Supporting Information

Almanza et al. 10.1073/pnas.1311145110

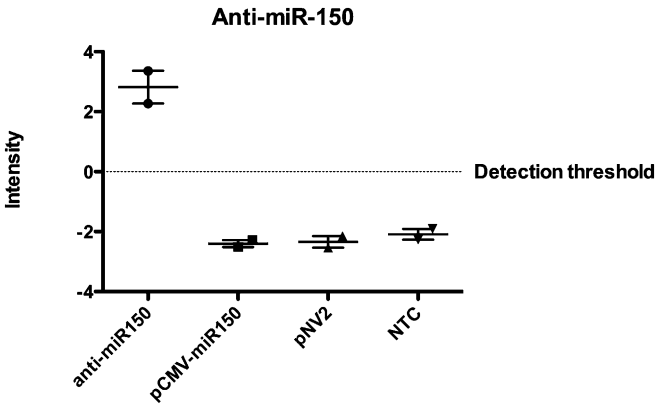

Fig. S1. Anti-microRNA-150 (anti-miR-150) is not amplified from plasmid pCMV-MIR^{a150}. Quantitative RT-PCR was performed as described in *Materials and Methods* using anti-miR-150 primers (ABI). Anti-miR-150 amplification is expressed as the ratio between the fluorescence intensity of the reporter dye (FAM) and that of the passive reference dye (ROX) used for normalization, according to the manufacturer's guidelines. Values above the detection threshold indicate amplification of the target sequence, whereas values below the detection threshold imply lack of amplification of the target sequence. The test reflects 30 standard cycles, including a denaturing step at 95 °C and an annealing/extension step at 60 °C. pNV2 = a ~15 Kb plasmid coding for a full-length chimeric Ig heavy chain gene modified in the CDR2 by insertion of the sequence (NANP-**NV**DP-NANP). From Xiong S, et al. (1997) Engineering vaccines with heterologous B and T-cell epitopes using Ig genes. *Nat Biotech* 15:882–886. NTC, nontemplate control.

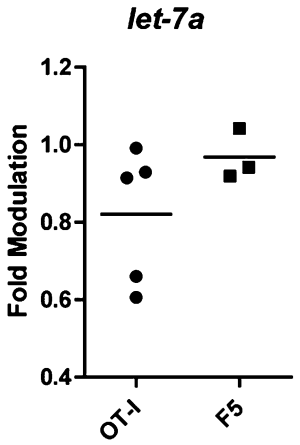

Fig. S2. Detection of endogenous let-7a in CD8 T lymphocytes from in vitro cross-priming cultures. Intracellular microRNAs were isolated using the mirVANA PARIS Kit (Life Technologies) as described in *Materials and Methods* and amplified using let-7a (002478) and snoRNA202 (001232) specific RT primers (ABI). Results are expressed as fold modulation (RQ) and refer to individual experimental data points run in duplicate. Unpaired *t* test, $P = 0.218$.

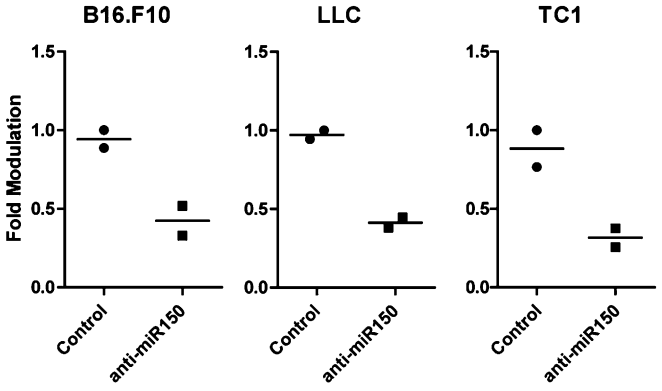

Fig. S3. Down-regulation of endogenous miR-150 in murine tumor cells treated with the supernatant of J558L cells transfected with pCMV-MIR^{a150}. Murine melanoma (B10.F10), lung (LLC), and prostate (TC1) tumor cells were cultured in a 24-well plate for 48 h in RPMI medium containing 1 mL supernatant of J558L cells transfected with pCMV-MIR^{a150} or the supernatant of untransfected J558L cells as a control. Results represent two experiments.

Table S1. Quantification of extracellular vesicle translocation efficiency to cross-primed CD8 T cells

Parameter	No cross-priming + EVs	Cross-priming + EVs
EV-positive cells, % ($n = 75$)	0	47%
EVs per cell	0	50

OT-I CD8 T cells were cross-primed in the presence of PKH67-labeled extracellular vesicles (EVs). The extent of successful EV translocation was quantified by enumerating the percentage of DAPI-positive cells containing punctate green fluorescent foci and the number of foci per positive cell.