Effectiveness of convalescent plasma therapy in severe COVID-19 patients

Kai Duan,1,2,3,4,5 Bende Liu,1,6 Cesheng Li,1,7 Huajun Zhang,1,2,8 Ting Yu,1,9 Jieming Qu,1,2,3,4 Min Zhou,1,2,3,4 Li Chen,1,8 Shengli Meng,1,7,9 Yong Hu,1,7,9 Cheng Peng,1,7,9 Mingchao Yuan,1,8 Jinyan Huang,1,2,8 Zejun Wang,1,2,8 Jianhong Yu,1,2,8 Xiaoxiao Gao,1,7 Dan Wang,1,7,9 Xiaoqi Yu,1,7,9,10 Li Li,1,2,11 Jiayou Zhang,1,2,12 Xiao Wu,1,2,10,11,12,13 Bei Li,1,2,10,11,12,13 Yanping Xu,1,2,3,9,10,11,13,14 Wei Chen,1,15 Yan Peng,1,7,9 Yeqin Hu,1,2,16 Linzhen Lin,1,7,9 Xuefei Li,1,8,9,10,11,16 Shihe Huang,1,2,8 Zhijun Zhou,1,2,8 Lianzhen Lin,1,7,9 Huichuan Yang,1,2,8 Xuefei Li,1,8 and Zhiwu Xia3

1China National Biotec Group Company Limited, 100029 Beijing, China; 2National Engineering Technology Research Center for Combined Vaccines, Wuhan Institute of Biological Products Co. Ltd., 430207 Wuhan, China; 3First People’s Hospital of Jiangxia District, 430200 Wuhan, China; 4Sinopharm Wuhan Plasma-derived Biotherapies Co., Ltd, 430207 Wuhan, China; 5Key Laboratory of Special Pathogens, Wuhan Institute of Virology, Center for Biosafety Mega-Science, Chinese Academy of Sciences, 430071 Wuhan, China; 6Department of Respiratory and Critical Care Medicine, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, 200025 Shanghai, China; 7National Research Center for Translational Medicine, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, 200025 Shanghai, China; 8Institute of Respiratory Diseases, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, 200025 Shanghai, China; 9Clinical Research Center, Department of Gastroenterology, Ruijin Hospital North, Shanghai Jiao Tong University School of Medicine, 200018 Shanghai, China; 10Wuhan Blood Center, 430030 Wuhan, China; 11State Key Laboratory of Medical Genomics, Shanghai Institute of Hematology, National Research Center for Translational Medicine, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, 200025 Shanghai, China; 12Research Laboratory of Critical Care Medicine, Ruijin Hospital and Ruijin Hospital North, National Research Center for Translational Medicine, Shanghai Jiao Tong University School of Medicine, 200025 Shanghai, China; and 13National Institute for Food and Drug Control of China, 102629 Beijing, China

Contributed by Zhu Chen, March 18, 2020 (sent for review March 5, 2020; reviewed by W. Ian Lipkin and Fusheng Wang)

Currently, there are no approved specific antiviral agents for novel coronavirus disease 2019 (COVID-19). In this study, 10 severe patients confirmed by real-time viral RNA test were enrolled prospectively. One dose of 200 mL of convalescent plasma (CP) derived from recently recovered donors with the neutralizing antibody titers above 1:640 was transfused to the patients as an addition to maximal supportive care and antiviral agents. The primary endpoint was the safety of CP transfusion. The secondary endpoints were the improvement of clinical symptoms and laboratory parameters within 3 d after CP transfusion. The median time from onset of illness to CP transfusion was 16.5 d. After CP transfusion, the level of neutralizing antibody increased rapidly up to 1:640 in five cases, while that of the other four cases maintained at a high level (1:640). The clinical symptoms were significantly improved along with increase of oxyhemoglobin saturation within 3 d. Several parameters tended to improve as compared to pretransfusion, including increased lymphocyte counts (0.65 × 10⁹/L vs. 0.76 × 10⁹/L) and decreased C-reactive protein (55.98 mg/L vs. 18.13 mg/L). Radiological examinations showed varying degrees of absorption of lung lesions within 7 d. The viral load was undetectable after CP transfusion in seven patients who had previous viremia. No severe adverse effects were observed. This study showed CP therapy was well tolerated and could potentially improve the clinical outcomes through neutralizing viremia in severe COVID-19 cases. The optimal dose and time point, as well as the clinical benefit of CP therapy, needs further investigation in larger well-controlled trials.


