

# Molecular vibration-sensing component in *Drosophila melanogaster* olfaction

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**A common explanation of molecular recognition by the olfactory system posits that receptors recognize the structure or shape of the odorant molecule. We performed a rigorous test of shape recognition by replacing hydrogen with deuterium in odorants and asking whether *Drosophila melanogaster* can distinguish these identically shaped isotopes. We report that flies not only differentiate between isotopic odorants, but can be conditioned to selectively avoid the common or the deuterated isotope. Furthermore, flies trained to discriminate against the normal or deuterated isotopes of a compound, selectively avoid the corresponding isotope of a different odorant. Finally, flies trained to avoid a deuterated compound exhibit selective aversion to an unrelated molecule with a vibrational mode in the energy range of the carbon–deuterium stretch. These findings are inconsistent with a shape-only model for smell, and instead support the existence of a molecular vibration-sensing component to olfactory reception.**

Olfactory systems perform remarkable feats of molecular recognition, but although much is known about the neurophysiology of olfaction (1–5), how olfactory receptors "read" molecular structure remains unknown. Parts of odorant molecules (odotopes) have been proposed to engage particular receptors in a "lock-and-key" manner and this molecular shape recognition mechanism is thought sufficient for odor discrimination (2). An alternative hypothesis (6) posits that molecular vibrations of all atoms, or of particular functional groups of odorant molecules, contribute to odor recognition, and odorants with similar vibrational spectra should elicit similar olfactory responses (7). Molecules in which deuterium replaces nonexchangeable hydrogens constitute appropriate probes to test these alternatives because deuteration does not alter atom size or bond length or stiffness (8). Thus, the conformation of a deuterated molecule should be identical to that of a hydrogen-only (i.e., normal) odorant and, according to the molecular recognition theory, the two isotopes should smell identical. However, atoms in a molecule vibrate in normal modes at particular energies that depend on the molecular structure. The doubling of nuclear mass upon deuteration will change all the vibrational modes of an odorant molecule to differing extents. The second hypothesis predicts that deuteration will alter its smell in comparison with the hydrogen-only isotope. A recent attempt to test the latter hypothesis in humans was rather inconclusive, potentially confounded by interference from previous experience, olfactory training or habituation of the subjects, or shortcomings of human olfaction as suggested by the authors (9). A major experimental difficulty is that, in general, isotopes are prepared and purified differently, and therefore perceived differences in smell could be attributed to different impurities. Purity can, in principle, be ensured by smelling single-molecule peaks exiting a gas chromatographic column, but this makes simultaneous two-compound comparisons difficult and animal experiments harder still.

Here we describe a different approach to test whether molecular vibrations contribute to recognition of odor character. We assumed that, if molecular vibrations are indeed detected, the deuterated isotopes of different odorants may share a common odor character, as deuteration shifts particular peaks—e.g., the carbon–deuterium (C–H) stretch—of the compound-specific vibrational spectra to lower frequencies. If so, the deuterium

odor character could be distinct and identifiable, irrespective of the structure and chemical properties of the odorant molecules that carry it. Significantly, we used *Drosophila* as unbiased and objective subjects to address this issue. They possess a relatively well understood olfactory system (10–13), exhibit keen olfactory discrimination (14–16), and can be conditioned to selectively avoid or seek odors with the use of established methodology (17, 18). We ask whether *Drosophila* can detect deuterium as a distinguishing molecular feature in odorant isotopes and a salient cue for conditioning. The results of these experiments provide support for the notion that flies can smell molecular vibrations.

## Results

**Spontaneous Differential Responses to Deuterated Odorants.** Although deuteration does not appreciably change molecular shape, atom size, or bond length or stiffness, it doubles hydrogen mass, thus affecting the overall vibrational modes of an odorant. Therefore, if recognition of molecular shape alone was the sole determinant for odor character (2, 3), then flies should not respond differentially to deuterated [ $d_x$ , where  $d_x$  denotes replacement of  $x$  nonexchangeable hydrogens with deuterium atoms] and nondeuterated/normal (i.e., H-) odorants. To address this hypothesis, we took advantage of the commercial availability of acetophenone (ACP) carrying three, five, or eight deuterium atoms ( $d_3$ ,  $d_5$ , and  $d_8$ ) in place of the respective hydrogens in the normal molecule (h-ACP). Equal amounts (75  $\mu$ L) of each odorant were diluted to 1 mL in isopropyl myristate and we quantified (Fig. 1A) the response of groups of flies to each odorant versus unscented air traversing the arms of a standard T-maze (*Materials and Methods*) (19, 20). When given a choice between normal ACP (i.e., h-ACP) and air, the excess flies present in the odor-delivering maze arm indicated that, at the dilution used, the odorant was attractive. However, replacing only three hydrogen atoms with deuterium eliminated this spontaneous preference as the flies distributed nearly equally in the arms of the maze. The  $d_5$ -ACP was mildly aversive and the fully substituted  $d_8$ -ACP was the most aversive of the four isotopes. Moreover, flies consistently avoided  $d_3$ ,  $d_5$ , and  $d_8$  ACP against equal concentration of h-ACP (Fig. S1). Therefore, a spontaneous switch in osmotaxis (14) to ACP was precipitated by substituting hydrogen atoms with deuterium.

