

African monkeys are infected by *Plasmodium falciparum* nonhuman primate-specific strains

Franck Prugnolle^{a,1,2}, Benjamin Ollomo^{b,1}, Patrick Durand^{a,1}, Erhan Yalcindag^{a,1}, Céline Arnathau^a, Eric Elguero^a, Antoine Berry^c, Xavier Pourrut^{a,d}, Jean-Paul Gonzalez^e, Dieudonné Nkoghé^d, Jean Akiana^f, Delphine Verrier^e, Eric Leroy^{a,d}, Francisco J. Ayala^{g,2}, and François Renaud^{a,2}

^aLaboratoire Maladies Infectieuses et Vecteurs Écologie, Génétique, Évolution et Contrôle, Unité Mixte de Recherche 224 Centre National de la Recherche Scientifique-Institut de Recherche pour le Développement-UM1, Institut de Recherche pour le Développement Montpellier, BP 64501, 34394 Montpellier Cedex 5, France; ^bUnité de Parasitologie Médicale, ^cUnité des Maladies Virales Emergentes, Unité Mixte de Recherche 190 Institut de Recherche pour le Développement-Université de la Méditerranée, and ^dUnité de Recherche en Ecologie de la Santé, Centre International de Recherches Médicales de Franceville, BP 769, Franceville, Gabon; ^eService de Parasitologie-Mycologie, Centre Hospitalier Universitaire de Toulouse and Unité Mixte de Recherche 152 UPS-Institut de Recherche pour le Développement, Université de Toulouse, 31059 Toulouse Cedex 9, France; ^fService Epidémiologie Moléculaire et Parasitaire, Département de la Médecine Préventive et des Essais Cliniques, Laboratoire National de Santé Publique, Brazzaville, République du Congo; and ^gDepartment of Ecology and Evolutionary Biology, University of California, Irvine, CA 92697-2525

Contributed by Francisco J. Ayala, June 10, 2011 (sent for review March 15, 2011)

Recent molecular exploration of the *Plasmodium* species circulating in great apes in Africa has revealed the existence of a large and previously unknown diversity of *Plasmodium*. For instance, gorillas were found to be infected by parasites closely related to *Plasmodium falciparum*, suggesting that the human malignant malaria agent may have arisen after a transfer from gorillas. Although this scenario is likely in light of the data collected in great apes, it remained to be ascertained whether *P. falciparum*-related parasites may infect other nonhuman primates in Africa. Using molecular tools, we here explore the diversity of *Plasmodium* species infecting monkeys in Central Africa. In addition to previously described *Hepatozoon* and *Plasmodium* species (*Plasmodium gonderi* and *Plasmodium* sp. DAJ-2004), we have found one African monkey to be infected by a *P. falciparum*-related parasite. Examination of the nuclear and mitochondrial genomes of this parasite reveals that it is specific of nonhuman primates, indicating that *P. falciparum*-related pathogens can naturally circulate in some monkey populations in Africa. We also show that at least two distinct genetic entities of *P. falciparum* infect nonhuman primates and humans, respectively. Our discoveries bring into question the proposed gorilla origin of human *P. falciparum*.

human malaria | laverania clade | parasite host transfer | cytochrome *b* | biological evolution

Until very recently, only one species (*Plasmodium reichenowi*) was known to be a phylogenetic sister lineage of *Plasmodium falciparum*, the agent of malignant human malaria. These two species were the only known representatives of the subgenus *Laverania* (1). In 2009 and 2010, several studies have revealed the existence of a number of distinct phylogenetic species belonging to this subgenus and infecting chimpanzees, bonobos, and gorillas in Africa (2–6), drastically modifying our understanding of the evolution of the *Laverania* lineage, and *P. falciparum* in particular (7).

Liu et al. (4) have proposed that the human strains of *P. falciparum* arose after a single gorilla-to-human transmission (4, 7). This scenario is based on the discovery of *P. falciparum*-related strains naturally circulating in lowland gorillas (2, 4, 6) that display greater genetic diversity than human strains (4). In contrast, no *P. falciparum* strains were found in wild chimpanzees or bonobos despite similar sampling efforts (4). Strains of *P. falciparum* had been found only in chimpanzees and bonobos living either in captivity or sanctuaries, with the possibility of a cross-species infection from humans to them (2, 3).

It, however, remains possible that *P. falciparum*-related strains circulate in wild chimpanzees and bonobos at a lower prevalence than in gorillas (8) and/or in geographic areas not yet explored. Given the propensity of *Plasmodium* parasites to switch hosts

(e.g., the transfer of *P. knowlesi* from macaques to humans (9) and *P. vivax* and *P. malariae* between humans and New World monkeys; ref. 10), one cannot exclude that *P. falciparum*-related strains also circulate in other African nonhuman primate species.

We report an analysis of the diversity of *Plasmodium* species infecting 10 monkey species in Africa and show that *P. falciparum* nonhuman primate-specific strains circulate in some of these primates. This finding challenges the gorilla origin of the human strains of *P. falciparum*.