Reviewers: W.I.L., Columbia University; and F.W., Beijing 302 Hospital.
The authors declare no competing interest.

This open access article is distributed under Creative Commons Attribution License 4.0 (CC BY).

Data deposition: Detailed data on patients that support the findings of this study have been deposited in the Open Science Framework (https://osf.io/gahz5).

1K. Duan, B. Liu, C.L., H.Z., T.Y., J.Q., M.Z., and L.C. contributed equally to this work.
2To whom correspondence may be addressed. Email: zchen@stn.sh.cn, zhangx@shsmu.edu.cn, or yangxiaoming@sinopharm.com.

This article contains supporting information online at https://www.pnas.org/lookup/suppl/doi:10.1073/pnas.2004168117/-/DCSupplemental.

First published April 6, 2020.

© 2020, Licenced to National Academy of Sciences. All rights reserved.

Significance

COVID-19 is currently a big threat to global health. However, no specific antiviral agents are available for its treatment. In this work, we explore the feasibility of convalescent plasma (CP) transfusion to rescue severe patients. The results from 10 severe adult cases showed that one dose (200 mL) of CP was well tolerated and could significantly increase or maintain the neutralizing antibodies at a high level, leading to disappearance of viremia in 7 d. Meanwhile, clinical symptoms and paraclinical criteria rapidly improved within 3 d. Radiological examination showed varying degrees of absorption of lung lesions within 7 d. These results indicate that CP can serve as a promising rescue option for severe COVID-19, while the randomized trial is warranted.
for an alternative strategy for COVID-19 treatment, especially among severe patients.

Convalescent plasma (CP) therapy, a classic adaptive immunotherapy, has been applied to the prevention and treatment of many infectious diseases for more than one century. Over the past two decades, CP therapy was successfully used in the treatment of SARS, MERS, and 2009 H1N1 pandemic with satisfactory efficacy and safety (9–12). A meta-analysis from 32 studies of SARS coronavirus infection and severe influenza showed a statistically significant reduction in the pooled odds of mortality following CP therapy, compared with placebo or no therapy (odds ratio, 0.25; 95% confidence interval, 0.14–0.45) (13). However, the CP therapy was unable to significantly improve the survival in the Ebola virus disease, probably due to the absence of data of neutralizing antibody titration for stratified analysis (14). Since the virological and clinical characteristics share similarity among SARS, Middle East Respiratory Syndrome (MERS), and COVID-19 (15), CP therapy might be a promising treatment option for COVID-19 rescue (16). Patients who have recovered from COVID-19 with a high neutralizing antibody titer may be a valuable donor source of CP. Nevertheless, the potential clinical benefit and risk of convalescent blood products in COVID-19 remains uncertain. Hence, we performed this pilot study in three participating hospitals to explore the feasibility of CP treatment in 10 severe COVID-19 patients.

Results

Neutralizing Activity of CP against SARS-CoV-2. The neutralizing activity against SARS-CoV-2 was evaluated by classical plaque reduction test using a recently isolated viral strain (1). Among the first batch of CP samples from 40 recovered COVID-19 patients, 39 showed high antibody titers of at least 1:160, whereas only one had an antibody titer of 1:32. This result laid the basis for our pilot clinical trial using CP in severe patients.

General Characteristics of Patients in the Trial. From January 23, 2020, to February 19, 2020, 10 severe COVID-19 patients (six males and four females) were enrolled and received CP transfusion. The median age was 52.5 y (interquartile range [IQR], 45.0 y to 59.5 y) (Table 1). None of the patients had direct exposure to Huanan Seafood Wholesale Market. The median time from onset of symptoms to hospital admission and CP transfusion was 6 d (IQR, 2.5 d to 8.5 d) and 16.5 d (IQR, 11.0 d to 19.3 d), respectively. Three patients were affected by clustering infection. The most common symptoms at disease onset were fever (7 of 10 patients), cough (eight cases), and shortness of breath (eight cases), while less common symptoms included sputum production (five cases), chest pain (two cases), diarrhea (two cases), nausea and vomiting (two cases), headache (one case), and sore throat (one case). Four patients had underlying chronic diseases, including cardiovascular and/or cerebrovascular diseases and essential hypertension. Nine patients received arbidol monotherapy or combination therapy with remdesivir (in one case not included in the current clinical trial), or ribavirin, or peramivir, while one patient received ribavirin monotherapy (Table 2). Antibacterial or antifungal treatment was used when patients had coinfection. Six patients received intravenous (i.v.) methylprednisolone (20 mg every 24 h).