To verify that flies are generally capable of spontaneous discrimination between airborne deuterated odorants and their normal counterparts, we additionally presented them with equal concentrations of normal and deuterated 1-octanol, normal and  $d_5$ -benzaldehyde, or a normal–perdeuterated ACP pair. In confirmation of the previous results, h-ACP was preferred versus an equal concentration of the  $d_8$  odorant (Fig. 1B). Reducing the concentration of the deuterated compound by 50% resulted in

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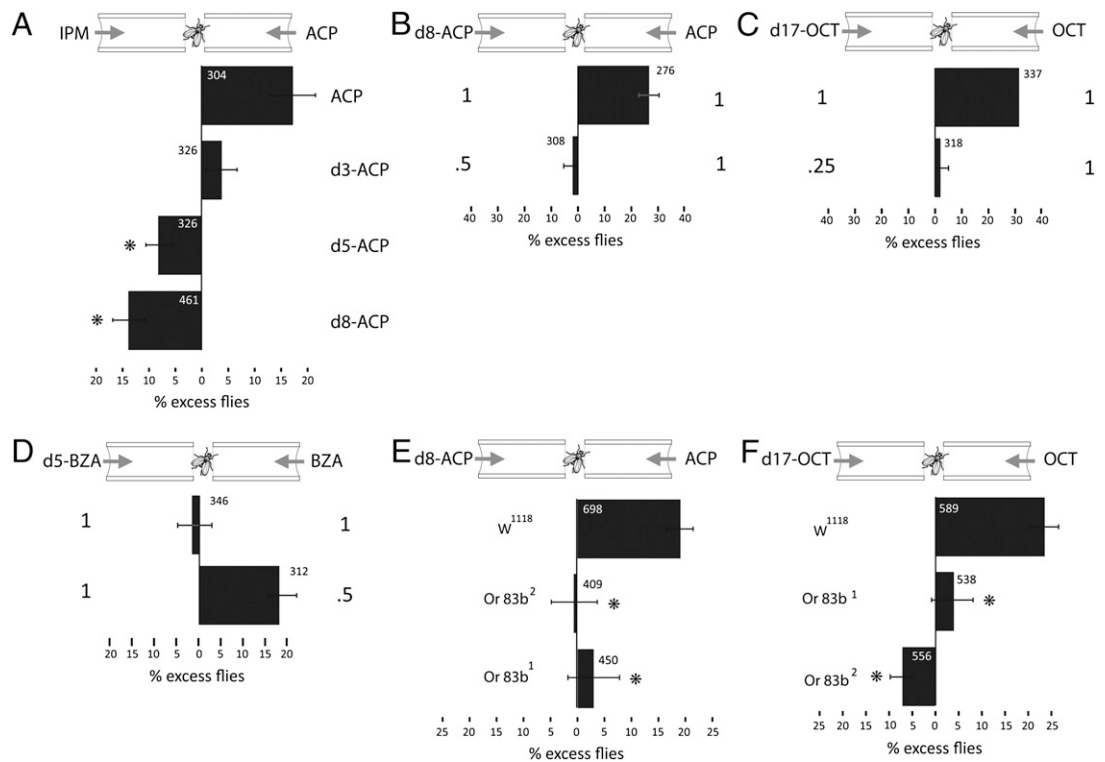
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**Fig. 1.** Differential spontaneous responses to odorants containing deuterium. The mean relative distribution of flies in the arms of the maze (% excess flies) carrying the indicated odorants  $\pm$  SEM is shown in all graphs. This metric reflects the prevailing distribution within the arms of the maze of groups of 40 to 60 flies tested each time. The total number of flies tested in each group is shown and the number of groups tested is  $n \geq 6$  for all groups. (A) Spontaneous responses to 75  $\mu$ L normal or d<sub>3</sub>, d<sub>5</sub>, or d<sub>8</sub>-ACP, each diluted with isopropyl myristate (IPM) to 1 mL. Flies spontaneously preferred h-ACP over the solvent. In contrast, incorporation of five or eight deuterium atoms results in significantly different distribution ( $P < 0.001$ ) from that toward h-ACP (attraction) to aversion. In contrast, the response to d<sub>3</sub>-ACP was not significantly different ( $P = 0.012$ ) from that of h-ACP. (B) Flies discriminate against d<sub>8</sub>-ACP if presented with equal amount (1:1) of h-ACP (75  $\mu$ L odorant/925  $\mu$ L IPM). However, a 50% reduction in the amount of deuterated odorant yielded an equal distribution of the flies in the arms (% excess flies not significantly different from zero), defined as a balanced maze. (C) Similarly equal amounts (200  $\mu$ L) of h-1-octanol (OCT) and d<sub>17</sub>-1-octanol yielded strong discrimination against the deuterated odorant, which was eliminated upon reducing it by 75% (1:0.25). (D) In contrast, equal amounts of normal and deuterated benzaldehyde (90  $\mu$ L) did not result in differential discrimination. In accord, decreasing the amount of h-BZA resulted in differential avoidance of d<sub>5</sub>-BZA. (E) The preferential discrimination against d<sub>8</sub>-ACP was eliminated in *Or83b*<sup>1</sup> and *Or83b*<sup>2</sup> mutants ( $P < 0.002$  and  $P < 0.001$ , respectively, Dunnett test). (F) Similarly, discrimination against d<sub>17</sub>-octanol was eliminated in *Or83b*<sup>1</sup> and *Or83b*<sup>2</sup> mutants ( $P < 0.001$  for both, Dunnett test).



