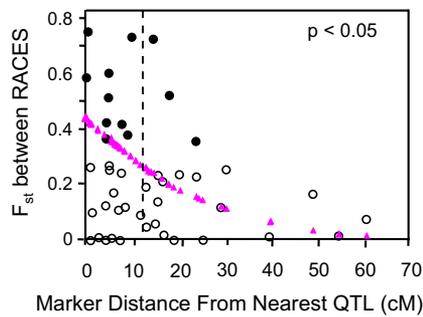
### Pea Aphids on Alfalfa and Red Clover: Ecological Speciation in Action?

The pea aphid complex is a worldwide group of phloem-feeding insects, found primarily on legumes (39). Although the pea aphid host races on alfalfa and red clover are in the same subspecies, *A. pisum pisum*, sympatric pea aphid populations on alfalfa, red clover, and other legumes are highly genetically divergent and ecologically specialized in the eastern United States (40), Europe (41–44), and South America (45).

Experimental studies in both the field and laboratory have documented extensive ecologically based reproductive isolation between the pea aphid host races in eastern North America because of strong selection against migrants to the alternate host (40, 46–48), environmentally mediated selection against hybrids (49), and habitat choice (49). It is unknown whether divergence between the pea aphid host races began in sympatry (e.g., ref. 2, pp.163 and 164). However, conditions of the initial split are far less relevant to the study of speciation than is the fact that divergent selection currently maintains genetically based phenotypic differentiation and significant ecologically based reproductive isolation between sympatric populations.

Via and West (3) estimated  $F_{st}$  between the pea aphid host races for 45 markers with known locations on a QTL map of genomic regions affecting early fecundity and behavioral acceptance of each plant. They then estimated the map distance from each marker to the nearest QTL for one of the key host use traits and found that  $F_{st}$  outliers were significantly clustered around the QTL involved in reproductive isolation ( $P < 0.05$ ; Fig. 4).

Surprisingly, the spatial distribution of the mapped  $F_{st}$  outliers suggests that the signature left by divergent phenotypic selection on neutral markers can extend far from major QTL: the average outlier was 10.6 cM from the nearest QTL. A similar result was found for traits involved in the divergence of whitefish morphs in postglacial lakes (50), where the average outlier was 16.2 cM from the nearest QTL. In both systems, the hitchhiking regions around divergently selected QTL are far larger than expected. After a selective sweep through a large panmictic population, linkage disequilibrium rapidly erodes except in areas of reduced recombination (51). Although reduced recombination is, in fact, the explanation for the



**Fig. 4.** The relationship between the  $F_{st}$  values of amplified fragment length polymorphism markers, and their map distance from the nearest QTL involved in reproductive isolation between pea aphids on alfalfa and red clover.  $F_{st}$  outliers are shown as solid circles, and the dotted line at 10.6 cM marks the average distance from an outlier to the nearest QTL. The pink triangles are the predicted values from a logistic regression of the probability that a marker was an outlier on its distance to the nearest QTL. [Figure has been modified and reproduced with permission from ref. 3 (2008, Wiley-Blackwell, Oxford, United Kingdom).]

large hitchhiking regions we observed, they arise not from suppression of recombination *per se*, but from a reduction in the “effective recombination” between locally adapted QTL alleles in divergent populations that are subdivided (i.e., no longer randomly mating).

In 1997, Charlesworth et al. (52) analyzed the potential for hitchhiking when populations are subdivided by divergent (local) selection. Using a simulation analysis, they found that regions of linkage disequilibrium of the size we observed (3) can be maintained around a divergently selected locus. This unexpected result occurs because subdivision reduces the opportunity for recombination between locally adapted QTL alleles. In other words, divergent selection leads to fewer than expected interpopulation matings, which reduces the effective recombination of locally adapted QTL alleles below what is expected based on a map from a controlled cross.

Because speciation is, by definition, a process in which populations become increasingly subdivided, reduced effective recombination under subdivision is an important aspect of speciation with gene flow, although it has been largely unappreciated. As populations become subdivided by divergent selection and ecological specialization increases, the opportunity for interracial recombination is increasingly reduced by selection against migrants, extrinsic selection against  $F_1$  hybrids, and/or habitat choice. When recombination between QTL alleles does occur, the local disadvantage of the recombinant QTL allele further reduces the frequency of migrant alleles in advanced generation hybrids or backcrosses.