On computer-assisted tomography (CT), all patients presented bilateral ground-glass opacity and/or pulmonary parenchymal consolidation with predominantly subpleural and bronchovascular bundles distribution in the lungs. Seven patients had multiple lobe involvement, and four patients had interlobular septal thickening.

Effects of CP Transfusion.

Improvement of clinical symptoms. All symptoms in the 10 patients, especially fever, cough, shortness of breath, and chest pain, disappeared or largely improved within 1 d to 3 d upon CP transfusion. Prior to CP treatment, three patients received mechanical ventilation, three received high-flow nasal cannula oxygenation, and two received conventional low-flow nasal cannula oxygenation. After treatment with CP, two patients were weaned from mechanical ventilation to high-flow nasal cannula, and one patient discontinued high-flow nasal cannula. Besides, in one patient treated with conventional nasal cannula oxygenation, continuous oxygenation was shifted to intermittent oxygenation (Table 2).

Reduction of pulmonary lesions on chest CT examinations. According to chest CTs, all patients showed different degrees of absorption of pulmonary lesions after CP transfusion. Representative chest CT

Table 1. Clinical characteristics of patients receiving CP transfusion

<table>
<thead>
<tr>
<th>Patient no.</th>
<th>Sex</th>
<th>Age, y</th>
<th>Clinical classification</th>
<th>Days of admission from symptom onset</th>
<th>Days of CP therapy from symptom onset</th>
<th>Clustering infection</th>
<th>Principal symptoms</th>
<th>Comorbidity</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>M</td>
<td>46</td>
<td>Severe</td>
<td>8</td>
<td>11</td>
<td>No</td>
<td>Fever, cough, sputum production, shortness of breath, chest pain</td>
<td>Hypertension</td>
</tr>
<tr>
<td>2</td>
<td>F</td>
<td>34</td>
<td>Severe</td>
<td>0</td>
<td>11</td>
<td>Yes</td>
<td>Cough, shortness of breath, chest pain</td>
<td>None</td>
</tr>
<tr>
<td>3</td>
<td>M</td>
<td>42</td>
<td>Severe</td>
<td>8</td>
<td>19</td>
<td>Yes</td>
<td>Fever, cough, sputum production, shortness of breath, sore throat, diarrhea</td>
<td>Hypertension</td>
</tr>
<tr>
<td>4</td>
<td>F</td>
<td>55</td>
<td>Severe</td>
<td>10</td>
<td>19</td>
<td>No</td>
<td>Fever, cough, sputum production, shortness of breath</td>
<td>None</td>
</tr>
<tr>
<td>5</td>
<td>M</td>
<td>57</td>
<td>Severe</td>
<td>4</td>
<td>14</td>
<td>No</td>
<td>Fever, shortness of breath</td>
<td>None</td>
</tr>
<tr>
<td>6</td>
<td>F</td>
<td>78</td>
<td>Severe</td>
<td>8</td>
<td>17</td>
<td>Yes</td>
<td>Fever, cough, sputum production, shortness of breath, muscle ache</td>
<td>None</td>
</tr>
<tr>
<td>7</td>
<td>M</td>
<td>56</td>
<td>Severe</td>
<td>4</td>
<td>16</td>
<td>No</td>
<td>Fever, cough, sputum production, arthralgia</td>
<td>Cardiovascular and cerebrovascular diseases</td>
</tr>
<tr>
<td>8</td>
<td>M</td>
<td>67</td>
<td>Severe</td>
<td>10</td>
<td>20</td>
<td>No</td>
<td>Fever, cough, headache, diarrhea, vomiting</td>
<td>None</td>
</tr>
<tr>
<td>9</td>
<td>F</td>
<td>49</td>
<td>Severe</td>
<td>1</td>
<td>10</td>
<td>No</td>
<td>Cough, shortness of breath</td>
<td>None</td>
</tr>
<tr>
<td>10</td>
<td>M</td>
<td>50</td>
<td>Severe</td>
<td>3</td>
<td>20</td>
<td>No</td>
<td>Shortness of breath</td>
<td>Hypertension</td>
</tr>
</tbody>
</table>