Results and Discussion

We analyzed blood samples from 338 monkeys from Gabon, including 10 different species of nonhuman primates, *Cercopithecus nictitans*, *Cercopithecus cephus*, *Cercopithecus pogonias*, *Cercopithecus neglectus*, *Cercopithecus solatus*, *Cercocebus torquatus*, *Mandrillus sphinx*, *Miopithecus talapoin*, *Miopithecus ougouensis*, and *Lophocebus albigena*. Approximately four percent (12 individuals) of the 338 monkeys were positive in a PCR array for *Plasmodium* cytochrome *b* (*cyt b*) (Table 1). As expected from previous studies (11–14), we detected infections by *Hepatozoon* spp. (12), *Plasmodium gonderi* (11, 12), and *Plasmodium* sp. DAJ-2004 (13, 14) (Fig. 1). Although *Plasmodium* sp. DAJ-2004 was originally isolated and molecularly characterized from a drill (*Mandrillus leucophaeus*) (14), we have now found that it also infects mandrills (*M. sphinx*) and some guenon species (*C. nictitans* and *C. cephus*).

Surprisingly, we found one individual *C. nictitans* infected with a *P. falciparum*-related parasite (isolate MO454; Fig. 1). The presence of this *P. falciparum*-related parasite in *C. nictitans* was confirmed three times by using independent extraction products and PCRs. All PCR products were cloned before sequencing to avoid potential chimeric sequences. The individual carrying this parasite was a wild-born, adult *C. nictitans* kept as a pet in the Gabonese village of Oguewa (Moyen Ogooué province; Fig. S1). The host species was genetically confirmed by the analysis of its 12S mitochondrial sequence (*Materials and Methods*).

We tested the hypothesis that this monkey living in close contact with humans could have become infected by a human

Author contributions: F.P., B.O., P.D., E.L., F.J.A., and F.R. designed research; P.D., E.Y., and C.A. performed research; A.B., X.P., J.-P.G., D.N., J.A., D.V., and E.L. contributed new reagents/analytic tools; F.P., P.D., and E.E. analyzed data; and F.P., F.J.A., and F.R. wrote the paper.

The authors declare no conflict of interest.

¹F.P., B.O., P.D., and E.Y. contributed equally to this work.

²To whom correspondence may be addressed. E-mail: fjayala@uci.edu, franck.prungnolle@ird.fr, or francois.renaud@ird.fr.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1109368108/-DCSupplemental.

Table 1. *Plasmodium* species determined by cytochrome *b* sequences in African monkeys

Host species	Sample size	Number of cytochrome <i>b</i> sequences obtained	<i>Plasmodium</i> species (number of sequences)
<i>Cercocebus torquatus</i>	14	0	–
<i>Cercopithecus cephus</i>	67	3	<i>P.sp. DAJ-2004</i> (1) <i>Hepatocystis sp.</i> (2)
<i>Cercopithecus neglectus</i>	3	0	–
<i>Cercopithecus nictitans</i>	29	2	<i>P.sp. DAJ-2004</i> (1) <i>P. falciparum</i> (1)
<i>Cercopithecus pogonias</i>	4	0	–
<i>Cercopithecus solatus</i>	2	0	–
<i>Lophocebus albigena</i>	4	0	–
<i>Mandrillus sphinx</i>	212	6*	<i>P.sp. DAJ-2004</i> (2) <i>P. gonderi</i> (4) <i>Hepatocystis sp.</i> (1)
<i>Miopithecus ogouensis</i>	2	0	–
<i>Miopithecus talapoin</i>	1	1	<i>Hepatocystis sp.</i> (1)

*One *Mandrillus sphinx* was mix-infected by *Plasmodium sp. DAJ-2004* and *P. gonderi*.

P. falciparum after its arrival at the village. We first tried to determine by qPCR the alleles present at two genes involved in chloroquine resistance: *pfcr* and *pfmdr1*. In Gabon, the preva-

lence of the two point mutations conferring resistance (*pfcr* 76T and *pfmdr1* 86Y) in human *P. falciparum* populations is high (>85%) (15, 16). Hence, we hypothesized that if the MO454