So, despite free recombination within subpopulations, the effective recombination between genotypes with different QTL alleles begins to decline as soon as divergent selection foils panmixia in early ecological speciation with gene flow, and it may also be a significant factor maintaining divergence after secondary contact. To emphasize that this form of hitchhiking is maintained only in divergent populations, and that it differs from hitchhiking after a selective sweep, I call it “divergence hitchhiking” (3).

The reduction in effective recombination between divergent races means that the nominal map distance estimated from  $F_{st}$  outliers to the nearest QTL (Fig. 4) overestimates the actual probability that recombination will separate a divergent outlier allele from a locally adapted allele at a nearby QTL. In other words, the “effective map distance” between a locally adapted QTL and a nearby marker is much less than that estimated from a linkage map made using controlled matings.

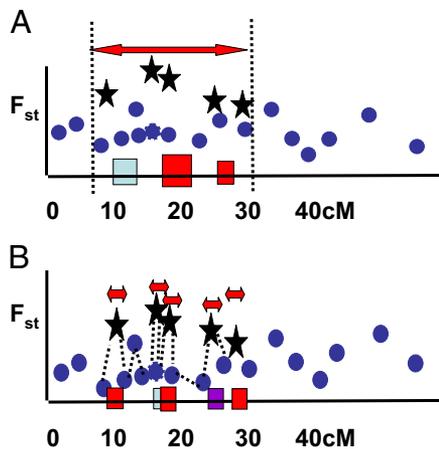
We can empirically approximate the effective marker–QTL distance in pea aphids by estimating the extent to which habitat choice and divergent selection limit the opportunity for interracial

mating and recombination (e.g., Fig. 2B). We used field sampling to estimate that habitat choice reduces migration to the alternate host to  $\approx 11\%$  (suggesting that 89% of possible migrants reject the alternate host plant; ref. 49). The fitness of migrants from alfalfa to clover is  $\approx 30\%$  of that of nonmigrants, whereas the fitness of migrants from clover to alfalfa is only  $\approx 5\%$  of that expected for nonmigrants, for an average relative fitness of migrants of  $\approx 17\%$  (estimated selection against migrants of  $s = 0.83$ ; ref. 49).  $F_1$  hybrids do not feed as effectively on either host as the parental specialist, which reduces their fecundity (and the realized number of recombinations) by  $\approx 50\%$  (49). So, without accounting for sexual selection against migrants, the probability of recombination between races for a marker 10.6 cM from a QTL is the original recombination probability (0.106) discounted by the probability that a migrant will choose the alternate habitat and survive there ( $0.11 \times 0.17$ ), and the relative fecundity of  $F_1$  hybrids (0.5), making the effective recombination rate for that marker  $(0.106) \times (0.11) \times (0.17) \times (0.5) = 0.001$ .

This calculation suggests that the average outlier, at a nominal distance of 10.6 cM from the nearest QTL on the linkage map (3), has an effective map distance to that QTL of only  $\approx 0.1$  cM. In these populations, even a marker 50 cM from a divergently selected QTL has an effective map distance of only 0.5 cM as a result of the large decrease in effective migration caused by extensive ecologically based reproductive isolation. This estimated difference between nominal recombination and effective recombination between subdivided populations shows why such a large genomic region is expected to remain in linkage disequilibrium with a QTL under divergent selection during ecological speciation with gene flow.

It is of interest that the magnitude of effective recombination in a given genomic region changes over time as ecological specialization evolves, because greater specialization increases the magnitude of resource-based selection against migrants and hybrids. This can be illustrated with a simple example. Imagine that early in divergence only a few QTL have differentiated such that extent of habitat choice and the disadvantage of migrants or  $F_1$  is only 25% as strong as at present. Then, only 22% of potential migrants would refuse the alternate host (78% accept), selection against a migrant would be  $s = 0.21$  (relative fitness of migrants = 0.79), and the relative fecundity of an  $F_1$  hybrid would be 0.875, making the effective rate of recombination of the average outlier with the nearest QTL  $(0.106) \times (0.78) \times (0.79) \times (0.875) = 0.057$ . Thus, at this earlier point in divergence, the average outlier would have an effective map distance of 5.7 cM from the nearest QTL. Although smaller than the nominal map distance of 10.6 cM, it is far from the tight effective linkage seen at present. The size of each region of divergence hitchhiking therefore depends not only on the strength of divergent selection directly on that genomic region, but also on the extent to which effective migration is reduced by the earlier divergence of other QTL alleles throughout the genome.