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balanced distribution of the flies in the maze arms, indicative of equal response to the two isotopes (i.e., balanced maze). Similarly, d<sub>17</sub>-1-octanol was preferentially discriminated against versus equal concentration of the normal odorant. In this case, however, the deuterated isotope appeared even more aversive, as 75% reduction in its concentration equalized aversion with h-1-octanol (Fig. 1C). In contrast, flies did not exhibit spontaneous differential avoidance of d<sub>5</sub>-benzaldehyde, which occurred only upon reducing the concentration of the normal odorant (Fig. 1D). These differences cannot result from reduced evaporation of the slightly heavier deuterated odorants because two of the three elicited increased aversion, known to be proportional to the concentration of airborne odorants in this T-maze system (14, 20, 21).

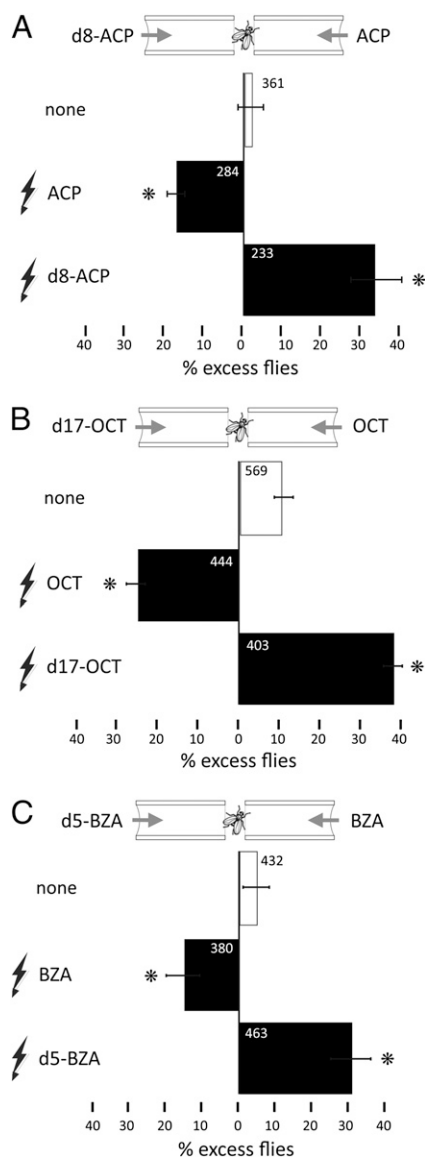
If the differential response to the deuterated odorants relied solely on olfaction and not any other sensory modality, it should be completely eliminated in anosmic mutants. To that end, we obtained two anosmic *Drosophila* strains carrying the null alleles *Or83b*<sup>1</sup> and *Or83b*<sup>2</sup> of the gene encoding the common subunit of the dimeric *Drosophila* olfactory receptor (13, 22, 23). Clearly, the spontaneous avoidance of deuterated d<sub>8</sub>-ACP and d<sub>17</sub>-1-octanol were eliminated and the mutant flies distributed equally in the maze arms as expected (Fig. 1E and F). Therefore, *Drosophila* use olfaction alone to discriminate between deuterated and normal odorants. Furthermore, the spontaneous discrimination against deuterated odorants indicates that they likely present to the flies recognizable, salient features distinct from their hydrogen counterparts.

**Conditioned Discrimination of Isotopes.** If the deuterated and normal isotopes of a pair exhibit salient odor character differences, then *Drosophila* should be able to associate either odorant with a punishing stimulus (20). To eliminate bias, the amounts of each odorant in a pair were adjusted as shown in Fig. 1B–D to yield

spontaneous preference as near zero as possible (i.e., balanced maze), so any postconditioning distribution changes would be a consequence of training. Flies were conditioned to associate electric foot-shock punishment with the presence of the deuterated or normal odorant of a pair. *Drosophila* successfully associated either odorant with punishment, demonstrated by the conditioned selective aversion of the shock-associated odor upon testing (Fig. 2). Flies shocked in the presence of the normal odorant distributed preferentially in the arm carrying its deuterated counterpart and vice versa. The shock-associated learning extended to h-benzaldehyde and d<sub>5</sub>-benzaldehyde, for which the flies exhibited no spontaneous preference, suggesting, as expected (24), that conditioned discrimination is independent from spontaneous preference.

This was not an exclusive property of the *w*<sup>1118</sup> control strain (14), as experiments were repeated with Canton-S, a different WT strain, with identical results (Fig. S2A and B). We also reversed the order of stimulus presentation, such that the shock-associated odor was delivered immediately before testing. Flies continued to selectively avoid the shock-associated odor (Fig. 2C), eliminating the possibility that they were simply attracted to the odor presented last in the absence of the shock reinforcer. Thus, deuterated and normal odorants present salient differences to the fly olfactory system, which can be used to predict punishment or its absence by association with either isotope.