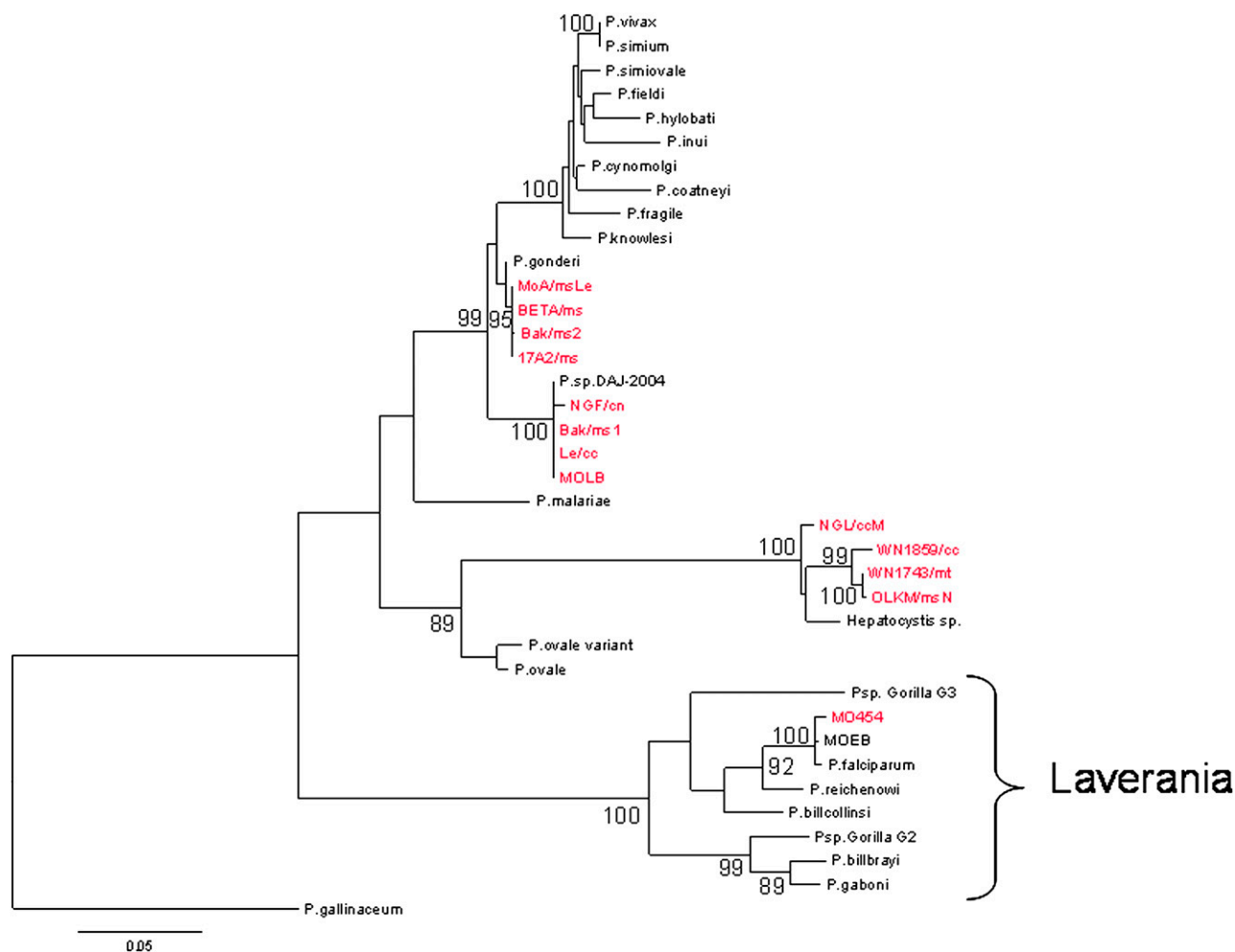


Fig. 1. Phylogenetic relationships among primate *Plasmodium* species based on a partial sequence (823 bp) of the cytochrome *b* gene. The tree was constructed by using maximum likelihood methods. The isolates genetically characterized in this study are highlighted in red. Bootstrap values >80% are indicated. *P. gallinaceum*, a parasite from birds, was used to root the tree. See Table S4 for the GenBank accession numbers of the sequences used in the tree. Note that *P. sp. Gorilla G2* and *P. sp. Gorilla G3* are the names given in ref. 4 to *P.sp GorA* and *P.sp Gor B*, respectively, as defined in ref. 6.

isolate was of very recent human origin, there would be a high probability to observe at least one resistance allele at one or both genes. We were unable to determine the genotype at *pfmdr1*. For *pfcr1*, the MO454 isolate (note that *Plasmodium* blood stages are haploid) displayed the wild allele at codon 76 (*pfcr1* K76). This allele is absent in the human *P. falciparum* population in Lambaréné (Moyen Ogooué province), a city <30 km away from the village where MO454 was collected (16) (Fig. S1). In another village of Gabon (Bakoumba, Haut Ogooué province; Fig. S1), the wild allele at codon 76 is only 6% (15). Such low prevalence of the wild-type allele in the human *P. falciparum* population in Gabon suggests a nonhuman origin for the MO454 isolate. However, we considered this evidence by itself was not sufficient to rule out its human origin.

We then determined the genotype of the MO454 isolate at seven nuclear microsatellite markers and 96 nuclear single nucleotide polymorphisms (SNPs) distributed all over the genome, and compared the multilocus genotypes of the MO454 isolate to a set of human African isolates genotyped for the same markers (see *Materials and Methods* for additional details) and collected from the same region of Africa (Gabon, Cameroun, and Congo). We computed assignment probabilities and performed a principal component analysis (PCA) for the two sets of markers. An assignment probability corresponds to the probability that a multilocus genotype belongs to a population given the allelic frequencies observed in this population. If the MO454 isolate was a very recent transfer from humans, it should display a high assignment probability to the human *P. falciparum* population. As shown in Fig. 2, MO454 displays a very low probability of assignment to the human *P. falciparum* population, whether computed using the microsatellite ($P = 0.0001$) or the SNP markers ($P = 0.0001$). The assignment probabilities computed for the human isolates were always higher (Fig. 2).

PCA results are congruent with the previous results. MO454 always displayed an outlier position on the first two axes compared with the human isolates, whether using the microsatellite or the SNP markers (Fig. 3). Thus, our analyses suggest that the MO454 isolate comes from a population of *P. falciparum* that is genetically differentiated and isolated from the current human *P. falciparum* population circulating in central Africa. These results, in turn, suggests that MO454 is not of very recent human origin and, therefore, that the monkey was most likely already infected before its arrival at the Gabonese village.