Divergence hitchhiking has the same general effect on interracial recombination and speciation as a chromosomal inversion that happens to contain 1 or more key QTL (e.g., refs. 53–55). However, unlike inversions, which must occur relatively infrequently at the site of key QTL, divergence hitchhiking appears automatically around any QTL under strong divergent selection. Moreover, regions of divergence hitchhiking are dynamic. They increase in size as the evolution of specialization reduces the effective migration between diverging races, and regions of divergence hitchhiking around loosely linked QTL may overlap and merge. Perhaps most importantly, however, regions of divergence hitchhiking leave no permanent signature because they do not involve physical alterations to chromosomes. These regions of reduced interracial recombination can only be detected while divergence elsewhere in the genome is low. They will not be seen in retrospective analyses of good species, because as speciation progresses they become assimilated into the overall genomewide pattern of genetic divergence and by the time speciation is complete, they have disappeared.



**Fig. 5.** Two views of how to delineate regions of divergence hitchhiking. (A) A region of divergence hitchhiking around several QTL is defined by the region covered by a cluster of  $F_{st}$  outliers. (B) Each outlier is within a separate hitchhiking region, bounded by a low  $F_{st}$  marker. Outliers are thought to be either under direct divergent selection or tightly linked to a selected gene. The double-headed arrows show the hypothesized extent of divergence hitchhiking. Stars denote  $F_{st}$  outliers and circles denote markers with  $F_{st}$  in the expected range. Boxes show the map location of QTL under divergent selection, color-coded by the trait they affect, and with box size corresponding to effect size. Dashed lines either mark the boundaries of the hitchhiking region (A), or connect the  $F_{st}$  values of adjacent markers (B).

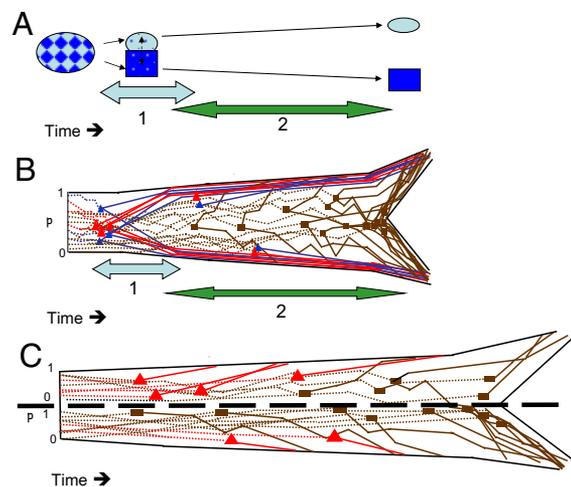
### How Many QTL Are There Within a Region of Divergence Hitchhiking?

Hawthorne and Via (25) found that QTL for different traits under divergent selection in the pea aphid host races tended to colocalize on the linkage map. Colocalization of QTL increases selection experienced by that genomic region, thereby increasing the size of the region of divergence hitchhiking, and facilitating both QTL detection and the accumulation of additional QTL. In addition, any given QTL may actually be a cluster of several genes. Thus, it seems likely that most regions of divergence hitchhiking will contain multiple genes that affect 1 or more traits under divergent selection.

**How Are Regions of Divergence Hitchhiking Delineated?** Determining the size of a given region of divergence hitchhiking is not entirely straightforward. There are 2 contrasting views:

**I. A Single Region of Divergence Hitchhiking Extends Across a Cluster of  $F_{st}$  Outliers Around a Given QTL or Group of QTL (Fig. 5A).** Via and West (3) proposed that a cluster of outliers defines a single region of divergence hitchhiking, which may include 1 or more QTL. They suggested that the boundaries of a given region of divergence hitchhiking be estimated by curve fitting in a genome scan of  $F_{st}$  values at various map distances around individual QTL under divergent selection (Fig. 5A). In this view, markers with low  $F_{st}$  values that lie within hitchhiking regions are interpreted as polymorphisms that predate divergence at the QTL. They are thus uninformative about population divergence and should not be used to mark the boundaries of divergence hitchhiking.