M, male; F, female.
images of patient 9 and patient 10 are shown on Fig. 1. Patient 9, a 49-y-old female admitted 1 day postonset of illness (dpoi), showed the most obvious pulmonary image improvement. At 10 dpoi, one dose of 200-mL transfusion of CP was given. The SARS-CoV-2 RNA converted to negative at 12 dpoi. Compared with the result at 7 dpoi, massive infiltration and ground-glass attenuation disappeared on CT image performed at 13 dpoi, accompanied by a much better pulmonary function. Patient 10, a 50-y-old male, was admitted 3 dpoi and was given a 200-mL transfusion of CP at 20 dpoi. His chest CT presented massive infiltration and ground-glass attenuation on admission and started to show a gradual absorption of lung lesions 5 d after CP transfusion. The SARS-CoV-2 RNA became negative at 25 dpoi.

**Table 2. Other treatments of ten patients receiving CP transfusion**

<table>
<thead>
<tr>
<th>Patient no.</th>
<th>Antiviral treatment</th>
<th>Antibiotic or antifungal treatment</th>
<th>Corticosteroids treatment</th>
<th>Oxygen support Before CP therapy</th>
<th>Oxygen support After CP therapy</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Arbidol 0.2 g q8h po.</td>
<td>Cefoperazone Sodium i.v.</td>
<td>None</td>
<td>High-flow nasal cannula, mechanical ventilation</td>
<td>Mechanical ventilation</td>
</tr>
<tr>
<td>2</td>
<td>Arbidol 0.2 g q8h po.</td>
<td>Cefoperazone Sodium i.v.</td>
<td>None</td>
<td>High-flow nasal cannula, mechanical ventilation</td>
<td>Mechanical ventilation</td>
</tr>
<tr>
<td>3</td>
<td>Arbidol 0.2 g q8h po.</td>
<td>Moxifloxacin i.v.</td>
<td>Methylprednisolone i.v</td>
<td>High-flow nasal cannula, mechanical ventilation</td>
<td>Mechanical ventilation</td>
</tr>
<tr>
<td>4</td>
<td>Ribavirin 0.5 g qd i.v.</td>
<td>Linezolid i.v.</td>
<td>Methylprednisolone i.v</td>
<td>High-flow nasal cannula</td>
<td>High-flow nasal cannula</td>
</tr>
<tr>
<td>5</td>
<td>Ribavirin 0.2 g q8h po.</td>
<td>Moxifloxacin i.v.</td>
<td>Methylprednisolone i.v</td>
<td>Low-flow nasal cannula</td>
<td>Low-flow nasal cannula</td>
</tr>
<tr>
<td>6</td>
<td>Ribavirin 0.2 g q8h po.</td>
<td>Cefoperazone Sodium i.v.</td>
<td>Methylprednisolone i.v</td>
<td>High-flow nasal cannula</td>
<td>None</td>
</tr>
<tr>
<td>7</td>
<td>Ribavirin 0.2 g q8h po.</td>
<td>Cefoperazone Sodium i.v.</td>
<td>Methylprednisolone i.v</td>
<td>High-flow nasal cannula</td>
<td>None</td>
</tr>
<tr>
<td>8</td>
<td>Arbidol 0.2 g q8h po.</td>
<td>None</td>
<td>None</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>9</td>
<td>Arbidol 0.2 g q8h po.</td>
<td>None</td>
<td>None</td>
<td>Low-flow nasal cannula</td>
<td>Low-flow nasal cannula (Intermittent)</td>
</tr>
<tr>
<td>10</td>
<td>Oseltamivir 75 mg q12h po.</td>
<td>None</td>
<td>None</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>10</td>
<td>Peramivir 0.3 g qd i.v.</td>
<td>None</td>
<td>None</td>
<td>None</td>
<td>None</td>
</tr>
</tbody>
</table>

Po., per os; i.v., i.v. injection; inh., inhalation; q8h, every 8 h; qd, per day; q12h, every 12 h; MIU, million IU.