Two scenarios can be considered regarding the nature of the population from which MO454 originates: (i) this population diverged from a population of human *P. falciparum* several generations ago, and had time to diverge from it because of adaptation to the host and/or genetic drift with limited exchanges of migrants. In this case, the population would have a human origin. (ii) The population has evolved for a long time in non-human primates. In this case, the population would have a simian origin.

To test these alternatives, we sequenced the entire mitochondrial genome of the MO454 isolate. Liu et al. (4) have shown that four SNPs of the *Laverania* mitochondrial genome make it possible to distinguish the *Plasmodium* infecting non-human primates from the *Plasmodium* infecting humans. We found that, for these four SNPs, the MO454 isolate displayed the same alleles as the ones found in nonhuman primate *Plasmodium* (Fig. 4), thus indicating that the MO454 strain did not come from a population of human origin. This hypothesis is further sustained by the high bootstrap value of the cluster of *P. falciparum* isolates of simian origin (which includes the MO454 isolate), which is clearly genetically differentiated from the cluster formed by the human isolates of worldwide origin (Fig. 4 and Fig. S2). In contrast, all strains isolated from captive bonobos by Krief et al. (3) fall within the *Plasmodium* genetic diversity found in humans and all display the four SNPs specific of the human *P. falciparum* (Fig. 4 and Fig. S2). This result, added to the presence of double or triple mutations associated with resistance to pyrimethamine in the *dhfr* gene (3), indicates that the bonobo strains arose from a direct transfer from humans.

Our results show that African monkeys can be infected by *P. falciparum* nonhuman primate-specific strains, in addition to the previously described *Hepaticystis* and *Plasmodium* species. Whether the *P. falciparum* infection we report in this study was due to a recent transmission from apes (e.g., gorillas) to monkeys or whether independent transmission foci exist in monkeys remains to be determined. In any case, our findings indicate that systematic surveys of the diversity of *Plasmodium* species circulating in primates and other mammals are needed to resolve the evolution of the *Laverania* subgenus. Our finding of a *P. falciparum*-related strain in a monkey suggests that *P. falciparum* may not be strictly specific to gorillas and humans, and opens up the possibility that other nonhuman primate species might also be naturally infected by *P. falciparum* parasites.

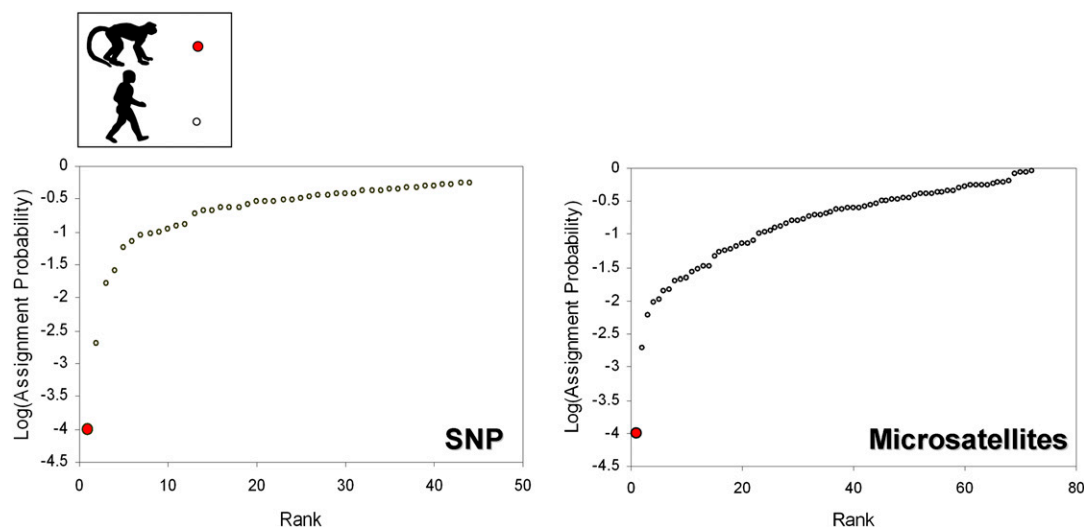


Fig. 2. Ranked logarithms of the assignment probabilities computed for human and nonhuman primate *P. falciparum* isolates obtained using SNP (Left) and microsatellite (Right) markers. The MO454's probability is represented by the red solid circle at bottom left.

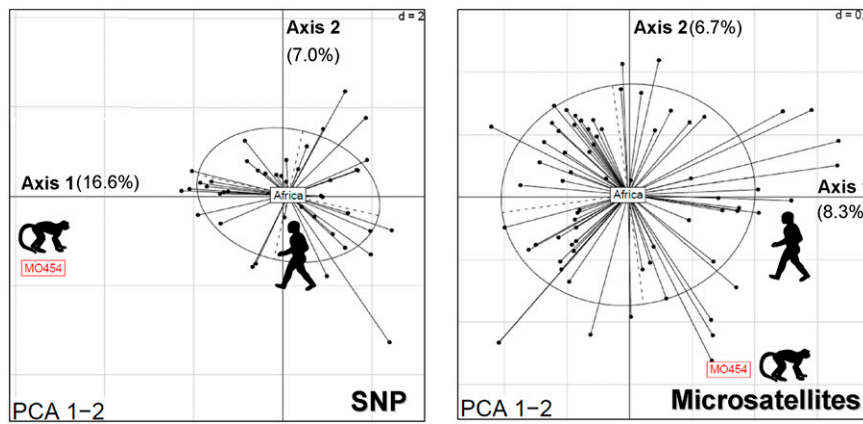


Fig. 3. Projection of the principal component analysis (PCA) on axes 1 and 2. Dots represent individuals (haplotypes). Ellipses are a graphical summary of the dispersion of the human *P. falciparum* haplotypes. The position of the MO454 isolate is indicated by the red box. The barycentre of the human population (“Africa”) is located at the center of the ellipse. *Left*, SNPs; *Right*, microsatellites. The percent of the variance explained by each axis is given next to each axis.