**II. Each Outlier Corresponds to a Gene or QTL Under Selection or Is Itself Under Selection (Fig. 5B).** Ting et al. (12) found that a DNA sequence just 1,100 bp away from the hybrid sterility gene *Odysseus* (*Ody*) was not divergent between the 2 parental species, and from this single observation they concluded that the hitchhiking region around *Ody* must be extremely small. Wood et al. (56) and Smadja et al. (57) extend this idea by suggesting that the nearest genomic region of low genetic divergence to an  $F_{st}$  outlier marks the boundary of its hitchhiking region (Fig. 5B).



**Fig. 6.** Visualizing the pattern of genetic divergence during ecological speciation. (A) The 2 stages of ecological speciation with gene flow. Stage 1 is noted by the blue arrow, and stage 2 is indicated by the green arrow. Events during each stage are as described in the text. (B) Diagram of gene trees within a species tree for ecological speciation with gene flow. Each gene is polymorphic in the original population, with a frequency ( $p$ ) as noted on the axis at the left of the drawing. As time goes on, some loci quickly diverge at about the same time under divergent selection. The gene frequency at a given locus is shown as a dotted line before it diverges between the incipient species (at the time marked by the symbols), and then as a solid line. Red symbols are selected QTL, and blue symbols are divergence hitchhikers. This divergence is mostly complete by the end of stage 1. During stage 2, a handful of additional loci diverge under divergent selection (red), and loci that were unaffected by divergent selection diverge by independent responses to uniform or balancing selection or by drift (brown symbols). (C) Gene trees in a species tree under allopatry. Symbols are as in B. The heavy dashed line indicates a geographical barrier to gene flow. If an allopatric population enters a new environment, there may be a period of rapid response to divergent selection similar to stage 1 in A. Otherwise, as shown here, genes diverge over a long time period under any combination of divergent selection, independent responses to uniform or balancing selection, or drift, eventually all coming into concordance to produce the branching pattern of the new species.

Several observations are inconsistent with this hypothesis. First, outliers are easy to find. Even in studies with just a few markers (3, 34, 35, 50), 5% or more of tested markers are generally  $F_{st}$  outliers. Taking 5% as a minimal estimate, this observation implies either that hitchhiking regions are large enough that they capture 5% of randomly chosen markers (3), or that so many genes are under divergent selection that 5 of them can be found with only 100 markers (56). If the latter were true, we would expect that in an average-sized genome of  $\approx 25,000$  genes,  $\approx 1,250$  of them (5%) would be involved in ecotypic differentiation and early speciation. This seems unlikely, given the prevailing view that speciation with gene flow typically involves just a handful of traits, each influenced by just a few major genes (22). Second, even with very strong phenotypic selection during population divergence, the selection coefficients on each of 1,250 genes would be far too small to generate either a detectable QTL or an  $F_{st}$  outlier.

### The Two Stages of Ecological Speciation with Gene Flow

Consideration of the mosaic nature of the genome during early ecological speciation with gene flow suggests that genetic change in this form of speciation occurs in 2 distinct stages:

**Stage 1: Response to Divergent Selection (Fig. 6 A and B).** During the first stage of ecological speciation with gene flow, genomic regions containing major QTL for key traits quickly diverge under selection and become resistant to gene exchange. This establishes the com-



## Why Hasn't Divergence Hitchhiking Been Seen Before?

One hazard of the retrospective spyglass approach to speciation is that patterns of genetic divergence early in the process of speciation can become obscured or even invisible over time as additional divergence between the new species accumulates. This is likely to be the fate of divergence hitchhiking during the process of ecological speciation.

Early in ecological speciation with gene flow, populations diverge under selection at genomic regions that affect key ecologically important traits, while gene flow continues across the rest of the genome (Fig. 3).  $F_{st}$  outliers found during this period (stage 1) will tend to map to these selected regions, as suggested by our analyses (3). However, by the time most of the genome is phylogenetically concordant and retrospective analyses begin, the genomic signature of the original divergent selection will have faded. Outliers might be found, but they should not be expected to mark the genomic regions under early divergent selection.

During stage 2 of ecological speciation with gene flow, genetic divergence occurs mostly in genomic areas that were not originally affected by divergent selection. As overall genomic divergence between the new species increases through drift or independent responses to selection, the distinctive genetic signature of divergence hitchhiking (excessive divergence in regions near divergently selected QTL) becomes assimilated into a more widespread pattern of genetic divergence between the new species. So, it is perhaps not surprising that decades of retrospective analyses have not seen these important, but transient, regions of interracial linkage disequilibrium around divergent QTL.