**Generalization of Conditioned Isotope Discrimination Across Odorant Pairs.** The results so far are consistent with an unambiguous difference between deuterated and normal odorants for all three pairs tested. Is the difference associated with the majority ( $\geq 99\%$ ) compound or with impurities? Although the odorants used were of the highest purity available (Figs. S3 and S4), they



**Fig. 2.** Conditioned avoidance of normal and deuterated chemically identical odorants. The mean relative distribution of flies in the arms of the maze (% excess flies) carrying the indicated odorants  $\pm$  SEM is shown. The total number of flies tested in each group is shown and mean and SEM were derived from at least six repetitions per group. *Drosophila* was conditioned to selectively avoid the indicated member of an odorant pair by coupling it to electric foot shock (indicated by the lightning symbol). The complementary odor of each pair was used as the unpunished control. Each experiment included a naive group (open bar) to establish balancing of the odors used for testing and was used as control for all statistical comparisons on how conditioning resulted in subsequent selective avoidance of the punished odorants. Asterisks indicate significant differences that are significant. (A) Flies selectively avoided h-ACP or d<sub>8</sub>-ACP as expected based on the conditioning scheme and illustrated by their differential distribution in the arms of the maze. The performance of both groups was significantly different from that of naive animals ( $P < 0.001$ , Dunnett test). (B) Selective discrimination ( $P < 0.001$  vs. naive distribution; open bars) against h-octanol and d<sub>17</sub>-1-octanol in accord with the punished odorant during training. (C) Conditioned discrimination against h-BZA ( $P = 0.001$  vs. naive) and d<sub>5</sub>-BZA ( $P < 0.001$  vs. naive, Dunnett test).

nevertheless may contain small amounts of impurities that, in principle, could account for the spontaneous preferences and the shock-associated learning. To examine this possibility, we asked whether *Drosophila* could recognize deuterium as a salient feature from one odorant pair to another (i.e., generalize). If dif-

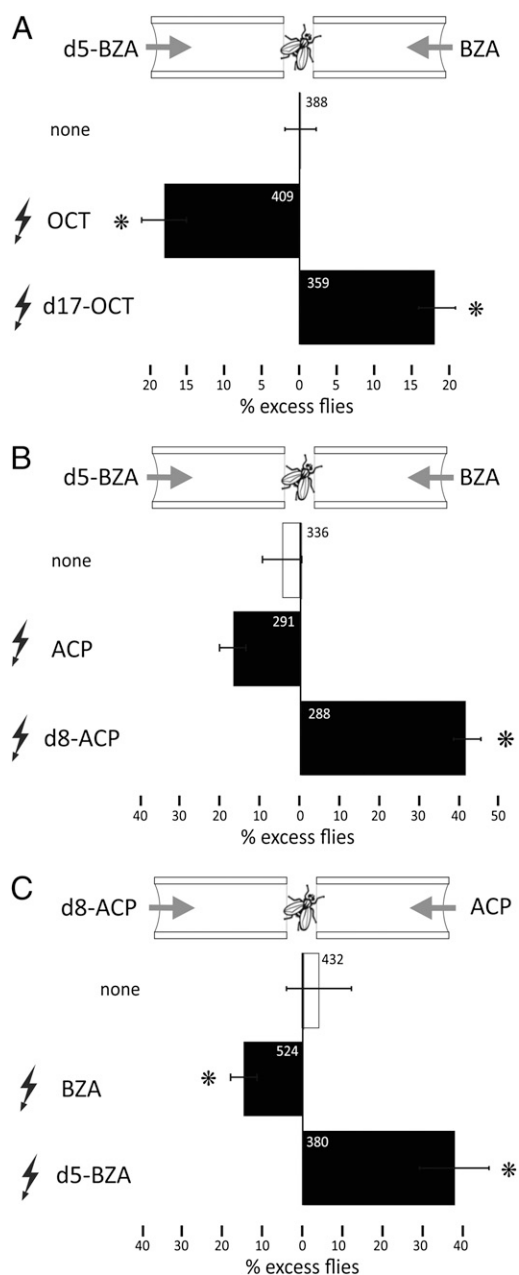
ferent impurities were present, salient features(s) of the training odors allowing their discrimination would be absent if testing with a different odorant pair. Hence, the flies would be unable to generalize (25) and thus distribute equally in the maze arms, as if faced with novel odors. In contrast, if the deuterium odorants had a common salient feature or odor character, then flies could, in principle, generalize between pairs.