Our discovery brings into question Liu et al.’s (4) proposal that human *P. falciparum* arose after a single gorilla-to-human transmission event. Indeed, if *P. falciparum* appeared in humans after a single cross-species transmission from a nonhuman primate species, then a transfer from a monkey is as likely as a transfer from an ape. An alternative possibility is that *P. falciparum* diverged from *P. reichenowi* after the divergence of the human and chimpanzee lineages and then experienced a recent radiation in primates in Africa, including great apes and monkeys (6, 17). The

fact that the genetic diversity observed in human isolates is lower than the genetic diversity observed in nonhuman primates might be due to recent demographic events (see e.g., ref. 18). There is a need for additional investigations to decide among these alternative hypotheses. Determining the mosquito species involved in the transmission of the nonhuman primate *P. falciparum* (which are not known) and their feeding behavior and host preferences might allow the assessment of the likelihood of an ape-to-human transmission. Similarly, estimating with accuracy the dates of

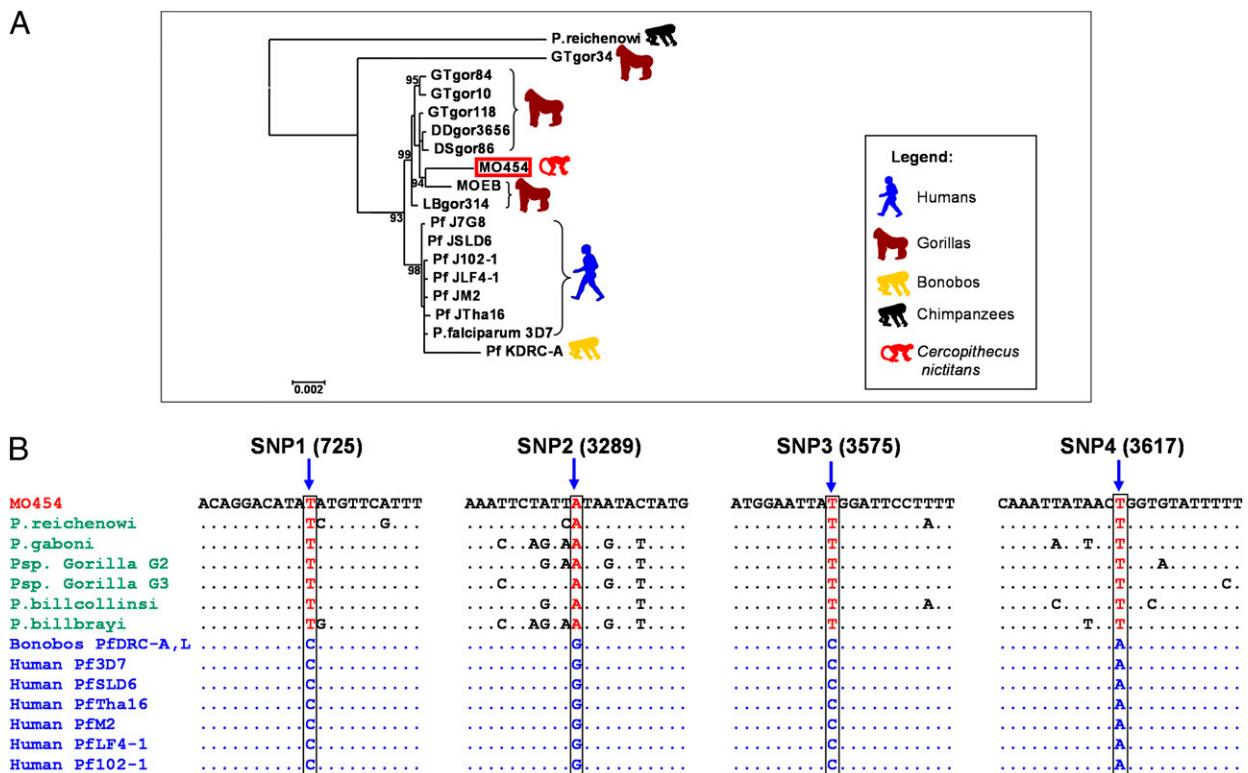


Fig. 4. (A) Phylogenetic relationships among *P. falciparum* isolates obtained with mitochondrial genomes ($\approx 5,800$ bp). The tree was constructed by using maximum likelihood methods. Bootstrap values (in %) are indicated at each node. *P. reichenowi* was used to root the tree. See Table S4 for GenBank accession numbers of the different sequences used in the tree. (B) The four diagnostic mitochondrial SNPs observed in MO454 compared with those present in *Plasmodium* isolates of nonhuman (i.e., chimpanzees, gorillas, and bonobos) and human origin. *Plasmodium* isolated from bonobos by Krief et al. (3), PfDRC-A, and PfDRC-L (as well as those from PfDRC-E and PfDRC-C, not shown in the figure) display the same SNP as human isolates. Note that *P. sp. Gorilla G2* and *P. sp. Gorilla G3* are the names given by ref. 4 to *P. sp. GorA* and *P. sp. Gor B*, respectively, as defined by ref. 6.

divergence between the different *P. falciparum* lineages and between *P. reichenowi* and *P. falciparum* would help to clarify the evolutionary history of *P. falciparum*.