$F_{st}$  outliers may still be found late in stage 2 or in hybrid zones between "good species" (e.g., ref. 68), but interpreting their cause becomes increasingly difficult as the overall level of genetic divergence between the new species grows. A high  $F_{st}$  marker seen late in speciation or in a hybrid zone is more likely to be the result of genetic drift or a recent selective sweep within 1 species than it is to be a signature of the divergent selection that caused speciation. Therefore, outliers found between new species or in hybrid zones, or between incipient species in secondary contact after a period of allopatric divergence, should not necessarily be expected to mark genomic regions affected by divergent phenotypic selection during the initial phases of ecological speciation.

Outlier analyses of races or morphs that have become partially reproductively isolated under divergent selection thus offer a privileged, but transient, view of the genetic mechanisms involved in early ecological speciation. It is in regions of divergence hitchhiking that ecological speciation with gene flow begins, and the divergence of the ecologically important QTL at their center determines the eventual pattern of branching seen later in the species tree. However, by the time that good species are recognized, the distinctive pattern of divergence hitchhiking around key QTL is gone, and the opportunity to analyze the genomic regions pivotal to ecological speciation with gene flow has been lost.

## The Real Difference Between Sympatric and Allopatric Speciation

In allopatric populations, where there is no possibility for gene exchange, virtually any type or strength of selection will eventually lead to reproductive isolation, and barriers to gene flow may be of virtually any kind. In their classic survey of reproductive isolation between *Drosophila* species, Coyne and Orr (66, 67) found that in allopatry, prezygotic (ecologically based) and postzygotic reproductive isolation (from DMIs) appeared to evolve at about the same rate.

In contrast, for speciation to occur without physical barriers to gene flow, divergent selection must be strong and "multifarious," i.e., affecting several different traits, which causes ecologically based isolation to evolve relatively rapidly (4, 17, 22, 23). Consistent with this, Coyne and Orr's comparative analyses (66, 67) suggest that in

sympatric populations, prezygotic isolation precedes the evolution of postzygotic isolation. The primacy of ecologically based isolation in speciation with gene flow is supported by empirical analyses of taxa in which divergent selection is thought to have been involved in speciation. They reveal extensive prezygotic ecologically based isolation, with little or no isolation attributable to postzygotic genetic incompatibilities (4, 17).

In the 2-stage model of speciation described here, allopatric speciation can occur without stage 1, but sympatric speciation cannot. There is essentially only 1 path for purely sympatric speciation: rapid divergence at genomic regions harboring QTL for traits under divergent selection, leading to significant ecologically based reduction of successful interbreeding between incipient species and ecological allopatry by the end of stage 1. Then, during stage 2, genetic incompatibilities can accumulate to reinforce the ecologically based isolation and make it permanent.

In contrast, allopatric speciation cannot be divided into distinct stages, because the accumulation of DMIs by independent responses to uniform or balancing selection can occur at the same time as the evolution of ecologically based isolation driven by divergent selection. In allopatry, any combination of divergent selection, uniform selection and genetic drift could produce speciation (Fig. 6C). Because the rapid divergence under selection that characterizes ecological speciation with gene flow is not required when populations are geographically isolated (although it can happen), allopatric speciation will often take much longer than speciation with gene flow.

## Divergence Hitchhiking Makes Sympatric Speciation Much More Likely Than Commonly Believed

Divergence hitchhiking neutralizes the most longstanding criticism of sympatric speciation, the difficulty of maintaining linkage disequilibrium between genes involved in resource use and those that produce assortative mating (e.g., ref. 2, pp. 127–137, and refs. 17, 24, and 25). Although it has been clear for some time that this problem is mitigated if the traits under divergent selection for resource use also affect mate choice (4), or if there is pleiotropy or physical linkage between the 2 classes of genes (17, 24, 25), these observations have done little to quell the controversy.

By providing a simple mechanism by which combinations of genes that produce assortative mating can accumulate and be protected from recombination, divergence hitchhiking removes the major constraint on sympatric speciation that prevented its acceptance for so long. The controversy over sympatric speciation has occupied a tremendous number of researchers over the past 50 years. If additional studies in other taxa show that divergence hitchhiking is a general phenomenon, we may finally be able to put this issue behind us.