Amelioration of routine laboratory criteria and pulmonary function. Lymphocytopenia, an important index for prognosis in COVID-19 (2), tended to be improved after CP transfusion (median: 0.65 × 10^9 per L vs. 0.76 × 10^9 per L), 7 out of 10 patients showing an increase of lymphocyte counts (Fig. 2). Concerning other laboratory tests, we observed a tendency of decrement of parameters indicative of inflammation and/or liver dysfunction as compared to the status before CP therapy. These included C-reactive protein (CRP) (median: 55.98 mg/L vs. 18.13 mg/L), alanine aminotransferase (median: 42.00 U/L vs. 34.30 U/L), and aspartate aminotransferase (median: 38.10 U/L vs. 30.30 U/L) (Table 3). The total bilirubin (median: 12.40 μmol/L vs. 13.98 μmol/L) remained unchanged, except for an obvious increment in patient 1 (Fig. 2). An increase of SaO2 (median: 93.00% vs. 96.00%), a measurement constantly performed in most patients in our trial, was found, which could indicate recovering lung function. This temporal relationship was notable despite the provision of maximal supportive care and antiviral agents.

Remarkably, patient 1, a 46-y-old male admitted 8 dpoi, had a very quick recovery, with much improved result of laboratory tests. He received antiviral drugs (arbidol and ribavirin) treatment and high-flow nasal cannula on admission. Mechanical ventilation was given at 10 dpoi for critical care support. CP transfusion was performed at 11 dpoi. At 12 dpoi, the SARS-CoV-2 test turned to negative, with a sharp decrease of CRP from 65.04 mg/L to 23.57 mg/L and increment of SaO2 from 86% to 90% (Fig. 3). The mechanical ventilation was successfully weaned off 2 d after CP transfusion. At 15 dpoi, a steady elevation of lymphocyte count and a drop of aminopherase level were observed, indicating improvement of immunological and hepatic function.

Increase of neutralizing antibody titers and disappearance of SARS-CoV-2 RNA. We determined neutralizing antibody titers before and after CP transfusion in all patients except one (patient 2) (Table 4). The neutralizing antibody titers of five patients increased and four patients remained at the same level after CP transfusion. SARS-CoV-2 RNA, assayed by RT-PCR, was positive in seven patients before CP transfusion. Of note is that SARS-CoV-2 RNA was decreased to an undetectable level in three patients on day 2, three patients on day 3, and one patient on day 6 after CP therapy. These results are in support of a neutralizing effect of CP on serum SARS-CoV-2.

Outcome of patients treated with CP as compared to a recent historic control group. A historic control group was formed by random selection of 10 patients from the cohort treated in the same hospitals and matched by age, gender, and severity of the diseases to the 10 cases in our trial. Baseline characteristics of patients between CP treatment group and control group showed no significant differences, while clinical outcomes of these two groups were different: three cases discharged while seven cases in much improved status and ready for discharge in CP group, as compared to three deaths, six cases in stabilized status, and one case in improvement in the control group (P < 0.001; SI Appendix, Table S1).
Adverse Effects of CP Transfusions. Patient 2 showed an evanescent facial red spot. No serious adverse reactions or safety events were recorded after CP transfusion.

Discussion

Our study explores the feasibility of CP therapy in COVID-19. All enrolled severe COVID-19 patients achieved primary and secondary outcomes. One dose of 200-mL CP transfusion was well tolerated, while the clinical symptoms significantly improved with the increase of oxyhemoglobin saturation within 3 d, accompanied by rapid neutralization of viremia.