What can be readily concluded from previous studies and from this one is that the diversity of *P. falciparum* is large, and so is the range of host species that it can naturally infect. It now becomes important to ascertain the origin of this diversity and to determine which genomic characteristics have allowed this lineage to become a generalist parasite. At least two distinct genetic entities of *P. falciparum* are present, one infecting apes and other nonhuman primates and another one infecting humans (Fig. 4). It will be worthwhile to ascertain the specific genomic adaptations of *P. falciparum* to the human host (19, 20).

The capacity of *P. falciparum* to switch hosts represents a risk that new strains may be transferred from nonhuman primates to humans, and vice versa. Although there is still no definitive evidence that nonhuman primates can serve as reservoirs of *P. falciparum* for humans, the contrary is not true. It is now apparent that the bonobos described by Krief et al. (3) were infected by human strains because they lived in close contact with humans. Should *Plasmodium* infections also prove seriously deleterious for nonhuman primate health, this additional factor could have major implications for the conservation of these already threatened species (International Union for Conservation of Nature, <http://www.iucn.org/>), although the evidence available indicates that *P. falciparum* is much less malignant for apes than it is for humans (19, 20). Given the steadily increasing contacts between humans and nonhuman primate populations due to human population growth and the increasing development of anthropic activities (e.g., logging, mining, commercial hunting) in tropical regions, the risk of emergence of these pathogens may not only have an impact on public health but also seriously compromise the preservation and persistence of nonhuman primate populations in the long term. There is a need to evaluate this risk. The identification as well as the description of the ecology and trophic behaviors of the vectors of the nonhuman primate *Plasmodium* species seems now all important.

Materials and Methods

Nonhuman Primate Sampling. All procedures in this study involving animals were approved by the Government of the Republic of Gabon and the Game and Wildlife Department of Libreville, Gabon (no. CITES 00956). The procedures complied with the relevant national and international guidelines. The samples were collected from wild-born nonhuman primates kept as pets by villagers in Gabon, as well as from 142 mandrills kept in a semifree ranging enclosure at the Centre International de Recherches Médicales de Franceville (CIRMF, Gabon). Blood samples were collected in 7-mL EDTA vacutainers from sedated animals (Ketamine; 10 mg/kg i.m.). Clots and plasma were obtained by centrifugation and stored at -20°C until transported to CIRMF and stored at -80°C until further analyses.

DNA Extraction, PCR Conditions, and Sequencing. Total DNA (*Plasmodium* and host) was isolated and purified from blood by using the DNeasy blood kit (Qiagen) according to the manufacturer's instructions. The *Plasmodium cytochrome b* (*cyt b*) gene was amplified and sequenced by using the protocols described in Prugnolle et al. (6) for diagnostic purposes. In addition to the *cyt b* gene, a set of other loci were amplified, genotyped, or sequenced in the isolate related to *P. falciparum* (see below).

First, we characterized the mutations present at codon 76 of the gene *pfcr*t and codon 86 of the gene *pfmdr*1 by using real-time PCR assays. Detection of *pfcr*t T76 mutations was done by using a real-time PCR method described (21). Detection of *pfmdr*-1 Y86 mutations was performed with the same quantitative real-time PCR assay by using fluorescence resonance energy transfer (FRET) hybridization probes. The method included amplification of a fragment of the *pfmdr*1 gene of *P. falciparum*, coupled with simultaneous detection of the product by probe hybridization and analysis of the melting curve by using real-time PCR. The *pfmdr*1 gene was amplified by using primers F186 (5'-GTATTATCAGGAGGAACATTACC-3') and R186 (5'-CCACAAACATAAATTAACGCA-3'). The amplified product was detected with two probes: the sensor 86 probe, 3'-labeled with fluorescein (5'-TTAATATCATCACCTAAATACATg-FL-3'), which hybridized with the re-

gion containing the mutation sites, and the anchor 86 probe, 5'-labeled with LC-Red 640 and 3'-phosphorylated (5'-LC640-TCTTAATATTACACC-AAACACAgATAT-PH-3'). Primers and probes were provided by Tib MolBiol (Berlin). The PCR and hybridization reactions were carried out on a Light-Cycler carousel-based system (Roche Diagnostics), in glass capillaries in a volume of 10 μL containing 2.5 μL of template DNA, 1 μL of MgCl_2 (25 mM), 1 μL of forward and reverse primers (5 μM each), 0.1 μL of sensor and anchor probes (20 μM each), and 1 μL of FastStart DNA Master Hybridization Probes (Roche Diagnostics). Briefly, qPCR was performed under standard conditions (1 cycle of 95°C for 9 min; 50 cycles of 95°C for 15 s, 50°C for 25 s, and 68°C for 7 s; 1 cycle of 95°C for 15 s, 40°C for 2 min, and a slow rise in the temperature to 65°C at a rate of 0.1 $^{\circ}\text{C}/\text{s}$ with continuous acquisition of fluorescence decline).