## Conclusions

The genetic changes that produce speciation have fascinated researchers for many years. To date, most research on this crucial aspect of evolution has taken a retrospective approach that I call the spyglass. However, population-level analysis of the ecological and genetic mechanisms that produce reproductive isolation between partially isolated ecotypes or races (the magnifying glass) can provide a very different perspective on the problem of speciation. Both the spyglass and the magnifying glass are useful tools in the genetic analysis of speciation; any truly general theory of how speciation occurs must be consistent not only with observations from fully differentiated species, but also with mechanisms seen at the population level in partially isolated ecological races. Speciation is a multidimensional problem, and we will not solve Darwin's mystery unless we scrutinize it from every possible vantage point.

**ACKNOWLEDGMENTS.** I thank John Avise and Francisco Ayala for the invitation to discuss these ideas at the Sackler Colloquium; Dolph Schluter and an anonymous referee for exceptionally thoughtful comments on the manuscript; Gina

Conte for developing the multiplexed microsatellites for the new linkage map; Justin Malin for some clever computational assistance; and Casey Mason-Foley, Kelly Mills, and Jeffrey Lew for PCRs without end. My work on speciation is

supported by National Science Foundation Grants DEB9796222, DEB0221221, and DEB0528288 and the U.S. Department of Agriculture National Research Initiative, Gateways to Genomics Program.