Severe pneumonia caused by human coronavirus was characterized by rapid viral replication, massive inflammatory cell infiltration, and elevated proinflammatory cytokines or even cytokine storm in alveoli of lungs, resulting in acute pulmonary injury and acute respiratory distress syndrome (ARDS) (17). Recent studies on COVID-19 demonstrated that the lymphocyte counts in the peripheral blood were remarkably decreased and the levels of cytokines in the plasma from patients requiring intensive care unit (ICU) support, including IL-6, IL-10, TNF-α, and granulocyte-macrophage colony-stimulating factor, were
significantly higher than in those who did not require ICU conditions (2, 18). CP, obtained from recovered COVID-19 patients who had established humoral immunity against the virus, contains a large quantity of neutralizing antibodies capable of neutralizing SARS-CoV-2 and eradicating the pathogen from blood circulation and pulmonary tissues (19). In the present study, all investigated patients achieved serum SARS-CoV-2 RNA negativity after CP transfusion, accompanied by an increase of oxygen saturation and lymphocyte counts, and the improvement of liver function and CRP. The results suggest that the inflammation and overreaction of the immune system were alleviated by antibodies contained in CP. The case fatality rates (CFRs) in the present study were 0% (0/10), which was comparable to the CFRs in SARS, which varied from 0% (0/10) to 12.5% (10/80) in four noncomparative studies using CP treatment (9, 20–22). Based on our preliminary results, CP therapy can be an easily accessible, promising, and safe rescue option for severe COVID-19 patients. It is, nevertheless, worth mentioning that the absorption of pulmonary lesions often lagged behind the improvement of clinical symptoms, as shown in patients 9 and 10 in this trial.

The first key factor associated with CP therapy is the neutralizing antibody titer. A small sample study in MERS-CoV infection showed that the neutralizing antibody titer should exceed 1:80 to achieve effective CP therapy (12). To find eligible donors who have high levels of neutralizing antibody is a prerequisite. Cao et al. (23) showed that the level of specific neutralizing antibody to SARS-CoV decreased gradually 4 mo after the disease process, reaching undetectable levels in 25.6% (IgG) and 16.1% (neutralizing antibodies) of patients at 36 mo after disease status. A study from the MERS-CoV−infected patients and the exposed healthcare workers showed that the prevalence of MERS-CoV IgG seroreactivity was very low (2.7%), and the antibodies titer decreased rapidly within 3 mo (24). These studies suggested that the neutralizing antibodies represented short-lasting humoral immune response, and plasma from recently recovered patients should be more effective. In the present study, recently recovered COVID-19 patients, who were infected by SARS-CoV-2 with neutralizing antibody titer above 1:640 and recruited from local hospitals, should be considered as suitable donors. The median age of donors was lower than that of recipients (42.0 y vs. 52.5 y). Among the nine cases investigated, the neutralizing antibody titers of five patients increased to 1:640 within 2 d, while four patients kept the same level. The antibody titers in CP in COVID-19 seem thus higher than those used in the treatment of MERS patient (1:80) (12).

The second key factor associated with efficacy is the treatment time point. A better treatment outcome was observed among SARS patients who were given CP before 14 dpi (58.3% vs. 15.6%; P < 0.01), highlighting the importance of timely rescue therapy (9). The mean time from onset of illness to CP transfusion was 16.5 d. Consistent with previous research, all three patients receiving plasma transfusion given before 14 dpi (patients 1, 2,...
and 9) in our study showed a rapid increase of lymphocyte counts and a decrease of CRP, with remarkable absorption of lung lesions in CT. Notably, patients who received CP transfusion after 14 dpi showed much less significant improvement, such as patient 10. However, the dynamics of the viremia of SARS-CoV-2 was unclear, so the optimal transfusion time point needs to be determined in the future.

In the present study, no severe adverse effects were observed. One of the risks of plasma transfusion is the transmission of the potential pathogen. Methylene blue photochemistry was applied in this study to inactivate the potential residual virus and to maintain the activity of neutralizing antibodies as much as possible, a method known to be much better than ultraviolet (UV) C light (25). No specific virus was detected before transfusion. Transfusion-related acute lung injury was reported in an Ebola virus disease woman who received CP therapy (26). As a result, the possibility that these antiviral agents could contribute to the recovery of patients, or synergize with the therapeutic effect of CP, as well as the optimal concentration of neutralizing antibodies and treatment schedule, should be further clarified. Third, the dynamic changes of cytokines during treatment were not investigated. Nevertheless, the preliminary results of this trial seem promising, justifying a randomized controlled clinical trial in a larger patient cohort.

In conclusion, this pilot study on CP therapy shows a potential therapeutic effect and low risk in the treatment of severe COVID-19 patients. One dose of CP with a high concentration of neutralizing antibodies can rapidly reduce the viral load and tends to improve clinical outcomes. The optimal dose and treatment time point, as well as the definite clinical benefits of CP therapy, need to be further investigated in randomized clinical studies.