To address this hypothesis, *Drosophila* were trained to selectively discriminate against deuterated or normal 1-octanol or deuterated versus normal ACP, but tested against the novel pair h-benzaldehyde versus d<sub>5</sub>-benzaldehyde. We found that flies trained to avoid h-1-octanol avoided h-benzaldehyde selectively when tested with the h-benzaldehyde/d<sub>5</sub>-benzaldehyde pair. Conversely, conditioning to discriminate against d<sub>17</sub>-1-octanol resulted in selective avoidance of d<sub>5</sub>-benzaldehyde (Fig. 3A). Similarly, flies trained to selectively avoid h-ACP discriminated preferentially against benzaldehyde and those trained against d<sub>8</sub>-ACP avoided the deuterated isotope selectively (Fig. 3B). Because h-ACP is attractive, the task of discriminating against the aversive h-benzaldehyde was expectedly more difficult than the converse choice, as indicated by the modest selective discrimination (Fig. 3B). To further substantiate these findings, we also trained with the h-benzaldehyde/d<sub>5</sub>-benzaldehyde pair, which do not elicit differential naive discrimination, and tested against normal and deuterated ACP. Again, flies exhibited differential avoidance of d<sub>8</sub>-ACP if conditioned with d<sub>5</sub>-benzaldehyde and of h-ACP if trained with h-benzaldehyde (Fig. 3C). Therefore, generalization occurred irrespective of the odorant pairs used for training and testing, suggesting that salient differences independent of their chemical identity and shape allow learned selective aversion to be transferred from the training to the testing pair.

Significantly, even if impurities were present, it is not expected that they would be similar in all three odorant pairs and account for differential conditioning, because the syntheses of the six compounds are different. Furthermore, the graded switch from attraction to aversion for a single molecule, ACP, clearly depended on the number of deuterium atoms it carried (Fig. 1A). Hence, it would be remarkable for an aversive impurity to fortuitously follow the number of deuterium atoms in that odorant. Nevertheless, could a single aversive impurity present only in every deuterated compound conceivably account for these results? If this were the case, it is difficult to explain how this impurity is ineffective in d<sub>5</sub>-benzaldehyde, consistent with the observed lack of spontaneous avoidance (Fig. 1D), but renders d<sub>8</sub>-ACP and d<sub>17</sub>-1-octanol aversive. Furthermore, it is possible that benzaldehyde masks the putative impurity, a potential explanation for the equal aversion of its two isotopes. However, if the putative impurity was masked, then, in contrast to our results (Fig. 3A and B), the benzaldehyde isotopes would not elicit differential discrimination in flies conditioned with 1-octanol/d<sub>17</sub>-1-octanol or ACP/d<sub>8</sub>-ACP. Collectively, then, impurities cannot account for the observed across odorant differential conditioned discrimination. In contrast, the generalization results are explained if we hypothesize that the salient cue(s) is a common property that differentiates the isotopes of each pair and distinguishes all deuterated odorants from their normal counterparts. As this cannot be molecular shape, differential evaporation, or impurities, it is likely that the common salient cue is the presence or absence of deuterium.

**Odorant Molecular Vibrations Are Salient Cues for Conditioning.** The doubling of hydrogen mass by deuteration will affect every molecular vibration in which movement of the hydrogen atoms occurs. When the vibrational mode involves mostly heavy atoms (i.e., C, N, O), the change in frequency is modest because of the relatively small change in mass of the deuterated versus normal compounds. In contrast, when most of the mode motion is in the hydrogen atoms themselves, as is the case for C-H stretch, wag, and scissor modes, the effect of deuteration on mode frequency will be large. A plausible biophysical mechanism for detection of molecular vibrations has been proposed previously (6), and a detailed study of the underlying physics was recently presented





**Fig. 3.** *Drosophila* can be conditioned to selectively avoid deuterium. The mean relative distribution of flies in the arms of the maze (% excess flies) carrying the indicated odorants  $\pm$  SEM is shown and the total number of flies in each group is denoted. The mean was derived from at least six repetitions per group. *Drosophila* was conditioned with the indicated pairs of isotopes, but tested against a different pair as indicated. The test odorants were adjusted such that avoidance of naive animals was as near zero as possible (open bar) and was used to compare the distribution of conditioned animals. (A) Animals conditioned with h-1-octanol, but tested after training with h-BZA versus d<sub>5</sub>-BZA exhibited significantly different distribution than that of naive animals ( $P < 0.001$  vs. naive for both, Dunnett test). Animals trained with d<sub>17</sub>-1-octanol selectively avoided the deuterated test odorant and vice versa. (B) Animals punished to h-ACP discriminated selectively against h-BZA, albeit not as robustly as previously described ( $P = 0.004$  vs. naive, Dunnett test), but ones trained to avoid d<sub>8</sub>-ACP avoided the deuterated BZA efficiently ( $P < 0.001$  vs. naive, Dunnett test). (C) Flies conditioned to with h-BZA/d<sub>5</sub>-BZA exhibited efficient selective avoidance of d<sub>8</sub>-ACP and h-ACP, respectively ( $P < 0.001$  vs. naive for both, Dunnett test).

(26). If indeed flies are detecting deuteration by sensing molecular vibrations, it is not unreasonable to suppose that they do so by detecting the modes most affected by deuteration, e.g., the C-H stretch. Each C-H bond has a slightly different stretch frequency clustered around approximately  $3,000\text{ cm}^{-1}$ . Deuteration reduces the stretch frequencies by approximately the square root of 2, to  $2,100$  to  $2,200\text{ cm}^{-1}$ . This in turn could confer the salient olfactory difference used to distinguish between isotopes and discriminate them differentially after conditioning.