1. Darwin C (1859) *On the Origin of Species by Means of Natural Selection or the Preservation of Favored Races in the Struggle for Life* (J. Murray, London).
2. Coyne JA, Orr HA (2004) *Speciation* (Sinauer, Sunderland, MA).
3. Via S, West JA (2008) The genetic mosaic suggests a new role for hitchhiking in ecological speciation. *Mol Ecol* 17:4334–4345.
4. Schluter D (2001) Ecology and the origin of species. *Trends Ecol Evol* 16:372–380.
5. Schluter D (2009) Evidence for ecological speciation and its alternative. *Science* 323:737–741.
6. Mayr E (1942) *Systematics and the Origin of Species* (Columbia Univ Press, New York).
7. Dobzhansky T (1937) *Genetics and the Origin of Species* (Columbia Univ Press, New York).
8. Dobzhansky T (1934) Studies on hybrid sterility. I. Spermatogenesis in pure and hybrid *Drosophila pseudoobscura*. *Z Zellforsch Microsk Anat* 21:169–221.
9. Muller HJ (1942) Isolating mechanisms, evolution, and temperature. *Biol Symp* 6:71–125.
10. Otte D, Endler JA (1989) *Speciation and Its Consequences* (Sinauer, Sunderland, MA).
11. Howard DJ, Berlocher SH (2003) *Endless Forms* (Sinauer, Sunderland, MA).
12. Ting C-T, Tsaur S-C, Wu C-I (2000) The phylogeny of closely related species as revealed by the genealogy of a speciation gene, *Odysseus*. *Proc Natl Acad Sci USA* 97:5313–5316.
13. Masly JP, Presgraves DC (2007) High-resolution genome-wide dissection of the two rules of speciation in *Drosophila*. *PLoS Biol* 5:1890–1898.
14. Phadnis N, Orr HA (2009) A single gene causes both male sterility and segregation distortion in *Drosophila* hybrids. *Science* 323:376–379.
15. Mihola O, Trachtulec Z, Vleck C, Schimenti JC, Forejt J (2009) A mouse speciation gene encodes a meiotic histone H3 methyltransferase. *Science* 323:373–375.
16. Willis J (2009) Origin of species in overdrive. *Science* 323:350–351.
17. Via S (2001) Sympatric speciation in animals: The ugly duckling grows up. *Trends Ecol Evol* 16:381–390.
18. Schemske D (2000) Understanding the origin of species. *Evolution (Lawrence, Kans)* 54:1069–1073.
19. Futuyma DJ (1987) On the role of anagenesis in speciation. *Am Nat* 130:465–473.
20. Haysm RG (1998) Linking evolutionary pattern and process: The relevance of species concepts for the study of speciation. *Endless Forms*, eds Howard DJ, Berlocher SH (Sinauer, Sunderland, MA), pp 19–31.
21. Ramsey J, Bradshaw HD, Schemske DW (2003) Components of reproductive isolation between the monkeyflowers *Mimulus lewisii* and *M. cardinalis* (Scrophulariaceae). *Evolution (Lawrence, Kans)* 57:1520–1534.
22. Rice WR, Hostert EE (1993) Laboratory experiments on speciation: What have we learned in 40 years? *Evolution (Lawrence, Kans)* 47:1637–1653.
23. Hendry AP, Nosil P, Rieseberg LJ (2007) The speed of ecological speciation. *Funct Ecol* 21:455–464.
24. Felsenstein J (1981) Skepticism toward Santa Rosalia, or why are there so few kinds of animals? *Evolution (Lawrence, Kans)* 35:124–138.
25. Hawthorne DJ, Via S (2001) Genetic linkage facilitates ecological specialization and reproductive isolation in pea aphids. *Nature* 412:904–907.
26. Hartl DL, Clark AG (1997) *Principles of Population Genetics* (Sinauer, Sunderland, MA), 3rd Ed.
27. Kondrashov AS, Yampolsky LY, Shabalina SA (1998) On the sympatric origin of species by means of natural selection. *Endless Forms*, eds Howard DJ, Berlocher SH (Sinauer, Sunderland, MA), pp 90–98.
28. Lewontin RC, Krakauer J (1973) Distribution of gene frequency as a test of the theory of selective neutrality of polymorphisms. *Genetics* 74:175–195.
29. Beaumont MA, Nichols RA (1996) Evaluating loci for use in the genetic analysis of population structure. *Proc R Soc London Ser B* 263:1619–1626.
30. Beaumont MA, Balding DJ (2004) Identifying adaptive genetic divergence among populations from genome scans. *Mol Ecol* 13:969–980.
31. Luikart G, England PR, Tallmon D, Jordan S, Taberlet P (2003) The power and promise of population genomics: From genotyping to genome typing. *Nat Rev Genet* 4:981–994.
32. Beaumont MA (2005) Adaptation and speciation: What can  $F_{ST}$  tell us? *Trends Ecol Evol* 20:435–440.
33. Storz JF (2005) Using genome scans of DNA polymorphism to infer adaptive population divergence. *Mol Ecol* 14:671–688.
34. Wilding CS, Butlin RK, Grahame J (2001) Differential gene exchange between parapatric morphs of *Littorina saxatilis* detected using AFLP markers. *J Evol Biol* 14:611–619.
35. Emelianov I, Marec F, Mallet J (2004) Genomic evidence for divergence with gene flow in host races of the larch budmoth. *Proc R Soc London Ser B* 271:97–105.
36. Rogers SM, Bernatchez L (2005) Integrating QTL mapping and genome scans toward the characterization of candidate loci under parallel selection in the lake whitefish. *Mol Ecol* 14:351–361.
37. Bonin A, Taberlet P, Miaud C, Pompanon F (2006) Explorative genome scan to detect candidate loci for adaptation along a gradient of altitude in the common frog (*Rana temporaria*). *Mol Biol Evol* 23:773–783.
38. Oetjen K, Reusch TB (2007) Genome scans detect consistent divergent selection among subtidal vs. intertidal populations of the marine angiosperm *Zostera marina*. *Mol Ecol* 16:5156–5167.