Each run of *pfcr*t and *pfmdr*1 PCR contained two DNA control samples of *P. falciparum*, F32-Tanzania (a chloroquine-sensitive, wild-type *pfcr*t K76 and *pfmdr*1 N86 strain) and FcM29-Cameroon (a chloroquine-resistant strain mutated for codon 76 of *pfcr*t and codon 86 of *pfmdr*1).

Second, we genotyped seven microsatellite markers. These microsatellites were originally discovered and designed on human *P. falciparum* strains but worked well on the primate *P. falciparum* (see Table S1 for a description of the microsatellites). PCR amplifications followed the protocols described in ref. 22. Fluorescence-labeled PCR products were sized on ABI Prism3110X genetic analyzer (Applied Biosystems), with a Genescan500LIZ internal size standard. For comparison purposes, the same microsatellites were genotyped in human *P. falciparum* samples coming from Gabon, Cameroon, and Congo (see Table S2 for a detail of the populations and sample sizes).

Third, we investigated the polymorphism at 96 SNPs distributed all over the genome and ascertained on human *P. falciparum*. This set of SNPs included SNPs located in coding (synonymous and nonsynonymous; introns and exons) as well as in noncoding regions (see Table S3 for a list of the SNPs). The SNP primers were defined according to the 3D7 sequence and genotyped by using the Golden Gate Illumina technology (Integragen Company). For comparison, the same SNPs were genotyped in a subset of the human samples genotyped with the microsatellites (Table S3).

Finally, we sequenced the entire mitochondrial genome of the *P. falciparum* isolate. The amplification was performed by following the protocol detailed in ref. 3. Approximately 5,800 bp (of 6,000) were amplified by using the oligos Forward 5'-GAGGATTCTCCACACTTCAATTCGTACTTC and Reverse 5'-CAGGAAATWATAGACCGAACCTTGGACTC with Takara LA Taq Polymerase (TaKaRa Mirus Bio). PCR products were cloned in the PGem-T vector (Promega) and sequenced by Eurofins. Note that in addition to the MO454 and for comparison purposes, we used the same method to sequence the entire mitochondrial genome of the *P. falciparum* isolate MOEB (isolated from a gorilla in Gabon) and we completed the mitochondrial genome of *P. gaboni* (GenBank accession no. FJ895307) described in Prugnolle et al. (6).

The host species *Cercopithecus nictitans* of the MO454 isolate was confirmed by the amplification and the sequencing of the host partial 12S rRNA gene using the protocol described in ref. 23.

Statistical Analyses. Phylogenetic tree. For phylogenetic analyses, we constructed trees by using maximum likelihood (ML) methods as described in ref. 6. The best-fitting ML model under the Akaike Information Criterion was GTR (General Time Reversible) + Γ (gamma distribution) for nucleotides as identified by ModelTest (24). The most likely DNA tree and corresponding bootstrap support values were obtained by PhyML (freely available at the ATGC bioinformatics platform; <http://www.atgc-montpellier.fr>) using nearest neighbor interchange branch swapping and 100 bootstrap replicates (25). Parasite isolates with their GenBank accession numbers are given in Table S4.

Assignment probability. Assignment probabilities were calculated for each individual according to the reference human *P. falciparum* population composed by the isolates coming from Gabon, Congo, and Cameroon.

To compute the assignment probability, we used the following procedure: first, genotype likelihood was computed by using the criterion of ref. 26. This criterion simply corresponds to the multinomial probability of the multilocus genotype given the allelic frequencies at each locus in the reference population, under an assumption of independence between loci. Note that each individual was removed from the reference population before calculating the observed allele frequencies (the leave-one-out procedure; ref. 27).

The assignment probability was then computed for each individual by using a MonteCarlo resampling method developed by ref. 28. This probability was computed by using the software GeneClass-2 (29) and 10,000 randomizations for both microsatellite and SNP markers.

Principal component analysis. Principal component analysis (PCA) was used to gain complementary insights into the relative position of the MO454

P. falciparum haplotype relatively to the diversity found in the human isolates from Central Africa. The PCA was performed on the matrix of binary allele profiles by using the R-packages Ade4 (30) and Adegenet (31). Missing values were replaced by the average frequencies of the corresponding alleles (31).

ACKNOWLEDGMENTS. We thank two anonymous referees for very helpful comments and suggestions, the staff of the CIRMF Primate Centre for pro-

viding the mandrill samples, and Jérôme Clain and Karen Day for helpful discussions and suggestions. This work was jointly supported by Centre National de la Recherche Scientifique, Institut de Recherche pour le Développement (IRD), and Centre International de Recherches Médicales de Franceville, as well as the Agence Nationale de la Recherche Malaria Genetic Adaptation to New Environments Programme Santé-Environnement et Santé-Travail 07 2012 and the Programme Maladie Infectieuses Emergentes 2009. B.O. was financed by an Institut de Recherche pour le Développement fellowship for 6 mo.