The fact that the C-D stretch occurs in the  $2,200\text{ cm}^{-1}$  region is useful because only a few triple-bonded functional groups possess vibrations in this region, such as acetylenes, nitriles, isonitriles, and azides. If our hypothesis is correct, flies should be able to generalize the C-D stretch vibration in deuterated compounds to a similar vibration not carried by the C-D stretch, but instead by a triple-bonded functional group similar only in stretch frequency.

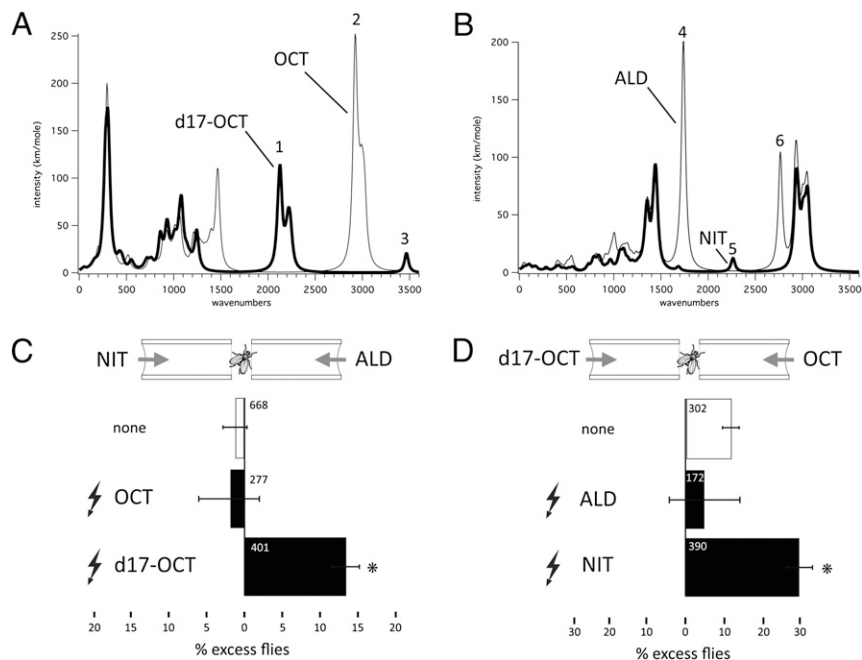
Therefore, the test pair should be molecules of similar odor, only one of which includes a triple bond. We took advantage of the fact that, at least to humans, many nitriles possess odors similar in character to the corresponding aldehydes. A well known such aldehyde-nitrile pair is citronellal (3,7-dimethyloct-6-enal) and citronellyl nitrile (3,7-dimethyloct-6-enenitrile), both citrus compounds with a lemongrass smell, with the nitrile slightly more metallic/oily (27). Consistent with what we perceive as their similar odor character, flies did not show spontaneous preference for or against citronellal or citronellyl nitrile (Fig. 4C). Moreover, in agreement with these empirical observations, the calculated IR spectra of the two compounds in the fingerprint region are very similar (Fig. 4B). The main difference lies in the vibrations of the aldehyde with a C=O stretch around  $1,740\text{ cm}^{-1}$  and the aldehydic C-H stretch around  $2,765\text{ cm}^{-1}$ . The nitrile lacks these two vibrations, replaced by a -CN stretch near  $2,265\text{ cm}^{-1}$ . The  $2,200\text{ cm}^{-1}$  region is the only one shared by d<sub>17</sub>-1-octanol and citronellyl nitrile but not by citronellal or octanol. Therefore, in conjunction with the d<sub>17</sub>-1-octanol/h-1-octanol pair, we now have two pairs of chemically unrelated molecules in which vibrations in the  $2,200\text{ cm}^{-1}$  region are present or absent.

If the salient character recognized in the deuterated odor is indeed the C-D stretch vibrations, flies should be able to generalize the odor character from C-D to C $\equiv$ N and vice versa. Because the C-D vibration is at the same energy as the nitrile's C $\equiv$ N stretch, the nitrile should be selectively avoided by animals conditioned to avoid d<sub>17</sub>-1-octanol. The results of experiments designed to test this idea are shown in Fig. 4C and D. Flies trained to discriminate against d<sub>17</sub>-1-octanol selectively avoided the citronellyl nitrile, whereas flies trained to avoid h-1-octanol did not show preferential avoidance (Fig. 4C). The latter outcome was not unexpected, as both citronellyl compounds have similar spectra aside from the functional group region, which do not overlap that of h-1-octanol, thus lacking recognizable salient features to guide behavioral responses. Furthermore, flies trained to avoid citronellyl nitrile discriminated selectively against d<sub>17</sub>-1-octanol, whereas discrimination against citronellal did not yield selective avoidance of h-1-octanol for the reasons outlined earlier. Therefore, in accord with our hypothesis, the presence of nitrile or deuterium resulted in cross-learning in either direction. Moreover, these results further constraint possible explanations invoking a hypothetical impurity, because they can be explained only if d<sub>17</sub>-1-octanol contains traces of citronellyl nitrile. This seems unlikely, not least because no nitrile solvent was used during its synthesis. As the nitrile is not deuterated, these results also make it very unlikely that the isotope effects we observe are caused by a property of isotopes unrelated to their vibrational modes.