39. Eastop VF (1971) Keys for the identification of *Acyrtosiphon* (Hemiptera: Aphididae). *Bull Br Mus Nat Hist Entomol* 26:1–115.
40. Via S (1991) The genetic structure of host plant adaptation in a spatial patchwork: Demographic variability among reciprocally transplanted pea aphid clones. *Evolution (Lawrence, Kans)* 45:827–852.
41. Sandstrom J (1996) Temporal change in host adaptation in the pea aphid, *Acyrtosiphon pisum*. *Ecol Entomol* 21:56–62.
42. Simon JC, et al. (2003) Host-based divergence in populations of the pea aphid: Insights from nuclear markers and the prevalence of facultative symbionts. *Proc R Soc London Ser B* 270:1703–1712.
43. Ferrari J, Godfray CHJ, Faulconbridge AS, Prior K, Via S (2007) Population differentiation and genetic variation in host choice among pea aphids from eight host plant genera. *Evolution (Lawrence, Kans)* 60:1574–1584.
44. Ferrari J, Godfray CHJ, Via S (2008) Population differentiation and genetic variation in performance of pea aphids on eight host plant genera. *Evolution (Lawrence, Kans)* 62:2508–2523.
45. Peccoud J, et al. (2008) Host range expansion of an introduced insect pest through multiple colonizations of specialized clones. *Mol Ecol* 17:4608–4618.
46. Via S (1989) Field estimation of variation in host plant use between local populations of pea aphids from two crops. *Ecol Entomol* 14:357–364.
47. Via S (1991) Specialized host plant performance of pea aphid clones is not altered by experience. *Ecology* 72:1420–1427.
48. Via S (1999) Reproductive isolation between sympatric races of pea aphids. I. Gene flow restriction and habitat choice. *Evolution (Lawrence, Kans)* 53:1446–1457.
49. Via S, Bouck AC, Skillman S (2000) Reproductive isolation between sympatric races of pea aphids. II. Selection against migrants and hybrids in the parental environment. *Evolution (Lawrence, Kans)* 54:1626–1637.
50. Rogers SM, Bernatchez L (2007) The genetic architecture of ecological speciation and the association with signatures of selection in natural lake whitefish (*Coregonus* sp., *Salmonidae*) species pairs. *Mol Biol Evol* 24:1423–1438.
51. Begun DJ, Aquadro CF (1992) Levels of naturally occurring DNA polymorphism correlate with recombination rates in *D. melanogaster*. *Nature* 356:519–520.
52. Charlesworth B, Nordborg M, Charlesworth D (1997) The effects of local selection, balanced polymorphism and background selection on equilibrium patterns of genetic diversity in subdivided populations. *Genet Res* 70:155–174.
53. Noor MAF, Grams KL, Bertucci LA, Reiland J (2001) Chromosomal inversions and the reproductive isolation of species. *Proc Natl Acad Sci USA* 98:12084–12088.
54. Rieseberg LH (2001) Chromosomal rearrangements and speciation. *Trends Ecol Evol* 16:351–358.
55. Machado CA, Haselkorn TS, Noor MAF (2007) Evaluation of the genomic content of effects of fixed inversion differences on intraspecific variation and interspecific gene flow in *Drosophila pseudoobscura* and *D. persimilis*. *Genetics* 175:1289–1306.
56. Wood HM, Grahame JW, Humphray S, Rogers J, Butlin RK (2008) Sequence differentiation in regions identified by a genome scan for local adaptation. *Mol Ecol* 17:3123–3135.
57. Smadja C, Galindo J, Butlin R (2008) Hitching a lift on the road to speciation. *Mol Ecol* 17:1477–1480.
58. Avise JC (2000) *Phylogeography: The History and Formation of Species* (Harvard Univ Press, Cambridge, MA).
59. Maddison WP (1997) Gene trees in species trees. *Syst Biol* 46:523–536.
60. Beltran M, et al. (2002) Phylogenetic discordance at the species boundary: Comparative gene genealogies among rapidly radiating *Heliconius* butterflies. *Mol Biol Evol* 19:2176–2190.
61. Machado CA, Hey J (2003) The causes of phylogenetic conflict in a classic *Drosophila* species group. *Proc R Soc London Ser B* 270:1193–1202.
62. Mallarino R, Bermingham E, Willmott K, Whinnett A, Jiggins CD (2004) Molecular systematics of the butterfly genus *Ithomia* (Lepidoptera: Ithomiinae): A composite phylogenetic hypothesis based on seven genes. *Mol Phylog Evol* 34:625–644.
63. Pollard DA, Iyer VN, Moses AM, Eisen MB (2006) Widespread discordance of gene trees with species tree in *Drosophila*: Evidence for incomplete lineage sorting. *PLoS Genet* 2:1634–1647.
64. Dopman EB, Perez L, Bogdanowicz SM, Harrison RG (2005) Consequences of reproductive barriers for genealogical discordance in the European corn borer. *Proc Natl Acad Sci USA* 102:14706–14711.
65. Campbell D, Bernatchez L (2004) Genomic scan using AFLP markers as a means to assess the role of directional selection in the divergence of sympatric whitefish ecotypes. *Mol Biol Evol* 21:945–956.
66. Coyne JA, Orr HA (1989) Patterns of speciation in *Drosophila*. *Evolution (Lawrence, Kans)* 43:362–381.
67. Coyne JA, Orr HA (1997) Patterns of speciation in *Drosophila* revisited. *Evolution (Lawrence, Kans)* 51:295–303.
68. Yatabe Y, Kane NC, Scotti-Saintagne C, Rieseberg LH (2007) Rampant gene exchange across a strong reproductive barrier between the annual sunflowers, *Helianthus annuus* and *H. petiolaris*. *Genetics* 175:1883–1893.