- Bray RS (1958) Studies on malaria in chimpanzees. VI. *Laverania falciparum*. *Am J Trop Med Hyg* 7:20–24.
- Duval L, et al. (2010) African apes as reservoirs of *Plasmodium falciparum* and the origin and diversification of the *Laverania* subgenus. *Proc Natl Acad Sci USA* 107: 10561–10566.
- Krief S, et al. (2010) On the diversity of malaria parasites in African apes and the origin of *Plasmodium falciparum* from Bonobos. *PLoS Pathog* 6:e1000765.
- Liu W, et al. (2010) Origin of the human malaria parasite *Plasmodium falciparum* in gorillas. *Nature* 467:420–425.
- Ollomo B, et al. (2009) A new malaria agent in African hominids. *PLoS Pathog* 5: e1000446.
- Prugnolle F, et al. (2010) African great apes are natural hosts of multiple related malaria species, including *Plasmodium falciparum*. *Proc Natl Acad Sci USA* 107: 1458–1463.
- Prugnolle F, et al. (2011) A fresh look at the origin of *Plasmodium falciparum*, the most malignant malaria agent. *PLoS Pathog* 7:e1001283.
- Holmes EC (2010) Malaria: The gorilla connection. *Nature* 467:404–405.
- Singh B, et al. (2004) A large focus of naturally acquired *Plasmodium knowlesi* infections in human beings. *Lancet* 363:1017–1024.
- Tazi L, Ayala FJ (2011) Unresolved direction of host transfer of *Plasmodium vivax* v. *P. simium* and *P. malariae* v. *P. brasilianum*. *Infect Genet Evol* 11:209–221.
- Coatney GR, Collins WE, Warren M, Contacos PG (1971) *The Primate Malariae* (US Government Printing Office, Washington, DC).
- Garnham PCC (1966) *Malaria Parasites and Other Haemosporidia* (Blackwell Scientific Publications, Oxford).
- Mu J, et al. (2005) Host switch leads to emergence of *Plasmodium vivax* malaria in humans. *Mol Biol Evol* 22:1686–1693.
- Escalante AA, Freeland DE, Collins WE, Lal AA (1998) The evolution of primate malaria parasites based on the gene encoding cytochrome b from the linear mitochondrial genome. *Proc Natl Acad Sci USA* 95:8124–8129.
- Maïga-Ascofaré O, et al. (2010) Adaptive differentiation of *Plasmodium falciparum* populations inferred from single-nucleotide polymorphisms (SNPs) conferring drug resistance and from neutral SNPs. *J Infect Dis* 202:1095–1103.
- Mayengue Pl, Kalmbach Y, Issifou S, Kreamsner PG, Ntoumi F (2007) No variation in the prevalence of point mutations in the Pfcrt and Pfmdr1 genes in isolates from Gabonese patients with uncomplicated or severe *Plasmodium falciparum* malaria. *Parasitol Res* 100:487–493.
- Silva JC, et al. (2010) Genome sequences reveal divergence times of malaria parasite lineages. *Parasitology*, 10.1017/S0031182010001575.
- Tanabe K, et al. (2010) *Plasmodium falciparum* accompanied the human expansion out of Africa. *Curr Biol* 20:1283–1289.
- Martin MJ, Rayner JC, Gagneux P, Barnwell JW, Varki A (2005) Evolution of human-chimpanzee differences in malaria susceptibility: Relationship to human genetic loss of N-glycolylneuraminic acid. *Proc Natl Acad Sci USA* 102:12819–12824.
- Varki A, Gagneux P (2009) Human-specific evolution of sialic acid targets: Explaining the malignant malaria mystery? *Proc Natl Acad Sci USA* 106:14739–14740.
- Vessièrè A, Berry A, Fabre R, Benoit-Vical F, Magnaval JF (2004) Detection by real-time PCR of the Pfcrt T76 mutation, a molecular marker of chloroquine-resistant *Plasmodium falciparum* strains. *Parasitol Res* 93:5–7.
- Anderson TJ, et al. (2000) Microsatellite markers reveal a spectrum of population structures in the malaria parasite *Plasmodium falciparum*. *Mol Biol Evol* 17:1467–1482.
- Kocher TD, et al. (1989) Dynamics of mitochondrial DNA evolution in animals: Amplification and sequencing with conserved primers. *Proc Natl Acad Sci USA* 86: 6196–6200.
- Posada D, Crandall KA (1998) MODELTEST: Testing the model of DNA substitution. *Bioinformatics* 14:817–818.
- Guindon S, Gascuel O (2003) A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Syst Biol* 52:696–704.
- Paetkau D, Calvert W, Stirling I, Strobeck C (1995) Microsatellite analysis of population structure in Canadian polar bears. *Mol Ecol* 4:347–354.
- Paetkau D, Shields GF, Strobeck C (1998) Gene flow between insular, coastal and interior populations of brown bears in Alaska. *Mol Ecol* 7:1283–1292.
- Paetkau D, Slade R, Burden M, Estoup A (2004) Genetic assignment methods for the direct, real-time estimation of migration rate: A simulation-based exploration of accuracy and power. *Mol Ecol* 13:55–65.
- Piry S, et al. (2004) GENECLASS2: A software for genetic assignment and first-generation migrant detection. *J Hered* 95:536–539.
- Dray S, Dufour AB (2007) The ade4 package: Implementing the duality diagram for ecologists. *J Stat Softw* 22:1–20.
- Jombart T (2008) adegenet: A R package for the multivariate analysis of genetic markers. *Bioinformatics* 24:1403–1405.