Materials and Methods

Patients. From January 23, 2020, to February 19, 2020, 10 patients in three participating hospitals (Wuhan Jinyintan Hospital; the Jiangxia District Hospital of Integrative Traditional Chinese and Western Medicine, Wuhan; and the First People's Hospital of Jiangxia District, Wuhan) were recruited in this pilot study. All patients were diagnosed as having severe COVID-19 according to the WHO Interim Guidance (30) and the Guideline of Diagnosis and Treatment of COVID-19 of National Health Commission of China (version 5.0) (31), with confirmation by real-time RT-PCR assay. The enrollment criteria were one of the conditions 2 to 4 plus condition 1: 1) age ≥ 18 y; 2) respiratory distress, RR ≥30 beats/min; 3) oxygen saturation level less than 93% in resting state; and 4) partial pressure of oxygen (PaO2)/oxygen concentration (FiO2) ≤ 300 mmHg (1 mmHg = 0.133 kPa). The exclusion criteria were as follows: 1) previous allergic history to plasma or ingredients (sodium citrate); 2) cases with serious general conditions, such as severe organ dysfunction, who were not suitable for CP transfusion. Written informed consent according to the Declaration of Helsinki was obtained from each patient or legal relatives. This study was approved by the Ethics Committee of the China National Biotec Group Co., Ltd. (Approval number 2020-0001). The registration number of this trial is ChiCTR2000030046.

Donors for Convalescent Plasma Transfusion. Ten donor patients who recovered from COVID-19 were recruited from three participating hospitals. The recovery criteria were as follows: 1) normoty of body temperature for more than 3 d, 2) resolution of respiratory tract symptoms, and 3) two consecutively negative results of sputum SARS-CoV-2 by RT-PCR assay (1-d sampling interval). The donor's blood was collected after 3 wk postonset.

Table 4. Comparison of serum neutralizing antibody titers and SARS-CoV-2 RNA load before and after CP therapy

<table>
<thead>
<tr>
<th>Patient no.</th>
<th>CP transfusion date</th>
<th>Serum neutralizing antibody titers</th>
<th>SARS-CoV-2 RNA load (Ct value)</th>
<th>Date</th>
<th>Serum neutralizing antibody titers</th>
<th>SARS-CoV-2 RNA load (Ct value)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>February 9</td>
<td>1:160</td>
<td>37.25</td>
<td>February 10</td>
<td>1:640</td>
<td>Negative</td>
</tr>
<tr>
<td>2</td>
<td>February 9</td>
<td>Unavailable</td>
<td>35.08</td>
<td>February 11</td>
<td>Unavailable</td>
<td>Negative</td>
</tr>
<tr>
<td>3</td>
<td>February 13</td>
<td>1:320</td>
<td>38.07</td>
<td>February 14</td>
<td>1:640</td>
<td>Negative</td>
</tr>
<tr>
<td>4</td>
<td>February 12</td>
<td>1:160</td>
<td>37.68</td>
<td>February 14</td>
<td>1:640</td>
<td>Negative</td>
</tr>
<tr>
<td>5</td>
<td>February 12</td>
<td>1:640</td>
<td>Negative</td>
<td>February 14</td>
<td>1:640</td>
<td>Negative</td>
</tr>
<tr>
<td>6</td>
<td>February 12</td>
<td>1:640</td>
<td>Negative</td>
<td>February 14</td>
<td>1:640</td>
<td>Negative</td>
</tr>
<tr>
<td>7</td>
<td>February 12</td>
<td>1:320</td>
<td>34.64</td>
<td>February 14</td>
<td>1:640</td>
<td>Negative</td>
</tr>
<tr>
<td>8</td>
<td>February 12</td>
<td>1:640</td>
<td>35.45</td>
<td>February 14</td>
<td>1:640</td>
<td>Negative</td>
</tr>
<tr>
<td>9</td>
<td>February 12</td>
<td>1:160</td>
<td>Negative</td>
<td>February 14</td>
<td>1:640</td>
<td>Negative</td>
</tr>
<tr>
<td>10</td>
<td>February 9</td>
<td>1:640</td>
<td>38.19</td>
<td>February 14</td>
<td>1:640</td>
<td>Negative</td>
</tr>
</tbody>
</table>
of illness and 4 d postdischarge. Written informed consent was obtained from each patient.