## Discussion

Flies, like humans, perceive odor quality and intensity and can be conditioned to discriminate differences in its concentration (28, 29), and at sufficiently different concentrations, the same odorant may appear as distinct qualities (30). Therefore, it can be argued that flies discriminate d<sub>8</sub>-ACP and d<sub>17</sub>-1-octanol from their normal isotopes based on putative (small) concentration

**Fig. 4.** *Drosophila* can be conditioned to selectively avoid a vibrational frequency. (A) Computed vibrational spectra (IR intensities) of h-OCT versus d<sub>17</sub>-OCT. Salient spectral peaks are indicated on the graphs and show that deuteration of octanol shifts the group of C-H stretch vibrations from around 3,000 cm<sup>-1</sup> to 2,150 cm<sup>-1</sup>. Deuteration also shifts downward all of the peaks on the fingerprint region (1,000–1,500 cm<sup>-1</sup>). (B) Computed IR intensities of citronellal versus citronellyl nitrile with the salient spectral peaks indicated. The spectra of citronellal and citronellyl nitrile are remarkably similar in the fingerprint region. They differ chiefly in the vibrations involving the terminal functional group: in citronellyl nitrile the aldehyde carbonyl stretch around 1,750 cm<sup>-1</sup> is absent, replaced by a nitrile stretch around 2,150 cm<sup>-1</sup>. The low-lying aldehyde C-H stretch vibration is also absent. The vibration band centered at 2,150 cm<sup>-1</sup> is the only one common to d<sub>17</sub>-1-octanol and citronellyl nitrile but not present in h-octanol or citronellal. The spectra were computed using the Amsterdam Density Functional software at DZP/PBE level of theory. (C) *Drosophila* selectively avoid the molecular vibrations of deuterium. Flies conditioned to selectively avoid d<sub>17</sub>-1-octanol exhibited strong preferential avoidance of citronellyl nitrile ( $P < 0.001$  vs. naive), but flies punished to h-octanol did not selectively avoid citronellal ( $P = 0.691$  vs. naive). The only common element potentially recognizable in the test odor pair to aid in selective avoidance is the overlap in the vibrational spectrum of the C-D bonds in d<sub>17</sub>-1-octanol and the C≡N triple bond in citronellyl nitrile as illustrated in A and B. In contrast, they were not selective toward a novel odor without any recognizable molecular features. (D) In the converse experiment, flies conditioned to selectively avoid citronellyl nitrile exhibited highly significant avoidance of d<sub>17</sub>-1-octanol as a testing odor ( $P < 0.001$  vs. naive), but flies punished to citronellal did not selectively avoid h-octanol ( $P = 0.999$  vs. naive).



differences because of the reduced evaporation of the heavier deuterated molecules. However, such differences would have to be greater than 30% (28), which is rather improbable under our experimental conditions, and flies seem to generalize concentration differences smaller than that (29). This explanation is additionally improbable because, except for benzaldehyde, the “heavier” deuterated compounds are more aversive than their normal isotopes. Moreover, this hypothesis is inconsistent with the identification of deuteration as a salient mediator of conditioned responses across chemically distinct odors and with the selective aversion of molecules with vibrational resonance in the range of the C-D stretch. Therefore, a parsimonious explanation for our results is strongly indicative of a molecular vibration-sensing component in *Drosophila* olfaction. This is also consistent with independent recent evidence suggesting contributions of the vibrational spectra of odorants in the electrophysiological response of isolated *Drosophila* olfactory receptors (31).

In the past there have been four main objections to the notion that olfaction detects molecular vibrations. First, that molecular shape is an adequate predictor of smell (2, 30, 32), which currently seems unlikely (33, 34). In fact, *Drosophila* has only 62 olfactory receptors (35), suggesting that a single receptor must generally respond to multiple odorants, but in addition that a single odorant can activate multiple receptors (10, 15). Clearly, therefore, the structural recognition mechanism alone does not suffice to explain odorant recognition, suggesting that additional properties of these volatile molecules likely contribute to the process. The second objection was to the main assertion of vibrational theory that odorants with similar spectra should produce similar olfactory responses and, conversely, molecules of identical molecular shape but distinct spectra should smell different. Our data are consistent with this hypothesis, at least in flies, and suggest that with gas chromatography-pure odorants, and perhaps with the use of aversive conditioning, similar effects may be revealed for vertebrates—even humans—as shown for perceptual discrimination of enantiomers (36). The third objection has been that no known biological mechanism could behave as the equivalent of a vibrational spectroscope. However, the proposal (6) that olfaction uses inelastic electron tunneling

spectroscopy (IETS) has made the idea physically plausible. IETS is a quantum mechanism whereby electrons move from a donor to an acceptor site at constant total energy, although the acceptor is energetically lower than the donor (37). To satisfy conservation of energy, tunneling occurs only if a molecule is present between donor and acceptor, possessing vibrational mode(s) at or near this excess energy, absorbing it, and becoming excited. A modified IETS mechanism appropriate to proteins was recently described (26), suggesting a testable model applicable to olfactory receptors. Finally, it was objected that enantiomers with identical vibrations should always smell the same, whereas some smell different (38). Given that proteins are chiral, a shape-only theory cannot account for the identical odors of most enantiomer pairs. In contrast, this objection is easily answered by IETS because it exhibits the pronounced polarization effects expected when sensing molecules in fixed orientations such as enantiomers (6).

Individual olfactory receptors in *Drosophila* and other species can mediate excitatory and inhibitory responses to different odorants (10). Our data suggest that the vibrational mode and frequency of particular atoms and active groups of odorant molecules may also provide discriminatory cues that could broaden the recognition repertoire of odorant receptors, but still retain specificity. Currently, we do not know whether the same or different olfactory receptors are recruited to sense the normal and deuterated versions of the odorant molecules, and this question is currently under investigation broadly guided by the following considerations. A single receptor may recognize, by broad odotope features, a given odorant whose particular vibrational resonance may contribute to the odor-specific activity patterns of odorant receptor neurons (39), potentially modifying its particular odor character. This hypothesis predicts that receptors with distinct shape selectivities must also recognize similar vibrational modes and frequencies to explain our data of selective avoidance of citronellyl nitrile after conditioning with d<sub>17</sub>-1-octanol and vice versa. By analogy to color vision, it is possible that the multiple olfactory receptors may be divided into spectral classes, the members of each class sensing the same vibrational range and differing in their affinity for molecules of different shapes and physical properties.

For example, we would expect to find several receptors in the  $2,150\text{-cm}^{-1}$  band, able to sense deuterated molecules but not their undeuterated counterparts.

If the proposed vibrational sensing component is arranged in broad frequency bands of  $100$  to  $200\text{ cm}^{-1}$  wide,  $10$  to  $20$  receptors would suffice to cover the entire vibrational range. However, why then do flies (and mammals) have many more receptors (2, 3)? A possible answer may be that the olfactory system has evolved to reconcile conflicting requirements, namely, to sense a wide range of odorants, thus being nonspecific, but also to possess high affinity to detect them in small concentrations. Therefore, multiple receptors may serve to ensure that one or more will always bind with adequate affinity to any molecule to provide broad olfactory recognition through the combined activities of multiple receptors. Testing these and other predictions is well within reach of available behavioral, imaging, and genetic techniques.

## Materials and Methods

*Drosophila* were cultured as described previously (40) under a 12-h dark, 12-h light cycle. Mixed-sex groups of 2- to 3-d-old flies were lightly anesthetized and segregated in groups of 50 to 70 animals in vials containing food. Twenty-four hours later they were changed to fresh vials and placed in the dark to adapt 2 to 3 h before behavioral experiments commenced. Details of the behavioral procedures can be found in *SI Materials and Methods*.

All behavioral experiments were performed at  $25^\circ\text{C}$  and 80% to 85% relative humidity. Training and testing were performed in complete darkness, with dim red light used only during manipulations not requiring exposure to odors. The standard T-maze was modified by replacing the odorant holding cups (20) with glass cylinders of 2.25 cm diameter and 14 cm height as described previously (19, 40). An air stream of 600 mL/min passing

over the meniscus carried the odors to the maze arms via silicon rubber tubing. The meniscus area was kept constant by maintaining the volume of the odorant and the solvent, isopropyl myristate (Fluka), at a total of 1 mL in all experiments. Deuterated compounds were from CDN Isotopes. The amounts of odorants added to isopropyl myristate to yield equivalent avoidances were 200  $\mu\text{L}$  of 1-octanol (Fluka) and 50  $\mu\text{L}$  of the fully deuterated  $d_{17}$ -1-octanol, 90  $\mu\text{L}$  of benzaldehyde (Fluka) and 90  $\mu\text{L}$  of perdeuterated benzaldehyde- $d_5$  150  $\mu\text{L}$  of ACP (Fluka) and 75  $\mu\text{L}$  of the perdeuterated  $d_8$ -ACP, 100  $\mu\text{L}$  of citronellal and 100  $\mu\text{L}$  of citronellyl nitrile (gift of International Flavors and Fragrances). As for  $d_8$ -ACP (C6D5COCD3), 75  $\mu\text{L}$   $d_3$ -ACP (C6H5OCD3),  $d_5$ -ACP (C6D5OCH3), and normal odorant were also used for discrimination experiments. Different sets of silicone rubber tubing holding cylinders and maze arms were used for each odor. Complete descriptions of the behavioral procedures are presented in *SI Materials and Methods*.

**Computation of IR Intensity Spectra.** Spectra were computed by using the Amsterdam Density Functional software package ([www.scm.com](http://www.scm.com)) at DZP/PBE level of theory. Frequencies were computed numerically by differentiation of energy gradients in slightly displaced geometries. The force constants and hence the frequencies were computed by comparison of the gradients (in the harmonic approximation of the energy surface). Under these conditions, intensities are proportional to the change in dipole occurring during atom movements in a given vibrational mode (41). Accuracy of mode calculations is typically fewer than 10 wave numbers for molecules comprising C, O, N, and H atoms.

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