**Plasma Preparation Procedure and Quality Control.** Apheresis was performed using a Baxter CS 300 cell separator (Baxter). Convalescence plasma for treatment was collected from 40 donors. The median age was 42.0 y (IQR, 32.5 y to 49 y). A 200- to 400-mL ABO-compatible plasma sample was harvested from each donor depending on age and body weight, and each sample was divided and stored as 200-mL aliquots at 4 °C without any detergent or heat treatment. The CP was then treated with methylene blue and light treatment for 30 min in the medical plasma virus inactivation cabinet (Shandong Zhongbaokang Medical Appliance Co., Ltd).

**SeroLOGY Test and Real-Time RT-PCR Detection of SARS-CoV-2 and Other Pathogens.** The neutralizing activity of plasma was determined by plaque reduction neutralization test using SARS-CoV-2 virus in the high biosafe level (BSL-4) laboratory of Wuhan Institute of Virology, Chinese Academy of Sciences. Neutralization titer was defined as the highest serum dilution with 50% reduction in the number of plaques, as compared with the number of plaques in wells in the absence of novel coronavirus antibody as blank control. The neutralizing activity of the receiver-binding domain of antibody in the CP was detected by a sandwich enzyme-linked immunosorbent assay (ELISA). SARS-CoV-2 IgG antibody titer was tested by ELISA. SARS-CoV-2 RNA was detected by RT-PCR assay, and the result was reported as cycle threshold (Ct) value (Shanghai BioGerm Medical Biotechnology Co., Ltd). Methylene blue residue was detected by the verified UV method. The serology screening for hepatitis B and C virus, HIV, and syphilis spirochete was negative. The protocols for neutralization assay, serological test, and real-time RT-PCR detection of SARS-CoV RNA are presented in SI Appendix.

**Treatment.** All patients were admitted to the ICU and received antiviral therapy and other supportive care, while some patients received antibiotic treatment, antifungal treatment, glucocorticoid, and oxygen support at the appropriate situation. One dose of 200 mL of inactivated CP with neutralization activity of >1:640 was transfused into the patients within 4 h following the WHO blood transfusion protocol.


**Data Collection.** Clinical information of all enrolled patients was retrieved from the hospital electronic history system, including the baseline demographic data, days of illness duration, presenting symptoms, different kinds of examination, and methods of treatment. Bacterial coinfection was identified by a positive culture from respiratory, urinary, or blood culture within 48 h of hospital admission. Complications, including acute renal failure, acute coronary syndrome, myocarditis, ARDS, and nosocomial infection, were recorded. The applications of assisted mechanical ventilation, intranasal oxygen inhalation, and medication regimen were recorded. The SARS-CoV-2 RNA from the serum sample was monitored during treatment.

**Outcome Measures and Definitions.** The clinical symptoms were recorded by attending physicians daily. The blood test and biochemical tests were carried out every 1 d to 2 d. SARS-CoV-2 RNA was detected every 2 d to 3 d. CT scan was repeated every 3 d to 5 d. The primary endpoint was the safety of CP transfusion. The secondary endpoints were the improvement of clinical symptoms and laboratory and radiological parameters within 3 d after CP transfusion. Clinical symptoms improvement was defined as temperature normalization, relief of dyspnea, and oxygen saturation normalization, and radiological improvement was defined as different degrees of absorption of lung lesions.

**Statistical Analysis.** Continuous variables were presented as the median and IQR. Graphs were plotted using GraphPad Prism 7.0. Statistical software used included SPSS 24.0.

**Data Availability Statement.** All data relevant to this manuscript and available to the authors at the time of publication are included in the main text or SI Appendix. Further detailed data on patients that support the findings of this study have been deposited in the Open Science Framework (https://osf.io/qahz5/).

**Acknowledgments.** This study was funded by Key projects of the Ministry of Science and Technology China “Preparation of specific plasma and specific globulin from patients with a recovery period of COVID-19 infection” (Project 2020YFC0841800). This work was also supported by Shanghai Guangci Translational Medicine Development Foundation. We thank all patients and donors involved in this study.